The Effect of Dietary Lipids on CVD Risk Factors in Ovariectomized Rats

Kwon, Ja Ryong · Ahn, Hae Seon · Lee, Sang Sun

Department of Food and Nutrition, Hanyang University, Seoul, Korea

ABSTRACT

This experiment was performed to investigate the effect of different dietary lipids on the risk factors of coronary vascular disease(CVD) in ovariectomized rats. Female rats of Sprague-Dawley strain were divided into sham-operated(sham) and ovariectomized(ovx) groups and then each group was divided into a beef tallow group, a soy bean oil group and a fish oil group. After 16 weeks of feeding on experimental diets, animals were sacrificed and blood, liver, kidney and perirenal fat pad were obtained.

Food intake and weight gain of fish oil group were significantly lower than other dietary lipid groups. Food intake and weight gain tended to be higher in ovx groups than in sham groups. The weight index(g/100g body weight) of liver and kidney was higher in the fish oil group than the other groups and weight index was lower in ovx groups compared to sham groups. The weight of the perirenal fat pad was the highest in the beef tallow group and the lowest in the fish oil group. The fish oil group showed the lowest total cholesterol(TC) and triglyceride (TG) levels in serum. Serum TG levels were lower in all ovx groups than in sham groups, but serum TC levels were not influenced by ovariectomy.

Fatty acid composition of serum reflects the recent dietary intake of fat. Linoleic acid content was the highest in soy bean oil group and eicosapentaenoic acid(EPA) and docosahexaenoic acid(DHA) contents were the highest in fish oil group. Fatty acid composition of adipose tissue, especially EPA and DHA contents in perirenal fat pad, was highest in the fish oil group. Saturated fatty acid(SFA) and monounsaturated fatty acid(MUFA) in serum and adipose tissue did not reflect fatty acid intake.

The activities of glucose-6-phosphate dehydrogenase, a lipogenic enzyme, in the blood of the beef tallow and soybean oil groups showed the tendency to be high and that of the fish oil group to be low in ovx. Carnitine acetyltransferase, a lipolytic enzyme, showed the highest activity in the liver of the fish oil group and was least active in the soy bean oil group. (*Korean J Nutrition* 30(4): 386~393, 1997)

KEY WORDS: dietary lipids · ovariectomy · glucose-6-phosphate dehydrogenase · carnitine acetyltransferase.

Introduction

Recently in Korea the incidence of cardiovascular disease(CVD) has been increasing¹⁾ and the mortality rate from this disease reached 29.9% in 1990²⁾. Al-

Accepted: May 16, 1997

though some studies^{3,4)} reported that menopause was not related to the risk of CVD, many studies^{5,8)} revealed a relationship between them. Premenopausal women have a lower incidence of CVD than men, but this characteristic disappears by natural and artificial menopause⁴⁾. Some of case-control studies of menopause also reported that postmenopausal women have higher cardiovascular or atherosclerotic risk than

^{*}This study was supported by Daewoo Foundation.

premenopausal women⁹¹⁰. The changes in the endocrine system because of menopause cause changes in serum lipid profiles.

Some studies¹¹⁾¹²⁾ reported that ovariectomy causes increases in body weight and body fat accretion. Obesity, which is the most common health problem in middle-aged women after menopause, increases blood volume and cardiac output and then causes heart hypertropy, hypertension, and coronary arterial disease (CAD) directly¹³⁾. It also causes hypercholesterolemia, hypertriglyceridemia, low HDL-cholesterol, and then cardiovascular disease indirectly14). There are a few studies about the effect of dietary lipid composition on fat accumulation. London et al. 15) reported that they observed only weak correlations between the content of saturated and monounsaturated fatty acids in adipose tissue and the dietary lipid intake of these two fatty acids. However, they found strong correlations between polyunsaturated fatty acids and dietary lipid intake. The purpose of this study was to investigate the effect of dietary lipids on the risk factors of CVD in ovariectomized rats.

Materials and Methods

1. Experimental diets

The experimental diets were based on the AIN-76¹⁶⁾ and the amount of lipids was about 15%30% of total

calories) of dietary weight. Table 1 shows the experimental diet composition. Three different kinds of dietary lipids were used ; beef tallow(Oddugi Co.), soybean oil(Dong-Bang oil Co.) and fish oil(Kohap Bio. Co.). To prevent oxidation of diets, α -tocopherol (Sigma Co. St. Louis, MO) was added to all experimental diets, which were stored under N_2 gas in a refrigerator.

2. Experimental animals and operation

Female Sprague-Dawley rats had an adaptation period with chow and their average weight was $227\pm3g$. Animals were divided into 6 groups with 10 animals in each group. Ovariectomies were performed under ethyl ether anesthesia through under-back incisions with removal of ovaries(ovx) and sham operations consisted of only under-back incisions without removal of ovaries(sham). Animals were housed individually in stainless steel hanging cages in a temperature($22\pm2\%$) and humidity(40-60%) regulated room. The room was lighted for 12 hours. Experimental diet and tap water were provided ad libitum.

