

Saponins from the Fructus of *Kochia scoparia*

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Abstract □ Two new triterpenoidal saponins, B(1) and C(2) were isolated from the fructus of *Kochia scoparia*. On the basis of chemico-spectral evidences, the structures of 1 and 2 were elucidated as oleanolic acid 3-*O*-β-D-ribofuranosyl-(1→2)-β-D-glucuronopyranoside and 3-*O*-β-D-xylofuranosyl-(1→3)-β-D-glucuronopyranosyl-olean-12-en-28-*O*-β-D-glucopyranosyl ester, respectively.

Keywords □ *Kochia scoparia*, Chenopodiaceae, two new oleanolic acid saponins, monodesmoside, bidesmoside, *Kochiae* fructus.

Kochiae fructus (Zi Bu Za) has been primarily used in the treatment of gonorrhoea and dermatitis, and as a diuretic¹⁾. The isolation and characterization of *Kochia* spp. saponins have been conducted by several researchers²⁻⁵⁾. However, no detailed chemical investigation of *Kochia scoparia* fructus appears to have been done. The present study deals with the isolation and structural elucidation of two new triterpenoidal saponins from the fructus of *Kochia scoparia* (L.) SCHRAD (Chenopodiaceae).

The water extract of the fructus of *Kochia scoparia* was fractionated on the Amberlite XAD-2 column using 95% MeOH as an eluent. Two new oleanolic acid saponins, compounds 1 and 2 have been isolated by HPLC using reverse phase column.

Compound 1 was obtained as colorless powder, gave positive reactions in Liebermann-Burchard, and Molish test, and showed carboxyl (1690 cm⁻¹) and glycosidic (1,100-1,000 cm⁻¹) absorption band in its IR spectrum. Acid hydrolysis of 1 afforded 1a as the sapogenin, and glucuronic acid and ribose at the molar ratio of 1:1 as a sugar moiety. Compound 1a was identified as oleanolic acid by direct comparison of its physicochemical constants with those of authentic oleanolic acid⁶⁻¹⁴⁾.

The ¹H-NMR spectrum of 1 showed seven quaternary methyl signals at δ 0.8-1.1 (3H, s, CH₃×7) with two anomeric proton signals at 4.3 and 4.7 ppm.

The ¹³C-NMR analysis (Table I) of 1 showed two anomeric carbon signals at 104.7, 104.1 ppm and two carboxyl carbon signals at 180.2 (C-28), 173.8 (glucuronic) ppm. The glucuronic acid and ribose units in 1 were suggested to be linked to the 3-C hydroxyl group of 1a by comparison with ¹³C-NMR spectral data of 1. Compound 1 was methylated according to the method of Hakomori to give permethylate whose anomeric proton signals were observed at 4.36 and 4.72 ppm.

Partial hydrolysis of 1 afforded 1b with ribose, and 1b gave oleanolic acid and glucuronic acid by acid hydrolysis. The mode of linkage of the glucuronic acid and ribose units were examined on the basis of the coupling constants (d, *J*=3.9 Hz, *J*=8.0 Hz) of the anomeric proton signals in the ¹H-NMR spectrum of 1.

Based on these evidences, the structure of 1 was elucidated as oleanolic acid 3-*O*-β-D-ribofuranosyl-(1→2)-β-D-glucuronopyranoside.

