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A Study on the Nucleotide Analysis of 18S rRNA and the Molecular Evolution of the Korean Decapods (II)

Kim, Won, Min, Gi Sik and Kim, Sang Hee

(Department of Molecular Biology, College of Natural Sciences, Seoul National University,
Seoul 151-742, Korea)

한국산 십각류의 18S 리보솜 RNA의 염기분석과 분자진화에 관한 연구(II)

김 원·민 기 식·김 상 희

(서울대학교 자연과학대학 분자생물학과)

적 요

중합효소연쇄반응(PCR)을 이용한 클로닝 기법과 Taq 염기서열분석법을 사용하여 갑각류에 속하는 뿔물맞이게(*Pugettia quadrident*) (십각목, 범배아목, 게하목)에 대한 18S 리보솜 RNA 유전자의 1차염기서열을 밝혔다. 본 종의 18S 리보솜 RNA 유전자는 십각류에 속하는 또 다른 종인 두드러기어리게 (*Oedignathus inermis*)보다 46개가 짧은 1837개의 염기로 이루어져있었다. 염기가 삽입되거나 결실된 부분을 고려하지 않았을때에 두 종간에 염기서열 유사도는 90.8%이었다. 염기서열이 가장 보존적인 부위는 1137-1206(70개) 부위로 이 부위에서는 두 종이 완전히 동일한 염기서열을 보이고 있었고, 변이가 연속적으로 가장 큰 부위는 46-55 부위였고 399-407 부위가 그 다음으로 많은 연속적 변이를 가지고 있었다. 18S 리보솜 RNA 유전자의 1차구조에 있어서 염기서열의 변이는 이 유전자 전체를 통해 고르게 분포하지 않았다.

Key words: *Pugettia quadrident*, 18S rRNA, Decapoda, PCR cloning.

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INTRODUCTION

Because of the conservative nature, the nucleotide sequences of 18S rRNA have been widely adopted in the construction of molecular phylogenies among the remotely related eukaryotes in recent years (Field *et al.*, 1988; Stock and Whitt, 1992). However, most studies used the partial sequences without complete understanding of the sequence variability of each species representing certain taxonomic categorical rank. Even though the results from those studies are congruent to those from the morphological data in many cases, the 18S rRNA shows sequence variability, such as conservative, variable, and highly variable, across the molecule. Therefore the nucleotide analysis of the complete sequences of many species will provide the further understanding of the evolutionary relationships among the organismal groups, especially when the groups being compared belong to the lower categorical rank.

As a part of study in examining sequence variability and general patterns of nucleotide substitution of the 18S rRNA gene with the complete sequences among crustaceans, the 18S rRNA gene from one decapod species, *Pugettia quadrident*, was cloned and sequenced. In crustacean decapods, there is only one species, *Oedignathus inermis*, in which the complete nucleotide sequences of 18S rRNA gene were known (Kim *et al.*, 1992). Therefore we compared the nucleotide sequences of *P. quadrident* with that of *O. inermis*.

MATERIALS AND METHODS

In this study, the 18S rRNA gene of *Pugettia quadrident* was sequenced by PCR cloning and Taq sequencing as Kim *et al.* (1992). This species belongs to the infraorder Brachyura, suborder Pleocyemata, order Decapoda in Crustacea (Bowman and Abele, 1982). The sequence datum of *O. inermis* is from Kim *et al.* (1992) and this species belongs to infraorder Anomura, order Decapoda. We aligned the nucleotide sequences of *O. inermis* and *P. quadrident* using FASTA program (Pearson and Lipman, 1988). The numbers indicating nucleotide position in the following text are those of *O. inermis* when the sequences are numbered without the insertion and/or deletion.

RESULTS

The total length of the 18S rRNA gene of *P. quadrident* is 1837 bases long, and 46 bases shorter than *O. inermis* sequences (Fig. 1). The occurrence of each nucleotide in the two species ranges in 23.3-26.9%. The G + C content is around 50% in each species (Table 1).