3. Sampling

After sixteen weeks of feeding on experimental diets, animals were sacrificed. Blood samples were obtained through heart puncture and centrifuged at $950 \times g$ for 15 minutes and separated serum was stored at -75%. Liver, kidney, and perirenal fat pad were removed and

Table 1. Composition of experiment	al diets	,
---	----------	---

(g/kg)

Table 1. Composition of expe	_		(8/1/8			
	S	Sham operation			Ovariectomy	
	BT ¹⁾	SB ²⁾	FO ³⁾	BT	SB	FO
. Corn starch	470	470	470	470	470	470
Sucrose	150	150	150	150	150	150
Casein	150	150	150	150	150	150
Methionine	3	3	3	3	3	3
Choline chloride	2	2	2	2	2	. 2
Beef tallow	150	50	50	150	50	50
Soy bean oil		100			100	
Fish oil			100			100
Vitamin mixture49	10	10	10	10	10	10
Mineral mixture ⁵⁾	35	35	35	35	35	35
α-cellulose	30	30	30	30	30	30
α-tocopherol	0.08	0.08	0.08	0.08	0.08	0.08

¹⁾ Beef tallow, 2)Soy bean oil, 3) Fish oil

⁴⁾ Vitamin Mixture(mg/100g): VD₃ 0.582, α-tocopherol-acetate 1200.0, Retinol-acetate 93.2, VK₃ 6.0, Thiamin-HCl 59.0, VB₁₂ 0.2, VC 588.0, Pyridoxine-HCl 29.0, D-biotin 1.0, Folic acid 2.0, Inositol 1176.0, Ca-pantothenate 235.0, Riboflavin 59.0, Nicotinic acid 294.0, Sucrose 96257.017

⁵⁾ Mineral Mixture(g/100g) : $CaCO_3$ 29.29, $CaHPO_4$ · $2H_2O$ 0.43, KH_2PO_4 34.31, NaCl 25.06, $MgSO_4$ · $7H_2O$ 9.98, $Fe(C_6H_5O_7)$ · $6H_2O$ 0.623, $CuSO_4$ · $5H_2O$ 0.156, $MnSO_4$ · H_2O 0.121, $(NH_4)_6Mo_7O_{24}$ · $4H_2O$ 0.0025, Na_2SeO_3 · $5H_2O$ 0.0015, $ZnCl_2$ 0.02, $ZnCl_2$ 0.005

weighed.

4. Analysis

1) Serum lipid concentration

Serum triglyceride, total cholesterol, and HDL-cholesterol were analyzed by enzymatic procedures using kits(Young-Dong Co.) LDL-cholesterol was calculated using Fredelwald¹⁷ formula as follow:

LDL - cholesterol = Total cholesterol - (HDL-cholesterol + triglyceride/5)

2) Serum and adipose tissue fatty acid composition

Serum fatty acid composition was measured using Fletcher's method and Lapage and Roy's method by gas liquid chromatography(GLC). The perirenal fat was homogenized with 10ml hexane: isoprophyl solution(3:2, v/v) and then centrifuged at $2500\times g$ for 10 minutes. After centrifugation, a white lipid layer was removed to a plastic tube and analysed in the same manner as was the serum lipid material.

3) Glucose-6-phosphate dehydrogenase(G-6-PDH) and Carnitine acetyltransferase (CAT) activity

Activity of G-6-PDH, a lipogenic enzyme, was measured in the blood using a modification of Kornberg and Horecker's method²⁰⁾ and Lohr and Waller's method²¹⁾. In this experiment, a kit(Sigma Co. kit No. 345) was used. Enzyme activity was expressed as unit per gram of hemoglobin where one unit is defined that amount of G-6-PDH activity that will convert 1 micromole of substrate per minute.

The activity of CAT, a lipolytic enzyme, was measured spectrophotometrically by following the appearance of CoASH using the general thiol reagent DTNB(Ellmans's reagent)²²⁾. The mitochondria were separated from the liver for measuring CAT activity²³⁾. Protein content was determined by the procedures of Lowry et al²⁴⁾.

5. Statistical analysis

All statistical analysis was performed using the statistical package for social science(SPSS). Data was presented as the means \pm SEM. Statistical evaluations were performed by ANOVA, and followed by Duncan's multiple range test for differences between means. A probability value of α =0.05 was chosen as the

level of statistical significance.