Compound 2 was obtained as colorless powder, gave positive reactions in Liebermann-Burchard, and Molish test, and showed ester (1730 cm⁻¹) and glycosidic (1,100-1,100 cm⁻¹) absorption band in its IR spectrum. The ¹H-NMR of 2 showed seven quaternary methyl signals at 0.82-1.30 ppm and three anomeric proton signals at 5.03, 5.41, 6.34 ppm, respectively. The ¹³C-NMR spectrum (Table I) of 2 showed three anomeric carbon signals at δ 106.3 (28-C, in-

Table I. ^{13}C -NMR chemical shifts (δ) of compounds **1** and **2** in $\text{C}_5\text{D}_5\text{N}$

Carbon No.	Oleanolic acid	1	2	C-3 sugar	1	2	C-28 sugar	2
3	78.1	88.2	89.3	GlcUA-1	104.7	106.3	Glc-1	95.8
12	122.6	121.3	122.9	GlcUA-2	85.0	74.2	Glu-2	74.2
13	144.9	143.8	144.2	GlcUA-3	76.0 ^a	86.4 ^a	Glu-3	79.4 ^a
23	28.8	27.4	28.3	GlcUA-4	73.1	71.4	Glu-4	71.2
24	16.0	16.3	17.0	GlcUA-5	74.2 ^a	78.2 ^a	Glu-5	78.9 ^a
28	180.2	180.0	176.5	GlcUA-6	173.8	172.2	Glu-6	62.3
				Rib-1	104.1			
				Rib-2	73.4			
				Rib-3	70.2 ^a			
				Rib-4	69.3			
				Rib-5	65.8			
				Xyl-1		106.9		
				Xyl-2		74.4		
				Xyl-3		77.6 ^a		
				Xyl-4		71.1		
				Xyl-5		67.4		

^{a,b}may be reversed.

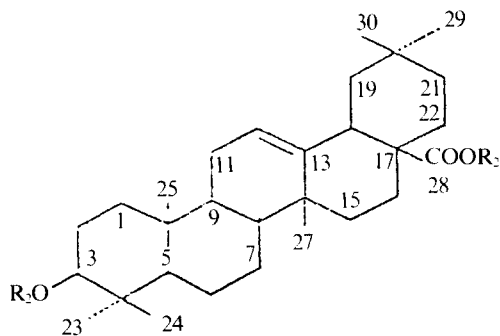
GlcUA: β -D-glucuronic acid, Glu: β -D-glucopyranose

ner carboxyl carbon) signals at δ 172.2 (3-C, inner glucuronic acid), 176.5 (28-C, carboxyl). Acid hydrolysis of **2** afforded oleanolic acid as the sapogenin and glucuronic acid, xylose and glucose at the molar ratio of 1:1:1 as a sugar moiety.

Alkali saponification of **2** gave **2a** and glucose. Compound **2a**, colorless powder gave positive reactions in Liebermann-Burchard, and Molish test, and showed carboxyl residues ($1725, 1655\text{ cm}^{-1}$) glycosidic ($1,100\text{--}1,000\text{ cm}^{-1}$) absorption band in its IR spectrum. The ^1H -NMR spectrum of **2a** showed seven quaternary methyl signals at δ 0.8–1.1 with two anomeric proton signals at 4.3, 4.77 ppm, respectively. ^{13}C -NMR analysis (Table I) of **2a** showed two anomeric carbon signals at 104.7, 104.1 ppm and two carboxyl signals at δ 180.2 and 173 ppm.

Acid hydrolysis of **2a** afforded oleanolic acid as the sapogenin, and glucuronic acid and xylose at the molar ratio of 1:1 as a sugar moiety. The glycosylation shift around 3-C as well as two anomeric carbon signals at 106.3 and 106.9 ppm in the ^{13}C -NMR spectrum of **2a** indicates that **2a** is 3-*O*-glycoside of oleanolic acid which has two monosaccharide units.

Negative FAB-Mass of **2a**, ion at m/z 763 (M^+), 631 (M^+ -xylose), 455 (M^+ -xylose+glucuronic acid)



	R ₁	R ₂
1.	-GlcUA—Rib	H
2.	-GlcUA—Xyl	Glc
1a.	H	H
2a.	-GlcUA—Xyl	H

indicates that sugar moiety of **2a** is consisted of a linear glycosyl unit with terminal xylose and inner glucuronic acid.