The total number of different, identical, and null nucleotides between two species are 167, 1655, and 76, respectively. The similarity between two species is 90.8% when the insertion and/or deletion sites were excluded. The number of different nucleotides in a row varies 1 to 10 and the one nucleotide difference is the majority (86 out of 167). If the insertion and/or deletion sites are included and these sites are considered consider as the unweighted nucleotide differences between two species, the similarity is 87.2% (Table 2). The ratio of transition/transversion is 0.72 in the present two species. There are 34 insertion regions

	10	20	30	40	50	60
Oed	TACCTGGTGTACCTGCCAGTAGTCATAATGCTTGTCAAAGATTAAAGCCATGCATGTCT					
Pug	TACCTGGTGTACCTGCCAGTAGTCATAATGCTTGTCGGTGTCTGGCTGCTGTCT					
	70	80	90	100	110	120
Oed	AACTACAAGCCGATGTAAGGTGAAACCGCGAATGGCTCATTAAATCAGCTATGTTCTT					
Pug	AACTACAAGCCGATTCAGGCAGAACCGCGAATGGCTCATTAAATCAGCTATGATTCTT					
	130	140	150	160	170	180
Oed	GGAACCTGACCCCCACTTACTTGGATAACTGTGGTAATTCTAGAGCTAATACATGCACAC					
Pug	GGATCTGTACCCACATTTACTTGGATAACTGTGGTAATTCTAGAGCTAATACATGCCA-					
	190	200	210	220	230	
Oed	AGAGTCCCCGACCCAGGGAGGGCGCTTTATTAGTCAAAACCGGTCGGGCTC--G					
Pug	-GAGTCTCTG-ACCGCAAGGAAGAGCGCTTTATTAGTCAAAACCGGTCGGGCTTCGG					
	240	250	260	270	280	290
Oed	GTCCGTAACCAACC-TGTGGTGAATCTGAATAACTTGTACTGAGCGCACGGTCTCCGACT					
Pug	GTCCGTCACCTGGTGTGAATCTGAATAACTTT-CTCA-CGCACGGTCTCCGCGAG					
	300	310	320	330	340	350
Oed	GGCGCCGCATCTTCAAGTGTCTGCCTTATCAGCTTCGATTCGA-GGTTATT-CGCCCTT					
Pug	GGC--CGCCTTTCAGTGT-TGCCTTATCAGCTTCGATTGTAAGGTTACTACGCCCTT					
	360	370	380	390	400	410
Oed	CCATGGCTATTACGGTTACGGGAATTCAAGGTTACGGGAGCTTCCGGAGAGGGAGCCTAGGA					
Pug	CAATGGCTATTACGG-TTACGGGTAAAT-CAGGGTTTATTCCGTTTCCCCCCTGGGA					
	420	430	440	450	460	470
Oed	ACGGCTACCATCTAACGGAGGGAGGAGCTAAATTACCCATTCCAGACCGGGGA					
Pug	ACGGGTACCATCTAACGGAGGGCGGAGGGACGGCAATTACCAAC-CCGGCACGGGGAA					
	480	490	500	510	520	530
Oed	GGTAGTGAACGAAAATAACCGATGCGAGACTCATCCGAGGCTTCAATCGGAATGAGTAC					
Pug	GGTAGTGAACGAAAATAACCGATGCGAGACTCATCCGAG-CCTCCAATCGGAATGAGTAC					
	540	550	560	570	580	590
Oed	ACTTTAAATCTTAAACGAGGGACCCATTGGAGGGCAAGTCTGGTGCCTTAAAGCTCCTAGTTGGAT					
Pug	ACTTTAAATGGTTAACGAGGATCCATTGGAGGG-CAAGTCTGGTGCCTAGCAGGCCGGT					
	600	610	620	630	640	650
Oed	TAATTCCAGCTCCAATA-CGTATATTAAAGTTGTTGGTTAAAAG-TCGTAGTTGGAT					
Pug	-ATTCCAGCTCCAATAGCGTATATTAAAGTTGTTGGTTAAAAGCTCCTAGTTGGAT					
	660	670	680	690	700	710
Oed	TACAGTTCCGGAACGTACGGTTACGGCCGGTGCCTACTGTCACGCCCGAACAGCCTGAA					
Pug	TTCAAGTTCTGGACTGACGGTTACGGCCGGTGTACTGTCACGCCCGAACACTTGTAC					
	720	730	740	750	760	770
Oed	ACATGGGCCCGCGCTCGCCGGGTCTCTTACCGAGTGTCCCGAGTGGCCGGCATGT					
Pug	-CAT---CCGCTGGCNAC-GGGTCCCCTTTCCCAATTCCCTG-GCCCGAGAGT					
	780	790	800	810	820	830
Oed	TTACTTTGAAAAAAATTAGAGTGCTACAGAGCAGGCTACATGAATTGGCTGAATGCTATG					
Pug	TTACTTTGAAAAAAATTAGAGTCTCAAAGCAGGCTACACTGAC-GGCCTGAATGCCCTATG					
	840	850	860	870	880	890
Oed	CATGGAATAATGGAATAGGACCTCGGTTCTATTAGTACTCGAGGCCAGGGTGAATTCTGGGA					
Pug	CATGGAATAATGGAATAGGACCTCGGTTCTATTAGTACTCGAGGCCAGGGTGAATTCTGGGA					
	900	910	920	930	940	950
Oed	AATGACTAATAGAAACAAGGCGGGGTATTAGTACTCGAGGCCAGGGTGAATTCTGGGA					
Pug	AATGACTAATGGAA-CAGGCGGGTCAATT-TATTGACGCTAGGGT-AAATTCTGGGA					
	960	970	980	990	1000	1010
Oed	CCGTCGCAAGACTTACAACGCAAAGTATTACCAAGGATGTTTCATTAATCAAGAACG					
Pug	CCGTCGCAAGACGAACTACTCGAAATCATTTGCCAAGGATGTTTCATTAATCAAGAACG					

