

Effect of Brain Angiotensin II Receptor Antagonists and Antisense Oligonucleotide on Drinking and Renal Renin in Rats

Hyeon-kyeong Cho, Eun-kyoung Yang, Hee-suk Han, Won-jung Lee, and M. Ian Phillips¹

Department of Physiology, School of Medicine, Kyungpook National University, Taegu 700–422, Korea; ¹Department of Physiology, College of Medicine, Box 100274, University of Florida, Gainesville, FL 32610, USA

The physiological roles of brain angiotensin II in mediating water deprivation-induced drinking and in regulating renal renin release were assessed in male Sprague-Dawley rats. Specific AT₁ receptor antagonists, losartan and SK 1080, and antisense oligonucleotide (AS-ODN) directed to AT₁ receptor mRNA were intracerebroventricularly (i.c.v.) administered in conscious unrestrained rats. When water was given 20 min after i.c.v. injection of AT₁ receptor antagonists in 48-h water-deprived rats, losartan and SK 1080 produced approximately 20% and 50% decrease in 1-h water intake, respectively. In contrast, i.c.v. treatment of the AS-ODN to AT₁ receptor mRNA for 24-h did not alter 1-h water intake in 24-h water-deprived rats, but prevented the increase in overnight water intake after 24-h water-deprivation. Six-day i.c.v. treatment of AS-ODN did not alter either the basal plasma renin concentration or renal cortical levels of renin and renin mRNA. The present results suggest that endogenous brain Ang II plays an important role in thirst and water intake through AT₁ receptors, but further studies are required to elucidate its regulatory role in renal renin synthesis.

Key Words: Intracerebroventricular, Losartan, SK 1080, Water-deprivation, Renin mRNA

INTRODUCTION

The discrete existence of the brain renin-angiotensin system (RAS) and its role in central cardiovascular and fluid control have been demonstrated (Phillips et al, 1993). Two major receptor subtypes for angiotensin II (Ang II), AT₁ and AT₂, have been identified (Bumpus et al, 1991; Toney & Porter, 1993), and AT₁ receptors are mainly found in the brain areas known to be involved in body fluid and blood pressure homeostasis (Steckelings et al, 1992). The intracerebroventricular (i.c.v.) administration of Ang II in rats elicits a series of responses, such as induction of drinking and salt appetite, blood pressure increase, secretion of vasopressin and adrenocorticotrophic hormone, and inhibition of renin secretion (Hoffman et al, 1977; Reid, 1984; Phillips, 1987; Ahn

et al, 1992).

With the development of competitive specific Ang II receptor antagonists, it has been possible to evaluate the physiological role of endogenous Ang II in the brain. Recently we have shown that acute central administration of losartan, a specific AT₁ receptor antagonist, significantly impaired cardiovascular and vasopressin responses to hemorrhage in conscious rats (Lee et al, 1995) and drinking in 48-h water-deprived rats (Lee et al, 1996). These results suggest that the brain RAS plays a significant role in regulation of blood pressure during hemorrhage and drinking in water deprivation.

Synthetic antisense oligodeoxynucleotides (AS-ODN) have been used successfully to inhibit genetic expression by sequence-specific hybridization of mRNA and synthesis of a particular protein in a number of biological systems. (Crooke, 1993; Phillips & Gyurko 1995) Blocking specific gene expression by AS-ODN inhibits action of specific protein without multiple, nonspecific side effects exerted by most pharmacological drugs. Phosphodiester AS-ODN directed to

Corresponding to: Won-jung Lee, Department of Physiology, School of Medicine, Kyungpook National University, Taegu 700-422, Korea. (Tel) 82-53-420-6926, (Fax) 82-53-424-3349, (E-mail) wjleek@knu.ac.kr

AT₁ receptor or angiotensinogen mRNA significantly reduced blood pressure for several days in hypertensive animals with a single injection into the brain (Gyurko et al, 1993; Phillips et al, 1994; Wielbo et al, 1995). In contrast to the prolonged effect of AS-ODN, the hypotensive effect of i.c.v. losartan in spontaneously hypertensive rats lasted only for several hours. AS-ODN to AT₁ receptor mRNA inhibited drinking response to i.c.v. Ang II (Meng et al, 1994). However, effects of i.c.v. AS-ODN on physiological thirst, such as water deprivation-induced drinking have not been tested.