Results and Discussion

1. Food intake and Body weight gain

The food intake, body weight gain, and food efficiency ratio(FER) are shown in Table 2. The food intake of the fish oil group was slightly lower than that of other lipid groups. The reason for the low food intake in the fish oil group was thought that the fish oil diet had a fish-like smell. The food intake tended to be higher in ovx than in sham. The body weight gain of the fish oil group was the lowest and weight gain was higher in ovx than in sham. These results were consistant with a previous study. The food efficiency ratio(FER) of the fish oil group was the lowest, but statistically not significant. FER of the ovariectomized groups was higher than the sham operation groups.

2. Organ weights

The organ weight and weight index, which were expressed as organ weight per 100g body weight to eliminate differences in body weight among the groups, are shown in Table 3. The liver weight and index of the fish oil group was the highest and the liver weights of the three lipid groups in ovx tended to be lower than those in sham. Perirenal fat pads were the smallest in fish oil groups for both ovx and sham animals. Results of the present experiment are consistant

Table 2. Food intake, weight gain, and FER(Food Efficiency Ratio)

	circy radio,		
Group	Food intake (g/d)	Weight gain (g/d)	FER
SBT	$21.73 \pm 0.99^{1)bc2)}$	1.69 ± 0.20^{ab}	$0.08 \pm 0.01^{\mathrm{ab}}$
SSB	$20.58 \pm 0.89^{\text{abc}}$	1.43 ± 0.14^{ab}	$0.07 \pm 0.00^{\text{ab}}$
SFO	18.79 ± 0.64^a	$1.23\!\pm\!0.15^a$	0.06 ± 0.01^a
OBT	$22.55 \pm 0.40^{\circ}$	2.12±0.11 ^b	0.09 ± 0.00^{b}
OSB	21.66 ± 0.66 ^{bc}	2.13 ± 0.24^{b}	$0.01\pm0.01^{\rm b}$
_ OFO	19.81 ± 0.40^{ab}	1.46 ± 0.25 ab	0.07 ± 0.01^{ab}
SF ³⁾	DL	DL, OVX	OVX
DL*OVA ⁴⁾	NS	NS	NS

- 1) Mean ± S.E.
- Values with different letters are significantly different among experimental groups at p<0.05 according to Duncan's multiple range test
- 3) Significant factor(SF) : Statistical significance was calculated at the α =0.05 level by 2-way ANOVA
- Interaction between dietary lipids(DL) and ovariectomy(OVX)

with the results of other studies comparing fish oil, coconut oil, corn oil, or olive oil²⁵. Also, Parrish et al ²⁶. reported that the fish oil group had a lower body fat weight than the lard group. Shimomura²⁷ reported that long-chain fatty acids in fish oil were oxidized easily and not deposited.

3. Serum lipids

Total cholesterol, HDL-cholesterol, LDL-cholesterol, and triglyceride in serum are shown in Table 4. These results were analysed by ANACOVA(Analysis of covariance) model using food intake as a covariance variable to remove the effect of food intake on serum lipid levels.

Total cholesterol in the fish oil group was significantly lower than other groups in both sham and ovx. It is known that saturated fatty acids tend to increase serum lipids while polyunsaturated fatty acids tend to decrease

serum total cholesterol and LDL-cholesterol²⁸⁾²⁹⁾. Polyunsaturated fatty acids are known to decrease total cholesterol by disturbing absorption in the small intestine³⁰⁾, increasing excretion of bile acid³¹⁾, disturbing synthesis of cholesterol in the liver30, and increasing mobilization of cholesterol through LDL-receptors to the tissue³²⁾. Simons et al.³³⁾ reported that the PUFA increase the excretion of cholesterol to bile acid to decrease serum total cholesterol. Especially, n-3 PUFA decreased triglyceride synthesis in the liver and release of VLDL from the liver to the blood. The serum TG level of the fish oil group was the lowest compared to beef tallow and soybean oil groups in ovx and sham. Generally, the PUFA were known to decrease serum TG by lowering lipogenic enzyme activity³⁴⁾ and VLDL synthesis in the liver 35/36). These experimental results were consistant with those of other studies37,389 that groups fed perilla oil or fish oil rich in n-3 PUFA showed low-