Oed	1020	1030	1040	1050	1060	1070
Pug	AAAAGTTAGAGGTTCGAAGGGCATCAGATACCGCCCTAGTTCTAACATAACGATGCTGA					
Oed	1080	1090	1100	1110	1120	1130
Pug	CTACGATCCGCCGGCGTTATTCCCATGACCGGGCGGCAGCTTCCGGAAACCAAAGTCTT					
Oed	1140	1150	1160	1170	1180	1190
Pug	CCAC-ATCCGCCGGAGTTATTCCCATGACCGGGCGGGAGCTTCCGGAAACCAAAGTCTT					
Oed	1200	1210	1220	1230	1240	1250
Pug	CACCAAGGAGTGGCATGCCGCTTAATTGACTCAACACGGGAACCTCACCAAGGCCAGAC					
Oed	1260	1270	1280	1290	1300	1310
Pug	ACCGGAAGGATTGACAGATTGAGAGCTTCTTGATTCGGTGGTGGTGGCATGGCC					
Oed	1320	1330	1340	1350	1360	1370
Pug	GTTCTTAGTTGGTGGAGCGATTGTCTGGTTAACCGATAACGAACGAGACTCTAGCCT					
Oed	1380	1390	1400	1410	1420	1430
Pug	ACTAACTAGTCGACGGATCTCCAGCAATTGGTGTCCAGTCGAACTTCTTCTAGAGGA					
Oed	1440	1450	1460	1470	1480	1490
Pug	TAACGGCAACTCTAACGCCGACGAGAACGATTCAGCAATAACAGGTCTGTGATGCCCTTAGAT					
Oed	1500	1510	1520	1530	1540	1550
Pug	AACGGGCAATTCTA-GCCGCACGAGA-TTGAGCAATAACAGGCTGTGATGCCCTTAGAT					
Oed	1560	1570	1580	1590	1600	1610
Pug	GGGTAACCCCTATGAAACCCCTTCATGATAGGGATTGGGCTTGCATTGTT-TCCCATG					
Oed	1620	1630	1640	1650	1660	1670
Pug	AACGAGGAATTCCCAAGTAAG-CGCAAGTCATCAGCTTGCCTGATTACCGTCTGCCCTT					
Oed	1680	1690	1700	1710	1720	
Pug	-GTACACACCCCCCG-TCGCTACTACCGATTGAATGATTAGTGAG-CTTCGGACTG-CGC					
Oed	1730	1740	1750	1760	1770	1780
Pug	CGTACACACGCCGGTCGCTACTACCGATTGAATGATTAGTGAGGCTTCGGATTGGCGC					
Oed	1790	1800	1810	1820	1830	1840
Pug	TCTTGGATGCCCTGGC-----CCGCCTTC-----CCGTGG--GCTTTTAG					
Oed	1850	1860	1870	1880		
Pug	GGCCCTCGGGCTGACGAAAGATGTCACACTTGATCATTTAGAGGAAGTAAAAGTCGTA					
Oed	ACAAGGTTCCGTAGGTGAAAC-TGCGGAAGGATCATT					
Pug	GCGCCTCGAGCTGACT-AAAGATGTCACACTTGATCATTTAGAGGAAGTAAAAGTCGTA					

Fig. 1. The nucleotide sequences of 18S rRNA gene of *O. inermis* and *P. quadridentis* aligned by FASTA program

$\Omega_{\text{ed}} = \Omega_{\text{inermis}}$; $P_{\text{ug}} = P_{\text{quadridentes}}$

Table 1. The total length, nucleotide composition, and G + C content of the 18S rRNA gene in *O. inermis* and *P. quadrident*s.