There is a possibility that central Ang II may be involved in renal renin release. The i.c.v. Ang II decreased plasma renin activity (Eriksson & Fyhrquist, 1976; Malayan et al, 1979; Ahn et al, 1992; Weekley, 1992; McKinley et al, 1994), whereas i.c.v. infusion of losartan with a high dose increased plasma renin activity in sodium-depleted sheep (McKinley et al, 1994). We observed that chronic i.c.v. administration of losartan increased the basal plasma renin concentration 3-fold (Yang et al, 1996), although acute i.c.v. losartan had no effect on the plasma renin concentration (Lee et al, 1995) as reported by others (Baum et al, 1983; Berecek et al 1991; DePasquale et al, 1992). Different responses to acute and chronic i.c.v. losartan might be because longer period of time is required for more losartan to get to critical brain target regions inaccessible to acute losartan.

We therefore attempted in the present study to compare the effects of blockade of brain Ang II-AT₁ receptors with short acting receptor antagonists and long acting AS-ODN on water deprivation-induced drinking and renal renin release. We injected the antagonists and AS-ODN directly to the brain ventricles of conscious unrestrained rats.

METHODS

Animals and surgical procedures

Experiments were performed on conscious, unrestrained male Sprague-Dawley (SD) rats (250~300 g). Each animal was anesthetized with sodium pentobarbital (40 mg/kg, i.p.) and a stainless steel cannula (22 gauge) was stereotaxically implanted into the left lateral cerebral ventricle following the procedure previously reported (Lee et al, 1995). The stereotaxic coordinates were 1.5 mm lateral, 0.8 mm posterior to

the bregma, and 4.0 mm below the skull surface. The animals were allowed to recover for 3 days and were initially screened for correctness of cannula location by an Ang II test. The average water intake after i.c.v. injection of 10 ng Ang II was 6.8 ± 0.5 ml/15 min (n=25). Any rat that drank <4 ml/15 min was not used in the study.

Effect of i.c.v. AT₁ receptor antagonists on drinking in water-deprived rats

Before the experiment, water bottles were removed from the cages for 2 days. In the morning of the experiment, 48-h water-deprived rats remained unrestrained in individual home cages. The rat was injected i.c.v. with either an AT₁ receptor antagonist (losartan potassium, 50 μ g=110 nmol or SK-1080, 10 μ g=20 nmol) or artificial cerebrospinal fluid (aCSF) as the vehicle. SK-1080 is a new non-peptide AT₁ receptor antagonist synthesized in the Korean Research Institute of Chemical Technology (Lee et al, 1999). Water was given 20 min after the injection and water intake was measured for 60 min.

Effect of i.c.v. AS-ODN on drinking in water-deprived rats

Basal overnight water intake (17 : 00~9 : 00) of rats with i.c.v. cannula was measured in each metabolic cage. Next day at 17 : 00, a 50 μ g dose of AS- or scrambled (SC)-ODN or 4 μ l aCSF was administered i.c.v. and water bottles were removed for 24 h. Another overnight water intake was measured in 24-h water-deprived rats.

AS- and SC-ODN were synthesized 15-mers to base +63 to +77 of AT₁ receptor mRNA (Murphy et al, 1991). The ODN sequences were modified by backbone phosphorothioation and were synthesized in the DNA Synthesis Laboratory, University of Florida. The ODN sequences were as follows: AS: 5'-TAAC-TGTGCCTGCAA-3', SC: 5'-AATTGGTGTGTTTC-GTTC-3'.

Effect of i.c.v. AS-ODN on renal levels of renin mRNA and protein

Rats were given two i.c.v. injections of 4 μ l aCSF, or 50 μ g/4 μ l AS- or SC-ODN at 48 h intervals. Two days after the second injection, rats were decapitated, and the trunk blood samples were collected

for measurement of plasma renin concentration. The kidney was removed, renal cortex was isolated and frozen in liquid nitrogen, and then kept in a deep freezer (-70°C) until total RNA isolation.

Plasma and renal cortical renin levels were measured with radioimmunoassay (Lee et al, 1995) and renal renin mRNA levels were measured with Northern blot with methods previously reported (Jo et al, 1996). Briefly, total RNA was extracted from renal cortex according to the method of Chomczynski & Sacchi (1987). For Northern blot analysis, 20 μg total RNA was subjected to electrophoresis on a 1% agarose-formaldehyde denaturing gel and transferred to a nitrocellulose membrane (Amersham). The filter was baked, prehybridized, and hybridized to full-length cDNA for rat renin (provided by Dr. Kevin R. Lynch, University of Virginia) and 18 S probe, both labeled with ^{32}P . The filter then was washed and exposed to x-ray film.