Table 3. Liver, kidney, and perirenal fat weight of experimental rats

Group	Liver (g)	Liver index (g/100g B.W.)	Kidney (g)	Kidney index (g/100g B.W.)	Perirenal fat (g)	Perirenal fat index (g/100g B.W.)
SBT	13.12±0.95 ^{1)ab}	$2.79\pm0.08^{ ext{b2}}$	$2.60 \pm 0.08^{ m abc}$	0.56 ± 0.03^{cd}	20.55 ± 0.87^{cd}	4.42±0.14bc
SSB	13.89 ± 1.40^{ab}	2.73 ± 0.13^{ab}	$2.73 \pm 0.18^{\circ}$	$0.57\!\pm\!0.03^{\text{cd}}$	15.49 ± 0.65 ab	3.56 ± 0.11 ab
SFO	$15.61 \pm 1.30^{\circ}$	$3.68\!\pm\!0.16^{\scriptscriptstyle d}$	$2.65\!\pm\!0.15^{\text{bc}}$	$0.63\!\pm\!0.01^{d}$	$12.05\!\pm\!0.99^{\circ}$	$2.89\!\pm\!0.22^{a}$
OBT	$12.83\!\pm\!1.09^{ab}$	$2.52\!\pm\!0.17^{\text{ab}}$	2.34 ± 0.05^{ab}	0.47 ± 0.01 °	23.13 ± 1.39^{d}	$4.59 \pm 0.24^{\circ}$
OSB	$11.86 \pm 0.96^{\circ}$	$2.37 \pm 0.14^{\circ}$	$2.39\!\pm\!0.08^{ab}$	$0.49\!\pm\!0.03^{ab}$	17.33 ± 1.86 ^{bc}	$3.35 \pm 0.27^{\circ}$
OFO	$13.82\!\pm\!0.83^{\text{ab}}$	$3.26 \pm 0.84^{\circ}$	$2.29\!\pm\!0.28^{a}$	$0.55\!\pm\!0.02^{\text{bc}}$	12.36 ± 2.11^a	2.74 ± 0.37^{a}
SF ³	DL, OVX	DL, OVX	DL, OVX	OVX	DL	DL
Interaction ⁴⁾	NS ⁵⁾	NS	NS	NS	NS	NS

¹⁾ Mean ± S.E.

Table 4. Total cholesterol, Triglyceride, HDL-C and LDL-C in serum

Group	TC	TG	HDL-C	LDL-C
SBT	127.13± 4.58 ¹⁾⁶	143.09±27.44 ^{d2)}	54.92± 7.63°	43.59± 5.34
SSB	123.72±14.84 ^b	$120.73 \pm 29.0^{\text{\tiny cd}}$	50.91 ± 5.79^{a}	54.51 ± 13.04
SFO	82.08 ± 5.16^{a}	32.01 ± 4.21^a	44.27 ± 5.52^{a}	43.91 ± 8.77
OBT	145.23±11.29 ⁶	85.06 ± 11.94 ^{bc}	$87.06 \pm 14.87^{\mathrm{b}}$	57.70± 9.66
OSB	132.00 ± 12.28^{b}	57.69 ± 6.20^{ab}	89.96±15.73 ^b	47.90 ± 9.69
OFO	68.61 ± 4.11^{a}	$31.73\pm\ 2.33^{a}$	34.54 ± 4.81^{a}	38.94 ± 10.30
SF ³⁾	DL	DL, OVX	NS	NS
Interaction4)	DL*OVX	DL*OVX	DL*OVX	NS

¹⁾ Mean ± S.E.

²⁾ Values with different letters are significantly different among experimental groups at p<0.05 according to Duncan's multiple range test

³⁾ Significant factor(SF): Statistical significance was calculated at the $\alpha = 0.05$ level by 2-way ANOVA

⁴⁾ Interaction between dietary lipid(DL) and ovariectomy(OVX)

⁵⁾ NS: Not significant

Values with different letters are significantly different among experimental groups at p<0.05 according to Duncan's multiple range test

³⁾ Significant factor(SF): Statistical significance was calculated at the α =0.05 level by 2-way ANACOVA

⁴⁾ Interaction between dietary lipid(DL) and ovariectomy(OVX)

er TG concentration than other lipids. In this experiment, serum HDL-cholesterol was higher in OBT and OSB groups than in other groups. Harris et al. ³⁹ reported that serum HDL-cholesterol was higher in postmenopausal women than premenopausal women. The group fed DHA-rich fish oil showed lower HDL-cholesterol levels than other groups. Serum LDL-cholesterol concentration was not different among the experimental groups.

4. Fatty acid composition of serum and adipose tissue

Serum fatty acid composition is shown in Table 5. The soybean oil group showed the highest content of linoleic acid and the fish oil group showed the highest DHA and EPA concentrations. These results were similar to these of other studies³⁹. The effect of dietary lipids on the fatty acid composition of perirenal fat is given in Table 6. The adipose tissue of the fish oil

