

ASSAY OF POTENTIAL ANTIMUTAGENICITY OF ETHNIC MEDICINAL PLANT EXTRACTS OF PAPUA NEW GUINEA BY USING SOS CHROMOTEST (*E. COLI* PQ 37)

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ABSTRACT: Thirty six crude drug samples have been prepared from different parts of twenty five plants belonging to different families, and antimutagenic activities were studied by using SOS chromotest (*E. coli* PQ 37). The following crude extracts of PNG medicinal plants which had a appreciable antimutagenic activity against mitomycin C were: *Artocarpus communis* (stem bark), *Cycas circinalis* (leaves), *Merremia peltata* (leaves), *Intsia palembanica* (leaves), *Annona muricata* (stem bark), and *Artocarpus altilis* (root bark).

INTRODUCTION

The people of third world countries have developed their own traditional medicines using different medicinal plants, which are as diverse as the people themselves. But the actual constituents and antimutagenicity and mutagenicity of the most of the folk medicines have not been studied. The previous research reports revealed that a number of traditional medicinal plants showed genotoxicities in microorganisms as well as in mammals (Wang and Hu, 1985; Chang and Chi, 1981; Chang and Chi, 1982; Chang *et al.*, 1982 (a), Chang *et al.*, 1982 (b), Kalantary *et al.*, 1986). Naturally occurring antimutagens in plant and animal sources have been studied by using different test techniques. A few research communications were available on antimutagens in crude drugs, which are frequently used as a traditional medicine, and studied the antimutagenicity of some of these plants (Kakinuma *et al.*, 1984 (a) Nakamura and Yamamoto, 1982; Kakinuma *et al.*, 1984 (b), Minakata *et al.*, 1983; Sakai *et al.*, 1986).

The present study aimed to investigate potential antimutagenic activities of ethnic medicinal plants collected in Papua New Guinea by using SOS chromotest.

MATERIALS AND METHODS

Plant Materials

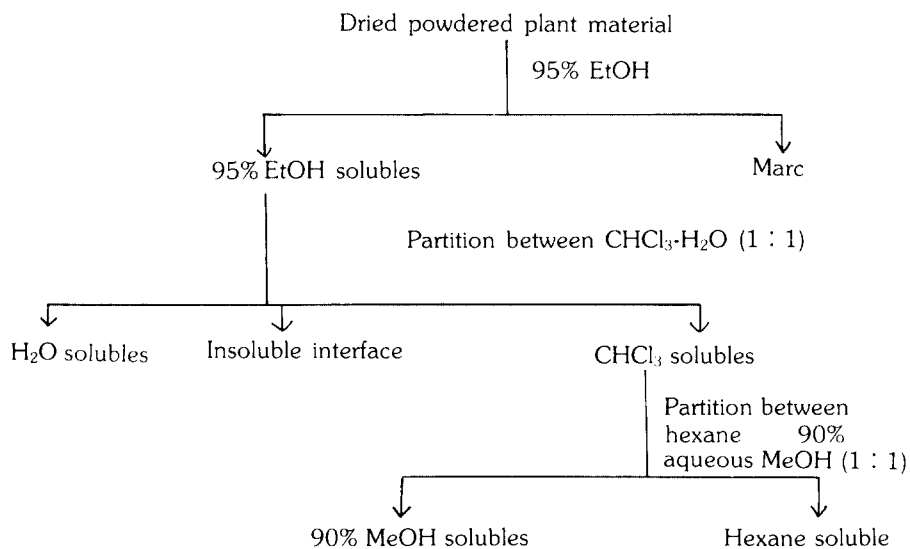
The following Papua New Guinea native medicinal plants were identified and collected in Papua New Guinea based on ethno-medicinal information:

Table 1. Papua New Guinean plants used for experiment.