Statistical analysis

Results are expressed as Mean \pm SEM. Statistical analysis was performed by unpaired or paired *t*-test. Differences were considered statistically significant at a value of $P < 0.05$.

RESULTS

Experiment 1

The effects of i.c.v. administration of AT₁ receptor antagonists on water intake in 48-h water-deprived SD rats are shown in Fig 1. Water intake of the control rats injected with aCSF was 16.0 ± 0.67 ml/h. With an AT₁ receptor antagonist, losartan, there was approx. 20% inhibition of drinking (12.6 ± 1.06 ml/h). However, another AT₁ receptor antagonist, SK-1080 with one-fifth the molar dose of losartan, produced a significantly greater inhibition in drinking (7.9 ± 0.48 ml/h) than losartan.

Experiment 2

The effects of i.c.v. administration of AS-ODN on AT₁ receptor mRNA and resultant effects on water intake in 24-h water-deprived rats are shown in Fig 2. The water deprivation increased the overnight water intake of the control rats injected with aCSF

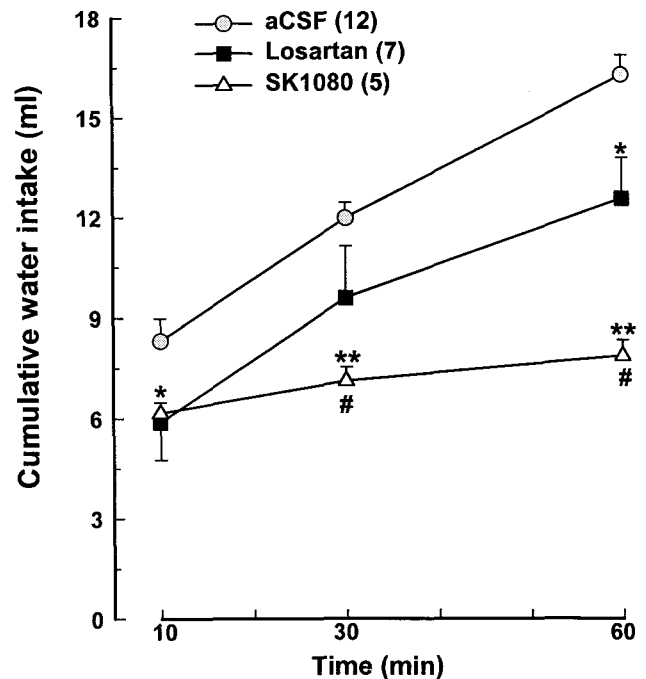


Fig. 1. Effect of i.c.v. administration of AT₁ receptor antagonists (50 μg losartan and 10 μg SK-1080), or artificial cerebrospinal fluid (aCSF) on water intake of rats deprived of water for 48 h. Numbers in parentheses represent numbers of rats. * $P < 0.05$, ** $P < 0.01$, vs. controls given aCSF, # $P < 0.05$, SK-1080 vs. losartan.

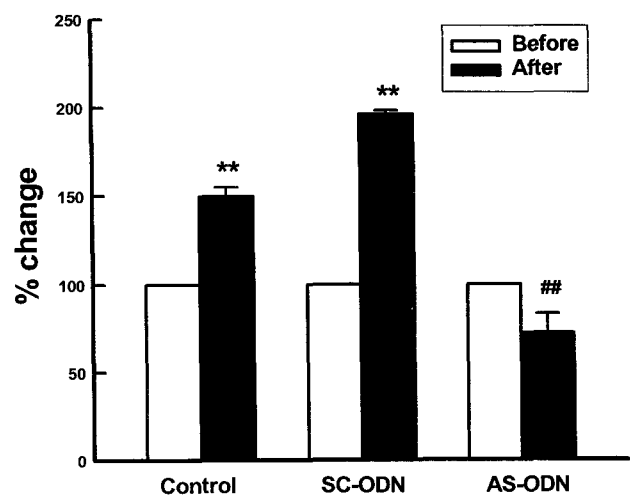


Fig. 2. Effect of i.c.v. administration of AS-ODN for AT₁ receptor mRNA on overnight water intake for 16 h before and after 24 h water deprivation ($n=5$ in each group). ** $P < 0.01$, before vs. after, ## $P < 0.01$, vs. controls given aCSF.