Table 5. Fatty acid composition of serum

(Relative	woight	0/ \
rkeiative	weignt	701

Fatty acid	SBT	SSB	SFO	OBT	OSB	OFO	SF ³⁾	Interaction ⁴⁾
C14:0	2.42±0.19 ^{1)abc}	2.11±0.16 ^{ab}	2.82±0.20 ^b	2.91±0.31°	1.96±0.11 ^a	3.26 ± 0.29^{d}	DL	NS
C16:0	17.69 ± 0.71^{bc}	18.02 ± 1.03^{ab}	$23.97 \pm 1.13^{\circ}$	20.03 ± 1.13^{b}	17.16 ± 0.50^a	$26.44 \pm 0.67^{\circ}$	DL	NS
C18:0	$32.06 \pm 1.89^{\circ}$	$30.22\!\pm\!1.30^{\rm c}$	25.66 ± 0.61^{b}	18.38 ± 2.36^a	32.11 ± 0.77^{b}	17.44±1.01 ^a	DL, OVX	NS
C18:1	$18.74 \pm 1.80^{\circ}$	9.85 ± 0.54^{ab}	$12.04 \pm 0.32^{\text{b}}$	22.23 ± 0.72^d	$9.36\!\pm\!0.55^{a}$	$17.96 \pm 0.95^{\circ}$	DL, OVX	DL*OVX
C18:2	6.76 ± 0.51^a	18.34 ± 2.93^{b}	6.56 ± 0.31^a	8.84 ± 0.75^{a}	17.75±0.87 ^b	9.35 ± 0.66^{a}	DL, OVX	NS
C18:3	0.07 ± 0.01^a	0.87 ± 0.13^d	0.31 ± 0.05^{b}	0.09 ± 0.03^a	$0.63 \pm 0.06^{\circ}$	0.24 ± 0.02^{ab}	DL	NS
C20:0	1.50 ± 0.09^a	1.94 ± 0.11^a	4.81 ± 0.15^{b}	1.77 ± 0.27^a	1.85 ± 0.28^a	4.06 ± 0.48^{b}	DL	NS
C20:1	0.33 ± 0.16^{b}	0.22 ± 0.11^{ab}	0.14 ± 0.01^{ab}	0.18 ± 0.03^{ab}	0.09 ± 0.02^a	0.15 ± 0.02^{ab}	OVX	NS
C20: 2	0.45 ± 0.07^{b}	0.18 ± 0.02^a	0.15 ± 0.03^a	$0.61 \pm 0.07^{\circ}$	0.11 ± 0.02^{a}	0.12 ± 0.01^{a}	DL	DL*OVX
C20:3	0.17 ± 0.11^{a}	$19.65 \pm 0.79^{\circ}$	17.24±1.15 ^b	ND ⁵⁾	ND	17.50 ± 0.83^{b}	DL, OVX	DL*OVX
C20:4	19.58±0.31 ^b	0.04 ± 0.03^a	0.60 ± 0.44^a	23.98 ± 0.53^{c}	27.99 ± 0.89^a	0.09 ± 0.02^a	DL, OVX	DL*OVX
C20: 5	0.42 ± 0.04^a	1.44 ± 0.50^a	$6.00 \pm 0.39^{\circ}$	0.89 ± 0.13^a	1.14 ± 0.11^{a}	3.05 ± 0.42^{b}	DL, OVX	DL*OVX
C22:6	0.01 ± 0.00^{a}	0.02 ± 0.00^{a}	0.10 ± 0.05^{a}	0.08 ± 0.02^{a}	0.05 ± 0.02^{a}	0.35 ± 0.14^{b}	DL	NS

¹⁾ Mean ± S.E.

Table 6. Fatty acid composition of adipose tissue

(Relative weight %)

Fatty acid	SBT	SSB	SFO	OBT	OSB	OFO	SF ³⁾	Interaction ⁴⁾
C14:0	$11.48 \pm 0.94^{1)a2}$	12.26± 2.42 ^a	19.76±1.03 ^{bc}	18.23±2.25 ^{bc}	14.60± 1.45 ^{ab}	$23.10 \pm 2.36^{\circ}$	DL, OVX	NS
C16:0	12.20 ± 4.36^{a}	$26.80\pm \ 9.20^{ab}$	11.43 ± 5.39^{ab}	31.41 ± 5.39^{ab}	31.93 ± 8.45^{ab}	34.34 ± 3.85^{b}	OVX	NS
C18:0	1.56 ± 1.56^a	0.53 ± 0.51^{a}	2.26 ± 1.52^a	4.51 ± 1.52^a	$5.46\pm\ 1.99^{a}$	10.13 ± 1.77^{b}	OVX	NS
C18:1	5.02 ± 5.02^{ab}	14.06± 6.86 ^b	ND ⁵⁾	2.94 ± 2.56^{ab}	4.61 ± 3.31^{ab}	7.67 ± 4.18^{ab}	NS	NS .
C18:2	$69.54 \pm 7.36^{\circ}$	44.55 ± 12.96^{bc}	58.86 ± 6.81^{bc}	40.49 ± 9.33^{abc}	37.86 ± 11.37^{ab}	14.67 ± 1.83^{a}	OVX	NS
C18:3	0.52 ± 0.06^a	4.67 ± 1.00^{b}	1.48 ± 0.09^{a}	0.55 ± 0.06^a	5.57 ± 0.73^{b}	1.44 ± 0.18^{a}	DL	NS
C20:0	0.02 ± 0.01^{a}	0.03 ± 0.02^{a}	3.20 ± 0.14^{b}	0.02 ± 0.01^{a}	0.08 ± 0.02^{a}	3.03 ± 0.31^{b}	DL	NS
C20:1	0.26 ± 0.03^a	$0.40 \pm\ 0.08^a$	1.06 ± 0.09^{b}	0.51 ± 0.17^a	0.65 ± 0.15^{a}	1.17±0.14 ^b	DL	NS
C20: 2	0.06 ± 0.01^a	0.38 ± 0.12^{b}	0.36 ± 0.07^{b}	0.15 ± 0.07^{ab}	0.38 ± 0.10^{b}	0.37 ± 0.07^{b}	DL	NS
C20:3	0.62 ± 0.09^{a}	0.87 ± 0.24^{ab}	$2.30\pm0.47^{\circ}$	0.84 ± 0.13^{a}	1.68 ± 0.41^{bc}	2.31 ± 0.22^{c}	DL	NS
C20: 4	trace5)	0.32 ± 0.22^{a}	0.88 ± 0.38^{b}	0.15 ± 0.14^{a}	$0.15\pm\ 0.07^{a}$	0.08 ± 0.01^a	DL, OVX	DL*OVX
C20:5	0.09 ± 0.00^{a}	0.17 ± 0.04^{ab}	1.04 ± 0.07^{c}	0.16 ± 0.07^{ab}	0.38 ± 0.08^{b}	$1.06 \pm 0.16^{\circ}$	DL	NS
C22:6	ND	0.02 ± 0.01^{a}	0.12 ± 0.07^{b}	0.06 ± 0.06^a	$0.02\pm\ 0.01^{a}$	0.09 ± 0.04^{ab}	DL_	NS