Scientific name	Family	Part Used
<i>Artocarpus communis</i> J.R. & G. Forster	Moraceae	Stem bark
<i>Plumeria rubra</i> L.	Apocyanaceae	Stem bark/leaves
<i>Alstonia spectabilis</i> R. Br	Apocyanaceae	Stem bark leaves
<i>Kleinhovia hospita</i> L.	Sterculiaceae	Stem bark/leaves
<i>Cycas circinalis</i> L.	Cycadaceae	Seeds/leaves
<i>Dracaena angustifolia</i> Roxb.	Agavaceae	Leaves
<i>Desmodium umbellatum</i> L.	Leguminosae	Leaves
<i>Timonius timon</i> (Spreng) Merr.	Rubiaceae	Leaves
<i>Securinega melanthesoides</i> (F. Muell) Airy Shaw	Euphorbiaceae	Leaves
<i>Merremia peltata</i> L.	Convolvulaceae	Leaves
<i>Intsia palembanica</i> Miq.	Leguminosae	Leaves
<i>Vitex trifolia</i> L.	Verbenaceae	Leaves
<i>Melanolepis multiglandulosa</i> (Reinw. ex Bl.)	Euphorbiaceae	Leaves
<i>Wedelia biflora</i> L.	Compositae	Leaves
<i>Cordia dichotoma</i> Forst	Boraginaceae	Leaves
<i>Pipturus argenteus</i> (Forst.) Wedd	Urticaceae	Leaves
<i>Pterocarpus indicus</i> Willd	Leguminosae	Leaves
<i>Annona cherimolia</i> Mill	Annonaceae	Seeds/Root bark/ Stem bark
<i>Annona muricata</i> L.	Annonaceae	Seeds/Root bark/ Stem bark
<i>Cananga odorata</i> Hook	Annonaceae	Stem bark
<i>Popowia polyandra</i> Merr.	Annonaceae	Stem bark
<i>Artocarpus altilis</i>	Moraceae	Seeds/Root bark/ Stem bark
<i>Morinda citrifolia</i> L.	Rubiaceae	Leaves/Stem bark/ Root bark
<i>Breynia cernua</i> Muel. Arg.	Euphorbiaceae	Leaves

Extraction

The following extraction procedures was used (Rupprecht *et al.*, 1990). The dried plant materials were extracted with 95% ethyl alcohol. This was concentrated, then followed by solvent partitioning using chloroform-water (1:1, v/v). The chloroform solubles were concentrated and partitioned between hexane-90% aqueous methanol (1:1, v/v). The 90% methanol solubles were concentrated under vacuum by rota evaporator. The dried samples were dissolved in DMSO and used for antimutagenicity assay.

Scheme 1. Standard flow sheet of extraction and initial partitioning.

Source: Reprecht *et al.*, 1990.

Table 2. β -Galactosidase activities induced by plant extracts.

Species	Plant Part	β -Galactosidase activity (unit) (4 mg/ml)*
Mitomycin C (0.03 μ g/ml)		339
DMSO		54
<i>Artocarpus communis</i>	stem bark	135
<i>Plumeria rubra</i>	stem bark	214
<i>Plumeria rubra</i>	leaves	243
<i>Alstonia spectabilis</i>	stem bark	329
<i>Alstonia spectabilis</i>	leaves	242
<i>Kleinhovia hospita</i>	stem bark	150
<i>Kleinhovia hospita</i>	leaves	195
<i>Cycas circinlis</i>	seeds	105
<i>Cycas circinalis</i>	leaves	129
<i>Dracaena augustifolia</i>	leaves	202
<i>Desmodium umbellatum</i>	leaves	198
<i>Securinega melanthesoides</i>	leaves	270
<i>Merremia peltata</i>	leaves	125
<i>Intsia palembanica</i>	leaves	131
<i>Vitex trifolia</i>	leaves	270
<i>Melanolepsis multiglandulosa</i>	leaves	289
<i>Wedelia bioflora</i>	leaves	353
<i>Cordia dichotoma</i>	leaves	142
<i>Pipturus argentens</i>	leaves	219

Continued from Table 2.

Species	Plant Part	β -Galactosidase activity (unit) (4 mg/ml)*
<i>Pterocarpus indicus</i>	leaves	238
<i>Annona cherimolia</i>	seeds	198
<i>Annona cherimolia</i>	root bark	145
<i>Annona cherimolia</i>	stem bark	212
<i>Annona muricata</i>	seeds	184
<i>Annona muricata</i>	stem bark	96
<i>Annona muricata</i>	root bark	311
<i>Cananga odorata</i>	stem bark	326
<i>Cananga odorata</i>	root bark	151
<i>Popwia polyandra</i>	stem bark	195
<i>Artocarpus altilis</i>	seeds	189
<i>Artocarpus altilis</i>	root bark	111
<i>Artocarpus altilis</i>	stem bark	142
<i>Morinda citrifolia</i>	leaves	210
<i>Morinda citrifolia</i>	stem bark	286
<i>Morinda citrifolia</i>	root bark	233
<i>Breynia cernua</i>	leaves	350

*concentration of each extract

Assay of Antimutagenicity by Using SOS Chromotest (*E. coli* PQ3)

The SOS Chromotest (*E. coli* PQ37) was described by Chang *et al.* (1987) elsewhere.

