Isoflavonoids of Belamcanda chinensis (II)*

Gang Hae Eu, Won Sick Woo, Ha Sook Chung and Eun Hee Woo Natural Products Research Institute, Seoul National University, Seoul 110-460, Korea

Abstract—Dimethyltectorigenin, irisflorentin, muningin and iristectorigenins A and B were isolated from the rhizomes of *Belamcanda chinensis*.

Keywords—*Belancanda chinensis* • Iridaceae • isoflavonoids • dimethyltectorigenin • irisflorentin • muningin • iristectorigenins A and B

In the previous paper, isolation of tectorigenin, irigenin and their glucosides together with 5,3'-dihydroxy-4',5'-dimethoxy-6,7-methylene-dioxyisoflavone was reported.¹¹ A further study on the chloroform soluble fraction has now led to the isolation of four additional isoflavones (Compounds 6-9). Compound 6, mp 180~182°, Compound 8, mp 284~286° and Compound 9, mp 186~188° showed FeCl₃ positive but Compound 7, mp 166~167° negative.

The UV of each compound, exhibiting band II peak at 264~269nm (Table I) was very similar to those reported for a number of isoflavonoids²⁾ and the IR showed the presence of conjugated carbonyl and aromatic ring system in all.

All compounds showed the hydroxyl group absorption except Compound 7, which showed, however, the methylenedioxyl group absorption.

Two singlet signals for H-2 and H-8 protons of each compound fell in the normal shift region for the isoflavonoid nucleus oxygenated at 5, 6 and 7 positions of ring A (Table II). Compounds 6 and 8 showed two two proton ortho-coupled doublets, indicating the presence of a 4'-substituted B ring, Compound 7 showed a two proton singlet, indicating a symmetric

Table I. UV spectral data of compounds $[\lambda_{\max}^{MeOH}]$ nm (log ϵ)]

(10g ε)]		
6	8	9
266.5(4.21)	263.5(4.39)	269. 5(4. 73)
330.0(3.24)	325.0(3.75)	331.0(4.66)
272.0(4.08)	283.5(4.38)	280.5(4.63)
358.5(3.28)	372.5(3.69)	333.0(4.53)
266.5(4.21)	263.5(4.37)	275.5(4.73)
330.0(3.24)	323.5(3.68)	342.0(4.36)
267.0(4.57)	264.0(4.39)	272.0(4.71)
330.0(3.24)	325.0(3.76)	342.0(3.95)
276.5(4.13)	264.0(4.36)	278.5(4.72)
312.0(3.63)	324.5(3.78)	318.2(4.26)
374.0(3.27)		366.4(3.85)
278.5(4.16)	264.0(4.32)	279.5(4.74)
316.5(3.52)	333.0(3.76)	318.0(4.27)
374.0(3.27)		374. 4(3. 88)
	6 266. 5(4. 21) 330. 0(3. 24) 272. 0(4. 08) 358. 5(3. 28) 266. 5(4. 21) 330. 0(3. 24) 267. 0(4. 57) 330. 0(3. 24) 276. 5(4. 13) 312. 0(3. 63) 374. 0(3. 27) 278. 5(4. 16) 316. 5(3. 52)	6 8 266. 5(4. 21) 263. 5(4. 39) 330. 0(3. 24) 325. 0(3. 75) 272. 0(4. 08) 283. 5(4. 38) 358. 5(3. 28) 372. 5(3. 69) 266. 5(4. 21) 263. 5(4. 37) 330. 0(3. 24) 323. 5(3. 68) 267. 0(4. 57) 264. 0(4. 39) 330. 0(3. 24) 325. 0(3. 76) 276. 5(4. 13) 264. 0(4. 36) 312. 0(3. 63) 324. 5(3. 78) 374. 0(3. 27) 278. 5(4. 16) 264. 0(4. 32) 316. 5(3. 52) 333. 0(3. 76)

trisubstituted B ring, Compound 9 showed an one proton ortho-coupled doublet, one proton meta-coupled doublet and one proton double doublet, indicating a trisubstituted B ring. Each NMR revealed the presence of three methoxyl group in Compound 6, four methoxyl group and a methylenedioxyl group in Compound 7, two methoxyl group in both Compound 8 and Compound 9, respectively. The appearance of a pair of retro-Diels-Alder fragment ions at

^{*} For previous report see ref. 1.

Table II. ¹H-NMR data of compounds (80 MHz)

Proton No.	6 (CDCl ₃)	PM 6 (CDCl ₃)	7 (CDCl ₃)	8 (DMSO-d ₆)	9 (DMSO-d ₆)
H-2	7.87(1H, s)	7.78(1H, s)	7.78(1H, s)	8.16(1H, s)	8.34(1H, s)
H-8	6.45(1H, s)	6.67(1H, s)	6.63(1H, s)	6.73(1H, s)	6.49(1H, s)
H-2'(H-6')	6.97(2H, d) J=8.8	6.94(2H, d) J=8.8	6.75(2H, s)	6.88(2H, d) J=8.8	7.14(1H, d) $J=1.8$
H-3′(H-5′)	7.46(2H, d) J=8.8	7.46(2H, d) J=8.8		7.33(2H, d) J=8.8	
H-5′					6.81(1H, d) J=8.1
H-6′					7.01(1H, dd) J=1.8, 8.1
H-5-OH	12.78(1H, s)				13.02(1H, s)
Other OH				9.40(bs)	9.02(bs)
$-OCH_3$	3.83(3H, s)	3.81(3H, s)	3.87(9H, s)	3.73(3H, s)	3.75(3H, s)
	3.91(3H, s)	3.89(3H, s)	4.08(3H, s)	3.90(3H, s)	3.79(3H, s)
	3.94(3H, s)	3.94(6H, s)			
$-O$ CH_2 $-O$			6.06(2H, s)		

m/z 196 and 132, 194 and 195, 196 and 118 and 182 and 148 in each MS spectrum, respectively, indicated that two methoxyls and one hydroxyl were located on ring A and one methoxyl on ring B in Compound 6, one methylenedioxyl and one methoxyl on ring A and three methoxyls on ring B in Compound 7, two methoxyls and one hydroxyl on ring A and one hydroxyl on ring B in Compound 8 and one methoxyl and two hydroxyl on ring A and one methoxyl and one hydroxyl on ring B in Compound 9.