¹⁾ Mean ± S.E.

Values with different letters in a row are significantly different among experimental groups at p < 0.05 according to Duncan's multiple range test

³⁾ Significant factor(SF): Statistical significance was calculated at the α=0.05 level by 2-way ANOVA

⁴⁾ Interaction between dietary lipid(DL) and ovariectomy(OVX)

⁵⁾ ND: Not Detected

²⁾ Values with different letters in a row are significantly different among experimental groups at p < 0.05 according to Duncan's multiple range test

³⁾ Significant factor(SF): Statistical significance was calculated at the α =0.05 level by 2-way ANOVA

⁴⁾ Interaction between dietary lipid(DL) and ovariectomy(OVX)

⁵⁾ ND: Not Detected

⁶⁾ Trace: Amount is less than 0.01

group had higher amounts of DHA and EPA than that of the other two groups. However, there is no relationship between fatty acid intake and the content of saturated and the monounsaturated fatty acids in adipose tissue. Recently, many studies examined if the fatty acid composition of adipose tissue reflects the dietary lipid intake, especially for certain fatty acids like linoleic acid. In an early study⁴⁰⁾, it was shown that linoleic acid percentages were higher in patients with a history of previous CHD than in patients without previous cardiac disease. Some studies⁴¹⁾ reported a positive correlation between the linoleic acid content of adipose tissue and CHD. But other studies 42) reported that there is negative correlation between the linoleic acid content of adipose tissue and CAD. In a clinical experiment⁴³⁾, the lower the DHA content in adipose tissue, the more severe the extent of atherosclerosis. This result supports that fish oil decreases the risk of CAD, but how DHA content in adipose tissue affects the risk factors of CAD is not clearly understood.

5. G-6-PDH and CAT activity

Table 7 shows the activity of G-6-PDH and CAT in six groups. The activity of G-6-PDH in the fish oil group with ovx was significantly lower than in other groups. Yang and Williams⁴⁴⁾ observed that DHA was more effective in suppressing fatty acid synthesis than was linoleic and linolenic acid. This present experi-

Table 7. Glucose-6-phosphate dehydrogenase activity in blood and Carnitine acetyltransferase activity in liver

Group	G-6-PDH (U/g Hb)	CAT (nmol/mg protein per min)
SBT	19.46±1.781)bc	0.24±0.05ab
SSB	$16.34 \pm 1.08^{ab2)}$	0.16 ± 0.08^a
SFO	20.60 ± 1.18^{bc}	0.29 ± 0.03^{ab}
OBT	20.88±1.01°	$0.29\!\pm\!0.09^{ab}$
OSB	19.67 ± 1.66 ^{bc}	0.11 ± 0.02^{a}
OFO	14.90 ± 1.07^{a}	$0.42\!\pm\!0.10^{\mathrm{b}}$
SF ³⁾	NS	DL
Interaction ⁴⁾	DL*OVX	NS

- 1) Mean + S.E.
- 2) Values with different letters are significantly different among experimental groups at p<0.05 according to Duncan's multiple range test
- 3) Sgnificant Factor(SF) : Statistical significance was calculated at the α =0.05 level by 2-way ANOVA
- 4) Interaction between dietary lipid and ovariectomy

ment showed results similar to the above experiments. The fish oil group showed the highest activity of carnitine acetyltransferase and the soybean oil group showed the lowest activity. The carnitine acetyltransferase activity of the soybean oil group was significantly lower than that of other two lipid groups in ovx. In rat liver, high-fat diets also cause an increase in peroxisomal β -oxidation⁴⁵⁾ and marine oil-rich diets seem to increase the activity of β -oxidative enzymes. The activity of CAT in rat liver was stimulated by a high fat diet and the effect of fish oil was higher than that of soybean oil group.