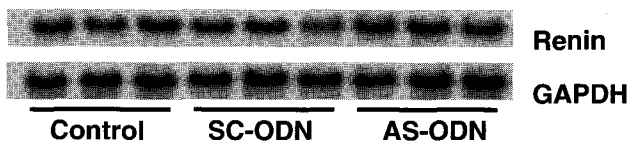


Fig. 3. Representative autoradiogram of Northern blot for renin and GAPDH mRNAs in the kidney from the rats i.c.v. injected twice at 48 h interval with AS-ODN for AT₁ receptor mRNA (n=6 in each group).

or SC-ODN from 37.8 ± 0.57 or 17.0 ± 1.62 ml/16 h to 56.7 ± 1.60 or 32.6 ± 1.70 ml/16 h after the water deprivation (% increase: 49.7 ± 4.03 or 95.8 ± 8.50), respectively. However, with AS-ODN, overnight water intake was even decreased from 28.2 ± 3.93 to 22.5 ± 2.9 ml/16 h after 24-h water deprivation (% decrease: 22.9 ± 9.46).

Experiment 3

The effects of i.c.v. AS-ODN on the plasma and renal renin levels, and renal renin mRNA are shown in Table 1 and Fig 3, respectively. Four-day i.c.v. treatment of AS-ODN altered neither the plasma renin concentration nor the levels of cortical renin and renin mRNA.

DISCUSSION

The present study demonstrates that blockade of the brain AT₁ receptor with antagonists and AS-ODN significantly inhibited the drinking response in water-deprived rats. However, the efficacy of AT₁ receptor antagonist and AS-ODN was different. Centrally administered AT₁ receptor antagonist produced more than 50% inhibition of 1-h water intake in 24-h water-deprived rats when water was given 20 min after the i.c.v. injection. In contrast, i.c.v. treatment of AS-ODN targeted to the AT₁ receptor mRNA for 24 h did not alter 1-h water intake after 24-h water deprivation (data not shown), but markedly decreased overnight water intake after 24-h water deprivation.

Reasons for different responses to AT₁ receptor antagonist and AS-ODN in the present study are not clear, but may be related to differences in distribution properties of the drugs. Inhibitory effects of AT₁ receptor antagonists are usually observed within 10 and 20 min after i.c.v. injection (Lee et al, 1995 & 1996).

Table 1. Basal plasma renin concentration (PRC) and renal renin content (RRC) in rats i.c.v. injected twice at 48 h interval with AS-ODN for AT₁ receptor mRNA

	aCSF	SC-ODN	AS-ODN
PRC, ng/ml/h	5.9 ± 0.6	8.8 ± 1.5	6.3 ± 0.99
RRC, $\mu\text{g}/\text{mg protein/h}$	9.1 ± 0.9	11.6 ± 1.3	8.9 ± 0.56

Values are mean \pm SE of 6 rats in each group.

However, the route of i.c.v. injection of AS-ODN does not seem to be optimal to reach all target cells involved in the responses. AS-ODN is known to be avidly taken up by cells and are concentrated in cells close to the injection site (Ogawa et al, 1995; Phillips & Gyurko, 1995). The AT₁ AS-ODN reduced AT₁ receptor levels in the hypothalamus, which is most proximal to the site of injection, whereas it did not significantly reduce that of the brainstem, which is rich in AT₁ receptors but distal to the injection site. These results suggest that AT₁ receptors in the hypothalamus area would be exposed to the highest concentration of the oligomers, resulting in reduced delivery of the oligomers to more distal site such as the brainstem. Nonetheless, significant reduction of overnight water intake in water deprived rats treated with the AS-ODN in the present study supports the previous suggestion of the necessary involvement of AT₁ receptors in the ventricular regions for the water deprivation-induced dipsogenic action.

