C-2 Modified Taxol Analogs with Improved Aqueous Solubility

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Paclitaxel¹ (Taxol), **1** is a novel anticancer agent isolated from the bark of the western yew² (*Taxus brevifolia*) that has been approved for treatment of advanced ovarian and breast cancers.



One of the problems in the pharmaceutical development of taxol is its extremely low aqueous solubility.³ An emulsion formulation has been developed, but this is not ideal because it causes hypersensitivity in some patients.⁴ Therefore, designing and synthesizing an improved aqueous solubility taxol analogs is highly desirable.

According to previous structure-activity relationship works⁵ of taxol, certain substituted groups at C-2 have improved activity. It encourages us to investigate new heteroaromatic substituted analogs at C-2 which are also thought to have improved water solubility.

Our synthesis was started with 10-deacetylbaccatin (III) **2** which was selectively protected as its tricthylsilyl (TES) ether. The C-10 hydroxyl group was then protected as the triethylsilyl (TES) ether through the use of n-butyllithium and triethylsilyl chloride. The C-13 hydroxyl group was subsequently protected as the trimethylsilyl (TMS) ether. This fully protected 10-deacetylbaccatin (III) **3** underwent selective reduction with Red-AI to give the diol **4** (Scheme1).⁶ Attachment of new C-2 analogs has always been problematic⁷ because of easy formation of THF ring between C-2 and C-20 in either basic or acidic media.

Thus, lithiation of diol **4** at low temperature (-78 °C) followed by addition of 2-furoyl chloride, 2-thiophenecarbonyl chloride, nicotinoyl chloride, and isonicotinoyl chloride afforded new C-2 heteroaromatic esters **5**, **6**, **7**, and **8**. Partial desilylation afforded 7,10-TES baccatin derivatives **9**, **10**, **11**, and **12** which coupled⁸ with optically active β -lactam **13** to give new C-2 ester analogs. Finally, fluoride assisted removal of the silyl ethers resulted in new heteroaromatic substituted taxol analogs **14**, **15**, **16**, and **17**° (Scheme 1). As far as we know, these four analogs are the first example of heteroaromatic substituted analogs at C-2.

These new C-2 modified analogs were evaluated against human breast (MCF-7), human ovarian (SK-OV-3), and human lung (Λ 549) cell lines (Table 1). For comparison,



Scheme 1. Reagents and Conditions: (i) TESCI. Pyr., rt. 97%; (ii) n-BuLi, THF, -78 °C, then TESCI. 89%; (iii) TMSCI, imidazole, DMAP, DMF, rt., 97%; (iv) Red-AI, THF, 0 °C, 97%; (v) LHMDS, THF, -78 °C, then, 2-furoyl chloride for 5 (76%), 2-thiophenecarbonyl chloride for 6 (72%), nicotinoyl chloride for 7 (52.5%), and isonicotinoyl chloride for 8 (48.4%); (vi) Pyr., 48% HF, CH₃CN, 9 (76.6%), 10 (78.6%), 11 (72.4%), and 12 (71.3%); (vii) LHMDS, THF, -45 °C, 13, then, Pyr., 48% HF, CH₃CN, 14 (71.9%), 15 (72.9%), 16 (49.1%), and 17 (45.7%).

Table 1. In Vitro Cytotoxicities of C-2 Modified Taxols

Cytotoxicity ED ₅₀ (µg/mL)"			
Compd	MCF-7 ^b	SK-OV-3 ^c	$\Lambda 549^d$
1	0.004	0,002	0.004
14	> 10.0	> 10.0	> 10.0
15	> 0.1	≥ 0.1	> 0.1
16	> 0.1	\geq 0.1	> 0.1
17	> 0.1	> 0.1	> 0.1

 $^{\circ}$ ED₃₀ is the concentration of compound that cause a 50% reduction in absorbance at 540 nm relative to untreated cells using SRB assay. hest MCF-7, SK-OV-3, A549 are human breast, ovarian, lung cancer cell line, respectively.

taxol was also evaluated. We found that all new analogs had negligible activity in all assay. In Table 2 the solubility data

Compd	Aqueous solubility"	
1	0.0008	
14	0.1664	
15	0.0701	
16	1647	
17	0.1887	

Table 2. Solubility Data for Taxol Analogs

" In mg mL, determined by reverse phase HPLC.

for C-2 modified analogs is presented. All analogs shows excellent water solubility compared to taxol.

In conclusion, the introduction in the C-2 position of heteroaromatic rings causes a significant increase of their water solubility. These new analogs offer the hope of acceptable solubility in the pharmaceutical development.¹⁰ New taxol analogs with improved water solubility and their evaluation study are under investigation in our laboratory.