Conclusion

From the above results, fish oil had a protective effect against the risk factors of CVD such as hypercholesterolemia, hypertriglyceridemia, and obesity. Although there was no significant difference in lipogenic enzyme activity, lipolytic enzyme activity was high in the fish oil group compared with the soybean oil group and the beef tallow group.

Literature cited

- Choe EH, Byun YC. Recent pattern of mortality in Korea. J Popul Associ Korea 8(2): 67, 1985
- National Bureau of Statistics, Economic Planning Board, Republic of Korea. Annual report on the cause of death statistics, 1982-1990
- 3) Gordon T, Kannel WB, Hjortland MC, McNamara PM. Menopause and coronary heart disease. The Framingham study. *Ann Intern Med* 89(2): 157-161, 1978
- 4) Kannel WB, Hjortland MC, McNamara PM, et al. The Framingham study. *Ann Intern Med* 85: 447-452, 1976
- 5) Bengtsson C, Rybo G, Westerberg H. Number of pregnancies, use of oral contraceptives and menopausal age in women with ischemic heart disease, compared to a population sample of women. In: Ischemic heart disease in women. Acta Med Scand(Suppl) 549: 75-81, 1973
- 6) Ritterband AB, Jaffe IA, Densen PM, et al. Gonadal function and the development of coronary heart disease. *Circulation* 27: 237-251, 1963
- Rosenberg L, Hennekens CH, Rosner B, et al. Early menopause and the risk of myocardial infarction. Am J Obstet Gynecol 139: 47-51, 1981
- 8) Kannel WB, Dawber TR, Kagan A, et al. Factors of risk in the development of coronary heart disease-six year fol-

- low-up experience. The Framingham study. Ann Intern Med 55: 33-50, 1961
- Robinson RW, HiGano N, Cohen WD. Increased incidence of coronary heart disease in women castrated prior to the menopause. Arch Intern Med 104: 908-913, 1959
- Parrish HM, Carr CA, Hall DG, King TM. Time interval from castration in premenopausal women to development of excessive coronary atherosclerosis. Am J Obstet Gynecol 97: 155-162, 1967
- 11) Mook DG, Kenney NJ, Roberts S, Nussbaum AI, Rodier WI. Ovarian-adrenal interactions in regulation of body weight by female rats. J Comp Physiol 81: 198-211, 1972
- 12) Tarttelin MF, Gorski RA. The effect of ovarian steroids on food and water intake and body weight in the female rat. Acta Endocr 72: 551-568, 1973
- 13) Borkan GA, Sparrow D, Wisniewski C, Vokon PS. Body weight and coronary disease risk. Am J Epidemiolo 124: 410-419, 1986
- 14) Park HS, Cho HJ, Kim YS, Kim CJ. The diseases associated with obesity in Korean adults. *J Korean Acad Fam Med* 13(4): 344-353, 1992
- 15) London SJ, Sacks FM, Caesar J, Stampfer MJ, Siguel E, Willett WC. Fatty acid composition of subcutaneous adipose tissue and diet in postmenopausal US Women. Am J Clin Nutr 54: 340-345, 1991
- 16) Reeves PG. AIN-76 diet: Should we change the formulation? *J Nutr* 119: 1081-1082, 1989
- 17) Fredelwald WT, Levy RI, Fedreiczon DS. Estimation of low density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clin Chem 18: 499, 1979
- Fletcher DL, Britton WM, Cason JA. A comparison of various procedures for determining total yolk lipid content. *Poul Sci* 63: 1759-1763, 1984
- Lepage G, Roy CC. Direct transesterification of all classes of lipids in a one-step reaction. J Lipid Res 27: 114-120, 1986
- 20) Kornberg A, Horecker BL. Glucose-6-phosphate dehydrogenase. In: Methods in enzymology. SP. Colowick, NO Daplan, Editors, vol. 1 Academic Press, New York, 323, 1955
- 21) Lohr GW, Waller HD. Glucose-6-phosphate dehydrogenase: In: Methods of enzymatic analysis. HU Bergmeyer, Editor, Academic Press, New York, 636, 1974
- Bieber LL, Markwell MAK. Carnitine acetyltransferases. Methods in enzymology 71: 351, 1981
- Alexander RR, Grittiths JM, Wilkinson ML. Basic Biochemical Methods 161-162, 1985
- 24) Lowry OH, Rosebrough WJ, Farr AL, Randall RT. Protein mesurement with folin phenol reagent. J Biol Chem