We observed suppression of plasma renin concentration after i.c.v. administration of Ang II (Ahn et al, 1992), as reported by other investigators in various species (Eriksson & Fyhrquist 1976; Malayan et al, 1979; Iwata et al, 1985; McKinley et al, 1994). Suppression of renal renin secretion rate in response to centrally administered Ang II was also observed in renal perfusion models of both the constant flow and constant pressure (Weekley, 1992). This result may suggest that changes in intrarenal hemodynamics do not mediate the response. However, in the medial basal forebrain lesioned rats (Weekley, 1992) and hypophysectomized dogs (Malayan et al, 1979), renal renin secretion was not decreased following i.c.v. injection of Ang II. Central administration of Ang II slightly decreased renal nerve activity (Unger et al, 1985), and renal denervation attenuated the central Ang II induced suppression of plasma renin activity

(Iwata et al, 1985). Taken together, these results may indicate an involvement of Ang II in the brain areas in the efferent path of the renal renin response.

Although large doses of Ang II administered directly into the brain can decrease the renin release, the physiological role of endogenous brain Ang II in regulation of renal renin release has not been clearly demonstrated. In sodium-depleted sheep, i.c.v. infusion of losartan with a high dose increased the plasma renin activity (McKinley et al, 1994). We observed that the basal plasma renin concentration increased 3-fold after chronic blockade of central AT₁ receptors with losartan (Yang et al, 1996), but was not altered after acute i.c.v. losartan (Lee et al, 1995). Different responses of the basal plasma renin concentration to acute and chronic i.c.v. losartan might be ascribed to the requirement of longer period of time for more losartan to get to critical brain target regions which is not easily accessible to acute losartan. To test this possibility in the present study, we injected the long acting phosphorothioated AS-ODN targeted to the AT₁ receptor mRNA into the brain twice at 48 h intervals for more effective blockade of the synthesis of the AT₁ receptors. After 4-day i.c.v. treatment of AS-ODN to AT₁ receptor, however, we observed no alteration in the basal plasma renin concentration, and renal levels of renin mRNA and protein. Chronic central blockade of Ang II with the AS-ODN failed to increase the basal plasma renin concentration unlike the chronic effect of losartan. On the other hand, a single i.c.v. injection of the same AS-ODN to the AT₁ receptor mRNA used in the present study produced a marked decrease in blood pressure for longer than a week in spontaneously hypertensive rats (Gyurko et al, 1993). Reasons of the lack of the plasma renin concentration response in the present study are not clear, but may be in part explained by that even repeated i.c.v. injections was not enough for the AS-ODN to effectively block the synthesis of the AT₁ receptors in critical brain areas. Further studies are required to clarify whether endogenous brain Ang II plays a significant physiological role in regulation of renal renin release.

ACKNOWLEDGEMENT

This study is supported by Korean Ministry of Education 1997 Research Fund (97-021-F00022).

REFERENCES

- Ahn DK, Oh ST, Yang EK, Park JS, Lee WJ. Effects of intracerebroventricular angiotensin II on the cardiovascular and endocrine systems in conscious normotensive and hypertensive rats. *J Kor Soc Endocrinol* 7: 364–372, 1992 (in Korean)
- Baum T, Becker FT, Sybertz EJ. Attenuation of pressor responses to intracerebroventricular angiotensin I by angiotensin converting enzyme inhibitors and their effects on systemic blood pressure in conscious rats. *Life Sci* 32: 1297–1303, 1983
- Berecek KH, Robertson JD, Thorstad MH. Central administration of a specific angiotensin II receptor antagonist on baroreflex function in spontaneously hypertensive rats. *J Hypertens* 9: 365–371, 1991
- Bumpus FM, Catt KJ, Chiu AT, DeGasparo M, Goodfried T, Husain A, Peach MJ, Taylor Jr. DG and Timmermans PBMWM. Nomenclature of angiotensin receptors. *Hypertension* 17: 720–721, 1991
- Chomczynski P, Sacchi N. Single-step method of RNA isolation by acid guanidium thiocyanate-phenol-chloroform extraction. *Anal Biochem* 162: 156–159, 1987
- Crooke ST. Progress toward oligonucleotide therapeutics: pharmacodynamic properties. *FASEB J* 7: 533–539, 1993
- DePasquale MJ, Fossa AA, Holt WF, Mangiapane ML. Central Dup 753 does not lower blood pressure in spontaneously hypertensive rats. *Hypertension* 19: 668–671, 1992
- Eriksson L, Fyhrquist F. Plasma renin activity following central infusion of angiotensin II and altered CSF sodium concentration in the conscious goat. *Acta Physiol Scand* 98: 209–216, 1976
- Gyurko R, Wielbo D, Phillips MI. Antisense inhibition of AT₁ receptor mRNA and angiotensinogen mRNA in the brain of spontaneously hypertensive rats reduces hypertension of neurogenic origin. *Regul Pept* 49: 167–174, 1993
- Hoffman WE, Phillips MI, Schmid PG, Falcon J, Weet JF. Antidiuretic hormone release and the pressor response to central angiotensin II and cholinergic stimulation. *Neuropharmacology* 16: 463–472, 1977
- Iwata T, Hiwada K, Kokubu T. Effects of renal denervation on renin release induced by intracerebroventricular administration of biological active peptides in conscious rats. *Neuropeptides* 6: 437–443, 1985
- Jo H, Yang EK, Lee WJ, Park KY, Kim HJ, Park JS. Gene expression of central and peripheral renin-angiotensin system components upon dietary sodium intake in rats. *Regul Pept* 67: 115–121, 1996
- Lee BH, Seo HW, Kwon KJ, Yoo SE, Shin HS. In vivo pharmacological profile of SK-1080, an orally active nonpeptide AT₁ receptor antagonist. *J Cardiovasc*