RESULTS AND DISCUSSION

Antimutagenic activities by using SOS Chromotest (*E. Coli* PQ37) for PNG medicinal plants crude extracts against mitomycin C were presented in Table 1. The results indicate that *Cycas circinalis* (seeds), *Annona nuricata* (stem bark), and *Artocarpus altilis* (root bark) showed significant antimutagenic effect (96-111 units) against mitomycin C compared to positive control (339 units). The nine other crude extracts which had a appreciable antimutagenic effect (~150 units) were: *Artocarpus communis* (stem bark), *Kleinhovia hospita* (stem bark) *cycas circinalis* (leaves), *Merremia peltata* (leaves), *Intsia palembanica* (leaves), *Cordia dichotoma* (leaves), *Annona cherimolia* (root bark), *Cananga Odorata* (root bark) and *Artocarpus altilis* (stem bark). The other plants crude extracts had a lower antimutagenic activities.

REFERENCES

- Chang, I.-M. and Chi, H.J. (1981): Toxicoicity and antitumor activities of Korean Medicinal Plants (1). *Korean J. Pharmacog.*, **12**, 125-130.
- Chang, I.-M., Kim, Y.S. and Han, B.H. (1982a): Toxicological evaluation of medicinal plants used for herbal drugs (II); Acute toxicity and effects on DNA

- biosynthesis in bone marrow cells and hemoglobin content in blood. *Korean J. Pharmacog.*, **13**, 14-19.
- Chang, I.-M. Kim, J.H. and Han, D.S. (1982b): Toxicological evaluation of medicinal plants used for herbal drugs (IV), acute toxicity and antitumor activities. *Korean J. Pharmacog.*, **13**, 62-69.
- Chang, I.-M. and Chi, H.J. (1982): Toxicological evaluation of medicinal plants used for herbal drugs (III): cytotoxicity and antitumor activities against Glioma (9ASK). *Korean J. Pharmacog.*, **13**, 55-61.
- Chang, I.-M., Guest, I.C., Lee-Chang, J., Paik, N.W., Jhoun, J. and RYun, R.Y. (1987): Assay of potential mutagenicity and antimutagenicity of Chinese herbal drugs by using SOS chromotest (*E. Coli* PQ37) and SOS UMU test (*S. Typhimurium* TA 1535/PSK 1002). Proceeding of the first Korea-Japan Toxicology Symposium Safety Assessment of Chemicals *In Vitro*. The Korean Society of Toxicology, pp. 133-145.
- Kakinuma, K., Okada, Y., Ikegawa, N., Kada, T. and Nomoto, M. (1984a): Antimutagenic diterpenoids from a crude drug *Isodonis herba* (Enmei-SO). *Agr. Biol. Chem.*, **48**, 1647-1948.
- Kakinuma, K., Koike, J. Kotani, J., Ikegawa, N., Kada, T. and Nomoto, M. (1984b): Cinnamaldehyde: Identification of an antimutagen from a crude drug, *Cinnamomi cortex*. *Agric. Biol. Chem.*, **48**, 1905-1906.
- Kalantary Gotvandi, H.N., Zong, M. and Chang, I.-M. (1986): Toxicological study on traditional Korean herbal drugs (V). *Korean J. Toxicol.*, **2**, 79-87.
- Minakata, H., Komura, H., Nakanishi, K. and Kada, T. (1983): Protoanemonin, an antimutagen isolated from plants. *Mutation Res.*, **116**, 317-322.
- Nakamura, H. and Yamamoto, T. (1982): Mutagen and antimutagen in ginger, *Zingiber officinale*. *Mutation Res.*, **103**, 119-126.
- Rupprecht, J.K., Hui, Y.H. and McLaughlin, J.L. (1990): Annonaceous acetogenins: A review *J. Natural Products*, **53**, 237-278.
- Sakai, Y., Nagase, H., Ose, Y., Sato, Y., Yamada, A., Hibi, M. and Yamada, F. (1986): Antimutagenicity of extracts from crude drugs in Chinese medicines. *Mutation Res.*, **174**, 1-4.
- Wang, Y.-M. and Hu, Y.-J. (1985): Toxicity and side effects of some hinese medicinal herbs in *Advanced in Chinese Medicinal Materials Research* (Chang H.M., Yeung, H.W., Tso W.-W. and Koo A. Ed.), World Scientific Pub. Singapore, pp. 109-123.