- 193: 265-275, 1951
- 25) Jones PJH. Effect of fatty acid composition of dietary fat on energy balance and expenditure in hamsters. Can J Physiol Pharmacol 67: 994-998, 1989
- 26) Parrish CC, Pathy DA, Angel A. Dietary fish oils limit adipose tissue hypertropy in rats. *Metabolism* 39: 217-219, 1990
- 27) Shimomura Y, Tramura T, Suzuki M. Less body fat accumulation in rats fed a safflower oil diet than in rats fed a beef tallow diet. J Nutr 120: 1291-1296, 1990
- 28) Fumeron F, Brigant L, Parra HJ, Bard JM, Fruchart JC, Apfelbaum M. Lowering of HDL₂-cholesterol and lipoprotein A-1 particle levels by increasing the ratio of polyunsaturated to saturated fatty acids. Am J Clin Nutr 53: 655-659, 1991
- 29) Berr F, Goetz A, Schreiber E, Paumgartner G. Effect of dietary n-3 versus n-6 polyunsaturated fatty acids on hepatic excretion of cholesterol in the hamster. *J Lipid Res* 34: 1275-1284, 1993
- 30) Spady DK, Dietschy JM. Interaction of dietary cholesterol and triglycerides in the regulation of hepatic low density lipoprotein transport in the hamster. *J Clin Inves* 81: 300-309, 1988
- 31) Smith MJ, Verkade HJ, Havinga R, Vonk RJ, Scherphof GL, Veld G, Kuipers F. Dietary fish oil potentiates bile acid-induced cholesterol secretion into bile in rats. *J Lipid Res* 35: 301-310, 1994
- 32) Kim SH, Lee LH, Lee JM, Kim WY, Kim MK. The evaluation of health and nutritional status between Korean and Western population based on lipid consumption pattern. KOSEF, 1993
- 33) Simons LA, Balasubramaian S, Hackie JB: Reduction in plasma cholesterol and increase in biliary cholesterol by a diet rich in n-3 fatty acids in rat. J Lipid Res 26: 684-689, 1985
- 34) Jung SE, Ha TY, Im JG, Cho SH. The study of biochemical change induced by fish oil diet in rat(I)-changes in hepatic lipogenic enzyme activity. Korean J Nutr 17(4): 290-296, 1984
- 35) Connor WE. Hypolipidemic effects of dietary omega-3 fatty acids in nornal and hyperlipidemic humans. In Health effects of polyunsaturated fatty acids in sea foods. Academic Press, 173: 210, 1986
- 36) Nestel PJ. Effects of n-3 fatty acids on lipid metabolism. Ann Rev Nutr 10: 149-167, 1990
- 37) Nam JH, Park HS. Differential effect of n6 and n3 polyunsaturated fatty acids on plasma lipids in rat fed low and high fat diets. *Korean J Nutr* 24(4): 314-325, 1991
- 38) Park HS, Lee SM. Effects of dietary n-3 fatty acids and fat unsaturation on plasma lipids and lipoproteins in

- rats. Korean J Nutr 25(7): 555-567, 1992
- 39) Harris WS, Connor WE, McMurry MP. The comparative reductions of the plasma lipids and lipoproteins by dietary polyunsaturated fats: Salmon oil versus vegetable oils. *Metabolism* 32(2): 179-184, 1983
- 40) Kirkeby K, Ingvaldsen P, Bjerkedal I. Fatty acid composition of serum lipids in men with myocardial infarction. *Acta Med Scand* 192: 513-519, 1972
- 41) Hodgson JM, Wahlquist ML, Boxall JA, Balazs ND. Can linoleic acid contribute to coronary artery disease? *Am J Clin Nutr* 58: 228-234, 1993
- 42) Riemersma RA, Wood DA, Butler S, et al. Linoleic acid

- in adipose tissue and coronary heart disease. Br Med J 292: 1423-1427, 1986
- 43) Seidelin KN, Myrup B, Fisher-Hansen B. N-3 fatty acids in adipose tissue and coronary artery disease are inversely related? *Am J Clin Nutr* 55: 1117-1119, 1992
- 44) Yang YT, Williams MA. Comparison of C18-, C20- and C22- unsaturated fatty acids in reducing fatty acid synthesis in isolated rat hepatocytes. *Biochim Biophys Acta* 531: 133-140, 1978
- 45) Neat CE, Thomassen MS, Osmundsen H. Induction of peroxisomal β-oxidation in rat liver by high-fat diets. *Biochem J* 186 : 369-371, 1980