The structural correlativity between **2a** and **1** confirms that the terminal moiety of ribose in **1** is replaced by xylose in **2a** by comparing the physical constants and spectral data of these two compounds.

The aqueous layer obtained from the alkali hy-

drolysis of **2** was acidified¹⁶⁾ and analyzed by GC and TLC, showing the presence of glucose.

These experimental results suggest that **2** is an ester composed of **2a** as the acid part, and glucose as the alcohol part.

The signal at 6.3 ppm (1H, d, $J=8.1$ Hz) in ¹H-NMR spectrum of **2** can be attributed to the anomeric proton of glucose linked to the 28-carboxyl group of **2a** in the ester form. The signal of the anomeric carbon of the glucose was observed at 95.8 ppm, supporting the glucose was linked to C-28 as an ester form.

Based on the above results, the structure of **2** was elucidated as 3-*O*-β-xylopyranosyl-(1→3)-β-D-glucuronopyranosyl-olean-12-en-38-*O*-β-glucopyranosyl ester.

EXPERIMENTAL METHODS

General procedure

Acid hydrolysis of **1** and **2** was carried out by refluxing with 2N-HCl:dioxane (1:1) at 90°C for 4 h and partial hydrolysis was performed by heating in the sealed tube with 1.5% H₂SO₄ at 70°C for 7 h. Alkaline saponification of **2** was carried out using 0.05 N KOH in MeOH for 1 h. Acid or alkali in the hydrolyzed solution was neutralized with Amberites MB 3. Melting points were measured with Mitamura Riken Kogyo MELT TEMP and uncorrected. IR spectra were taken by Shimadzu IR 408, 435. GC was performed on Shimadzu GC-9A gas chromatograph. ¹H-NMR spectra were measured on a Bruker AM-200, JEOL JNN-GX-500 spectrometer and ¹³C-NMR spectra were measured on a Bruker AM-200, JEOL JNN-GX-500 spectrometer, using tetramethylsilan as an internal standard. Chemical shifts are given in ppm. Mass spectra were recorded on a JMS-DX300 (FAB-Mass), Hewlett Packard 5985B, Finnigan automated gas chromatography spectroscopy (EI-CI, GC-Mass). Elemental analysis was performed by Perkin Elmer 240 EA. Optical rotation was measured with Union automatic digital polarimeter PM-101. Preparative liquid chromatography was carried out on a column of YMC packed column ODS-5, S-5, 120A ODS with Japan TOSOH system. For TLC, silica gel 60 F₂₅₄ (thickness 0.2 mm, Merck) and RP-8 (thickness 0.25 mm, Merck) were used. Solvent system for silica gel column was consisted of CHCl₃:MeOH:H₂O (70:30:

4, v/v/v). Solvent system for HPLC was 73% MeOH (0.05% TFA H₂O).

Extraction and isolation

Air dried fructus of *Kochia scoparia* (1 kg) collected in Seoul, Korea¹⁷⁻¹⁹⁾, was extracted with hot water. The water extract was passed through an Amberlite XAD-2 column and the adsorbed material was eluted with 95% methanol. The methanol eluates were concentrated to dryness to give a crude saponin (30 g), which was passed through a silica gel column employing a solvent system of CHCl₃:MeOH:H₂O (70:30:4). The resulting two fractions showed the presence of four different saponins by TLC. The four different saponins were named as saponins A-D according to their R_f values.

Among them, compounds **1** (R_f 0.55) and **2** (R_f 0.35) were separated by using preparative HPLC according to their t_R (41.4, 20.3).

Saponin B(**1**)-A white powder, mp. 220-222°C [α]_D²⁰ +13.4 (C=0.60, MeOH). *Anal. calcd.* for C₄₁H₆₈O₁₅·2H₂O:C, 61.7; H, 8.5. Found: C, 61.1; H, 8.3. IR ν_{max}^{KBr} cm⁻¹ 3400 (brs, OH), 1727, 1651 (COOH), 1450 (C=C), 1055, 1042 (CHOH). ¹H-NMR (pyridine-d₅): 0.71 (3H, s), 0.74 (3H, s), 0.87 (6H, s), 0.97 (3H, s), 1.09 (3H, s), 1.20 (3H, s), 4.31 (1H, d, anomeric H, ribose), 4.72 (1H, d, anomeric, H, glucuronic acid), 5.15 (brs, 12-H). ¹³C-NMR data are listed in Table I.