- Pharmacol* 33: 375–382, 1999
- Lee WJ, Yang EK, Ahn DK, Park YY, Park JS, Kim HJ. Central ANG II-receptor antagonists impair cardiovascular and vasopressin response to hemorrhage in rats. *Am J Physiol* 268: R1500–R1506, 1995
- Lee WJ, Kim KS, Yang EK, Lee JH, Lee EJ, Park JS, Kim HJ. Effect of brain angiotensin AT1, AT2, and cholinergic receptor antagonism on drinking in water deprived rats. *Regul Pept* 66: 41–46, 1996
- Malayan SA, Keil LC, Ramsay DJ, Reid IA. Mechanism of suppression of plasma renin activity by central administered angiotensin II. *Endocrinology* 104: 672–675, 1979
- McKinley M, Evered M, Mathai M, Coghlan JP. Effects of central losartan on plasma renin and centrally mediated natriuresis. *Kidney Int* 46: 1479–1482, 1994
- Meng H, Wielbo D, Gyurko R, Phillips MI. Antisense oligonucleotide to AT1 receptor mRNA inhibits central angiotensin induced thirst and vasopressin. *Regul Pept* 54: 543–551, 1994
- Murphy TJ, Alexander RW, Griendling KK, Riunge MS, Bernstein KE. Isolation of cDNA encoding the vascular type-1 angiotensin II receptor. *Nature* 351: 233–236, 1991
- Ogawa S, Brown HE, Okano HJ, Pfaff DW. Cellular uptake of intracerebrally administered oligodeoxynucleotides in mouse brain. *Regul Pept* 59: 143–149, 1995
- Phillips MI. Functions of angiotensin in the central nervous system. *Annu Rev Physiol* 49: 413–435, 1987
- Phillips MI, Speakman EA, Kimura B. Levels of angiotensin and molecular biology of the tissue renin angiotensin system. *Regul Pept* 43: 1–20, 1993
- Phillips MI, Wielbo D, Gyurko R. Antisense inhibition of hypertension: a new strategy for renin-angiotensin candidate genes. *Kidney Int* 46: 1554–1556, 1994
- Phillips MI, Gyurko R. In vivo applications of antisense oligonucleotides for peptide research. *Regul Pept* 59: 131–141, 1995
- Reid IA. Actions of angiotensin II on the brain: mechanisms and physiological role. *Am J Physiol* 246: F533–1984
- Toney GM, Porter JP. Functional role of brain AT1 and AT2 receptors in the central angiotensin II pressor response. *Brain Res* 603: 57–63, 1993
- Unger T, Schneider B, Becker H, Petty M, Demmert G, Ganten D, Lang R. Differential effects of central angiotensin-II and substance P on sympathetic nerve activity in conscious rats. *Circ Res* 56: 563–575, 1985
- Weekley LB. Renal renin secretion and norepinephrine secretion rate in response to centrally administered angiotensin II: Role of the medial basal forebrain. *Clin Exp Hypertension A14*: 923–946, 1992
- Wielbo D, Sernia C, Gyurko R, Phillips MI. Antisense inhibition of hypertension in the spontaneously hypertensive rat. *Hypertension* 25: 314–319, 1995
-