Species	Total length(base)	No. of each nucleotide				G + C content
		A	C	G	T	
<i>O. inermis</i> *	1883	468 (24.8%)	439 (23.3%)	507 (26.9%)	469 (24.9%)	50.20%
<i>P. quadrident</i> s	1837	444 (24.2%)	437 (23.8%)	481 (26.2%)	474 (25.8%)	49.97%

* Data from Kim *et al.* (1992)

Table 2. Pairwise comparison of nucleotide sequences in Art/Oed* and Oed/Pug. Art = *A. salina*; Oed = *O. inermis*; Pug = *P. quadrident*s.

	Art/Oed	Oed/Pug
No. of nucleotide		
— identical	1552	1655
— different	237	167
— null (insertion and/or deletion)	115	76
Similarity(%)		
— with null sites	81.5	87.2
— without null sites	86.8	90.8
Transition / transversion (ratio)	1.26	0.72

* Data from Kim *et al.* (1992)

in *O. inermis* and 14 in *P. quadrident*s. The number of nucleotide in each insertion region varies between 1 and 12. The longest insertion region is found at the position of 1758-1769 and the second most at 1742-1749 (Fig. 1). The sequence variation in the primary structure of 18S rRNA gene are not evenly distributed throughout the molecule as indicated by Kim and Abele (1990).

DISCUSSION

Kim *et al.* (1992) compared the nucleotide sequences of *A. salina* (class Branchiopoda) and *O. inermis* to see the sequence variability between class levels and the present study compared the nucleotide sequences between infraorder levels (between Anomura and Brachyura) in Decapoda. Result from the present study show that the molecular data are congruent to the morphological data in terms of the higher similarity between infraorder levels than between class levels (Table 2).

It has been known that the nucleotide sequences of the 3'-terminal region is well conserved among many species. In the present two decapod species, the 3'-terminal region (position of 1804-1883) also shows

only one nucleotide insertion and/or deletion among total 81 nucleotides. Therefore this region appears to be quite conservative, while the most conservative (identical) region locates at the position of 1137-1206 and it is 70 bases long. The most variable nucleotide differences in a row occur at the position between 46-55 and the second most between 399-407. The nucleotide sequences of these regions are identical between *A. salina* (class Branchiopoda) and *O. inermis*. Thus the differences between *O. inermis* and *P. quadridentata* in these regions are due to *P. quadridentata* sequences. Because *A. salina* and *O. inermis* are considered to be more primitive than *P. quadridentata*, the different sequences of *P. quadridentata* in these regions seems to be the nucleotide substitutions occurred in the lineage leading *P. quadridentata*.

We also compared the above three species together to see the insertion and/or deletion pattern between class levels by using Multialign programs (Corpet, 1988; sequence datum of *A. salina* from Nelles et al. 1984). The results of the multialignment show that there are 54 insertion regions where two decapod species show nucleotide sequences but no sequences in branchiopod species. Thus these regions may be important in recognizing the difference of class levels of Crustacea.

Our results indicate that many additional complete sequences are necessary to determine the sequence variability of 18S rRNA gene of each species among different taxonomic categorical groups. These sequence variability will give more elucidation to construct the molecular phylogenies and to see the molecular evolutionary patterns among the groups of other eukaryotic organisms as well as crustaceans.

ABSTRACT

The primary sequence of the 18S rRNA gene of a crustacean *Pugettia quadridentata* (Decapoda: Pleocyemata: Brachyura) was determined by the PCR cloning and Taq sequencing. The 18S rRNA gene of this species is 1837 bases long, and 46 bases shorter than that of another crustacean decapod *Oedignathus inermis*. The similarity between two species is 90.8% when the insertion and/or deletion sites were excluded. Within the molecule, the most conservative (identical) region locates at the position of 1137-1206 and it is 70 bases long. The most long consecutive nucleotide differences occur at the position between 46-55 and the second most between 399-407. The sequence variation in the primary structure of 18S rRNA gene are not evenly distributed throughout the molecule.

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