Saponin C(**2**)-A white powder, mp. 197-201°C (uncorr.) [α]_D²⁰ +32.5 (C=0.05, MeOH). *Anal. calcd.* for C₄₇H₇₈O₂₀·H₂O:C, 57.6; H, 8.2. Found: C, 58.6; H, 8.1. IR. ν_{max}^{KBr} cm⁻¹ 3400 (brs, OH), 1730 (COO-R), 1635 (C=C), 1100-1000 (CHOH). ¹H-NMR δ : 0.82, 0.89, 0.91, 1.00, 1.01, 1.09, 1.29, 1.30, (each 3H, s), 5.03, 5.41, 6.34, (1H, anomeric H, glucuronic acid, xylose and glucose), 5.42 (s, 12-H). ¹³C-NMR data are listed in Table I.

Aglycone of compound 1 and 2

Compounds **1** and **2** (each 100 mg) were hydrolyzed, respectively by using the method described in general procedure. The hydrolysate was diluted with H₂O and extracted with CHCl₃. The residue from the CHCl₃ extract was chromatographed on silica gel column using an eluting solvent system of CHCl₃:MeOH (7:3). The eluates (from **1** and **2**) were concentrated, respectively, and the resulted residues were recrystallized, respectively in MeOH to give

the same aglycone of **1a**, colorless needle, mp. 307-309°C (uncorr.), which was identified as oleanolic acid by direct comparison with the authentic sample. After being neutralized with Amberlite MB-3, the filtrate was concentrated to a small volume and examined by TLC and GC, to show the presence of glucuronic acid, ribose (t_R , 17.8, 6.8) from **1**, glucuronic acid, xylose and glucose (t_R , 20.5, 9.5, 15.3) from **2**.

Alkali saponification of compound 2

Compound **2** (100 mg) was hydrolyzed by alkali using the method, described in general procedure. The reaction mixture was diluted with H₂O then, neutralized with Amberlite MB-3, and extracted with EtOAc:MeOH (2:1). The organic layer thus obtained was washed with H₂O and concentrated to give **2a**.

Compound **2a**, colorless powder (MeOH), 225-227°C (uncorr) $[\alpha]_D^{20} +15.5$ (0.5, MeOH). *Anal. calcd.* for C₄₁H₆₈O₁₅: C, 61.5; H, 8.2. *Found:* C, 61.1; H, 8.3. IR ν_{max}^{KBr} cm⁻¹ 3400 (OH), 1727, 1651 (COOH), 1450 (C=C), 1055, 1042 (CHOH). ¹H-NMR 0.82, 0.89, 0.92, 1.00, 1.09, 0.29, 1.03 (3H, s, CH₃), 5.02 (1H, d, $J=7.9$, anomeric H), 5.36 (1H, d, $J=7.5$, anomeric H).

Permethylate 3 of compound 1

Compound **1** was methylated by the method of Hakomori¹⁵⁾. Compound **1** (50 mg), DMSO 6.0 ml and NaH 350 mg were reacted with streaming N₂ for 2 h, in the ultrasonicator. After cooling, 30 ml of CH₃I was added and allowed to stand for another 1 h in the ultrasonicator. The reaction mixture was diluted with H₂O and extracted with CHCl₃. The CHCl₃ extract was evaporated and recrystallized from MeOH to give **3** as a colorless needles, mp. 124-126°C (uncorr.).