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- 9. 14: mp. 156-157 °C; ¹Η NMR (CDCl₃, 300 MHz) δ 1.03 (s, 3H, Me16), 1.14 (s, 3H, Me17), 1.68 (s, 3H, Me19), 1.69 (s, 3H, Me18), 1.79 (m, 1H, H6 β), 2.18 (m, 2H, H14), 2.31 (s, 3H, 4Ac). 2.48 (m, 1H, H6 α). 3.80 (d, 1H, J = 7.0Hz, H3), 4.17 (m, 1H, H7), 4.19 (d, 1H, J = 8.3 Hz, H20 β). 4.35 (d, 1H, J = 8.3 Hz, H20 β), 4.75 (d, 1H, J = 2.2 Hz, H2[']), 4.88 (d, 1H, J = 9.5 Hz, H5), 5.15 (s, 1H, H10), 5.51 (d, 1H, J = 7.0 Hz, H2), 5.73 (dd, 1H, J = 2.4, 9.0 Hz, H3[']), 6.14 (dd, 1H, J = 8.5, 8.8 Hz, H13), 6.54 (dd, 1H, J = 1.7, 3.4 Hz, furoyl), 7.21 (d, HI, J = 9.0 Hz, NH), 7.38 (m, 111, furoyl), 7.38 (m, 811, 2'-NBz and 3'-Ph), 7.62 (d, 111, J = 1.7 Hz, furovl), 7.71 (m, 211, 2]-NBz and 3]-Ph). 15: mp. 161-163 °C: ¹Η NMR (CDCl₃, 300 MHz) δ 1.06 (s, 3H, Me16), 1.16 (s, 3H, Me17), 1.71 (s, 3H, Me19), 1.71 (s. 311, Me18), 1.81 (m. 111, H6B), 2.24 (m. 211, H14), 2.33 (s, 3H, 4Ac), 2.53 (m, 1H, H6α), 3.82 (d, 1H, J = 7.1 Hz, H3), 4.23 (dd, 1H, J = 7.1, 11.0 Hz, H7), 4.24 (d, 1H, J = 8.4 Hz, H20 β), 4.40 (d, 1H, J = 8.3 Hz, H20 α), 4.75 (d, 1H, J = 2.4 Hz, H2'), 4.91 (d, 1H, J = 8.6 Hz, H5), 5.15 (s, 1H, H10), 5.53 (d, 1H, J = 7.1 Hz, H2), 5.73 (dd, 1H, J = 2.4, 9.0 Hz, H3²), 6.13 (dd, 1H, J = 7.6, 8.3 Hz, 1113), 7.13 (dd, 111, J = 3.7, 4.9 Hz, thiophene), 7.15 (d, 111, J = 9.1 Hz, NH), 7.41 (m, 8H, 2'-NBz and 3'-Ph), 7.63 (dd, 111, J = 1.2, 4.9 Hz, thiophene), 7.74 (m, 2H, 2'-NBz and 3'-Ph), 7.88 (dd, 111, J = 1.2, 3.7 Hz, thiophene). **16**: mp. 156-157 °C; ¹H NMR (CDCl₃, 300 MHz) δ 1.10 (s, 311, Me16), 1.19 (s, 311, Me17), 1.74 (s, 311, Me19), 1.77 (s. 3H, Me18), 1.85 (m, 1H, H6\beta), 2.30 (m. 2H, H14), 2.36 (s. 3H, 4Ac), 2.56 (m. 1H, H6a), 3.89 (d. 1H, J = 7.3 Hz, H3), 4.19 (d. 1H, J = 8.2 Hz, H20 β), 4.21 (m, 1H, 2OH). 4.26 (d. 1H, J = 8.2 Hz, H20 α). 4.27 (dd. 1H. J = 6.8. 13.5 Hz, H7), 4.75 (d. 1H, J = 2.8 Hz, H2c), 4.92 (dd, 1H, J = 1.1, 9.3 Hz, H5), 5.17 (s, 1H, H10), 5.67 (d, 111, J = 7.3 Hz, 112), 5.74 (dd, 111, J = 2.2, 8.9 Hz, H3'). 6.16 (t, 111, J = 8.3 Hz, 1113), 7.04 (d, 111, J = 8.9 Hz. NH), 7.42 (m, 1H, nicotinovl), 7.57 (m, 10H, 2'-NBz and 3'-Ph), 8.38 (d, 111, J = 7.9 Hz, nicotinovl), 8.81 (s, 111, nicotinovl), 9.30 (s. 111, nicotinovl), 17; mp 171-173 °C; ¹H NMR (CDCl₃, 300 MHz) δ L07 (s, 3H, Me16), 1.19 (s, 3H, Me17), 1.73 (s, 3H, Me19). 1.77 (s, 3H, Me18), 1.86 (m, 1H, H6 β), 2.28 (m, 2H, H14), 2.38 (s, 3H, 4Ac), 2.54 $(m, 1H, H6\alpha), 3.88 (d, 1H, J = 7.3 Hz, H3), 4.15 (d, 1H, J)$ = 8.2 Hz, H20 β). 4.17 (m. 1H, 2'OH), 4.22 (d, 1H, J = 8.2 Hz, H20 α), 4.22 (m. 1H. H7), 4.78 (d, 1H, J = 2.5 Hz, H2'), 4.90 (dd, 1H, J = 1.1, 9.3 Hz, H5), 5.17 (s, 1H, H10), 5.64 (d, 111, J = 7.3 Hz, 112), 5.78 (dd, 111, J = 2.3, 9.3 Hz, H3'), 6.20 (t, HI, J = 8.1 Hz, H13), 7.09 (d. HI, J = 9.1Hz, NH), 7.55 (m, 10H, 2]-NBz and 3]-Ph), 7.90 (d, 1H, J = 1.8 Hz, isonicotinov1), 7.91 (d. 111, J = 1.6 Hz, isonicotinov1), 8.79 (d. 111, J = 1.6 Hz, isonicotinov1), 8.80 (d, 111, J = 1.8 Hz, isonicotinovl).
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