IR ν_{max}^{KBr} cm⁻¹ 1723, 1745 (COOR), 1460 (C=C), 1096, 1025 (OCH₃). *Anal. calcd.* for C₄₈H₈₀O₁₄: C, 65.5; H, 9.1 *Found:* C, 65.0; H, 9.3. ¹H-NMR (CDCl₃) 0.71, 0.83, 0.88, 0.89, 0.92, 1.01, 1.11 (3H, s, CH₃), 3.25, 3.45, 3.48, 3.60, 3.61 (3H, s, OCH₃), 4.36, 4.72 (1H, d, anomeric H), 5.26 (brs, 12-H).

Methanolysis of compound 3

Twenty mg of **3** was boiled with 8% HCl-MeOH (3 ml) on water bath for 3 h. The hydrolysate was neutralized and the filtrate was evaporated. The me-

thylated sugar was identified as 3,4-dimethyl-6-carboxymethylglucuronopyranoside and 2,3,4-trimethyl-ribopyranoside by GC (t_R , 6.36, 14.73).

LITERATURE CITED

- Choi, M. S.: Korean Folk Medicine, p.167 (1987).
- Coxworth, E. C. M. *et al.*: *Kochia* seed as a component of diet of turkey poult. Effects of different methods saponin removal or inactivation. *Can. J. Anim. Sci.*, **54**, 712 (1972).
- Singh, N. *et al.*: Chemical investigation of *Kochia indica*. *Indian J. Chem.*, **21**, 49 (1966).
- Tandon, J. S. *et al.*: Chemical examination of *Kochia trichophylla*. *Indian J. Chem.* **4**, 545 (1966).
- Kernan, J. A., Coxworth, E., Fleming, S.: Microdetermination of triterpene sapogenin content of *Kochia scoparia* seed using gas-liquid chromatography, *J. Arg. Food Chem.*, **21**, 232 91973).
- Shen, Y. L. *et al.*: Isolation of two saponins from *Anemone flaccida* and effects on reverse transcriptase. *Shoyakugaku Zasshi*, **42**, 35 (1988).
- Shao, C. J. *et al.*: Saponins from leaves of *Acanthopanax senticosus*, *Chem. Pharm. Bull.*, **36**, 601 (1988).
- Kisu, H., *et al.*: Studies on the constituents of *Clematis Spp.* *Chem. Pharm. Bull.* **30**, 859 (1982).
- Shao, C. J. *et al.*: Saponins from leaves of *Acanthopanax senticosus*. *Chem. Pharm. Bull.*, **37**, 42 (1989).
- Tschesche, R. *et al.*: Uber die Saponin des Spinnats. *Liebigs Ann. Chem.*, **726**, 125 (1969).
- Taniyama, T. *et al.*: Saponin and sapogenol from the seed of *Glycine Max.* *Chem. Pharm. Bull.*, **36**, 2829 (1988).
- Kinjo, J. *et al.*: Studies on the constituents of *Pueraria lobata*, *Chem. Pharm. Bull.*, **36**, 1174 (1988).
- Mizutani, K. *et al.*: Saponins from *Anemone Ricularis*, *Planta Medica*, **45**, 327 (1984).
- Shao, C. J. *et al.*: Saponins from *Kalopanax Septemlobus*, *Chem. Pharm. Bull.*, **37**, 311 (1989).
- Hakomori, S. I.: A rapid permethylation of glycolipid and polysaccharide catalyzed by methylsulfanyl carbanion dimethylsulfoxide, *J. Biochem.* **55**, 205 (1964).
- Takabe, S. T. *et al.*: Triterpenoidal glycoside from the root of *Tetrapanax papyriferum* K.

- Koch. III. *Chem. Pharm. Bull.* **33**, 470 (1985). (1986).
17. Ahn, H. S.: Names of plants of Korea, p.32 19. Yook, C. S.: Coloured medicinal plants of Korea, p.174 (1989).
18. Kim, Y. S.: Coloured plants of Korea, p.55