In Compound 6, a bathochromic shift of band II in UV with AlCl₃ and appearance of an exchangeable proton signal around δ13 ppm indicated that 5-hydroxy group was not substituted, therefore it was identified as 5-hydroxy-6, 7, 4'-trimethoxyisoflavone. Methylation of Compound 6 with dimethylsulfate gave the same product as 5, 6, 7, 4'-tetramethoxyisoflavone (PM 6) prepared from tectorigenin. Compound 6 was isolated first from *Dalbergia sissoo*.³⁾

In Compound 7, its UV spectrum was not

changed in the presence of shift reagents. Two methoxyl carbon signals appeared around δ 60 ppm, indicating the presence of two *ortho*-disubstituted methoxyl groups, which were allocated to C-5 and C-4' positions, respectively. Therefore it was characterized as 6, 7-methylenedioxy-5, 3', 4', 5'-tetramethoxyisoflavone. As expected, methylation of 5, 3'-dihydroxy-4', 5'-dimethoxy-6, 7-methylenedioxyisoflavone gave Compound 7. Compound 7 (irisflorentin) was first isolated *Iris florentina*.

In Compound 8, a bathochromic shift of band I on addition of NaOH was observed but no band shift in the presence of AlCl₃ and NaOAc, indicating that hydroxyl groups at C-5 and C-7 were methylated, therefore it was characterized as 6, 4'-dihydroxy-5, 7-dimethoxy-isoflavone, and confirmed by derivatization into permethylated tectorigenin (PM 6). Compound 8 (muningin) was isolated previously from *Pterocarpus angolensis*.⁵⁾

In Compound 9, a bathochromic shift of band II in the presence of AlCl₃ and NaOAc and

increase of intensity of band I in the presence of NaOMe, indicating the presence of 5-hydroxyl, 7-hydroxyl and 4'-hydroxyl groups. Therefore, Compound 9 was assumed to be 5,7,4'-trihydroxy-6,3'-dimethoxyisoflavone (iristectorigenin B, 9b). However, the melting point of Compound 9 compared badly with that reported the authors.⁶⁾ In order to explain this dissimilarity, Compound 9 was subjected to degradation. Oxidation with H₂O₂ resulted in formation of both vanillic acid and isovanillic acid. Thus Compound 9 was concluded to be a mixture of iristectorigenin A(9a) and B(9b), which were first isolated from *Iris tectorum*.^{6,7)}

¹³C-chemical shifts (Table III) of each compound are in agreement with the values for each structure. The assignments were based

Table III. ¹³C-NMR chemical shifts of compounds (20 MHz)

Carbon No.	(CDCl ₃)	PM 6 (CDCl ₃)	7 (CDCl ₃)	(DMSO-d ₆)
2	152.55	150.34	152.85	154.3
3	123.50	125, 41	125.57	122.0
4	181.06	174.94	174.94	180.7
5	153.54	153.09	141.81	152.9
6	131.90	140.70	135.57	131.8
7	158.99	157.74	154.82	157.7
8	90.47	96.14	93.10	94.1
9	153.70	154.56	150.65	153.4
10	106.77	113. 14	113.79	105.1
1'	123.04	124.41	127.40	122.1
2'	130.09	130.20	107.20	113.9
3'	114.21	113.92	153.16	147.6
4'	159.86	159.62	138.62	147.1
5 ′	114.21	113.92	153.16	115.6
6 ′	130.09	130. 20	107.20	122.0
OCH_3	60.69	61.97	61.05	60.1
		61.30	60.69	
	56.25	56.17	56.35	56.2
	55. 32	55. 26	56.35	
$-O$ CH_2			102.19	

on shift comparison with literature data. 8,9) The early assignments 1) for C-7 and C-9 in all the compounds previously reported have to be reversed. Original incorrect assignments were purely based on those of the reference 10.

	R_1	R_2	R_3	R_4	R_5	R_6
6	H	CH ₃	CH ₃	Н	CH_3	Н
PM 6	CH_3	CH_3	CH_3	H	CH_3	Н
7	CH_3	C	H_2 —	OCH_3	CH_3	OCH ₃
8	CH_3	H	CH_3	H	H	H
9a	\mathbf{H}	CH_3	H	OH	CH_3	H
9 b	H	CH_3	Н	OCH_3	H	H

Experimental

Separation of compounds

The CHCl₃ soluble fraction of the MeOH extract was chromatographed over SiO₂ column and eluted with a gradient of CHCl₃-MeOH to afford fractions, but most of the fractions consisted of mixtures. Among them the fraction containing Compounds 6-9 was subjected to SiO₂ column chromatography using hexane-EtOAc (gradient) to give each compound.

Characterization of compounds

5-Hydroxy-6, 7, 4'-trimethoxyisoflavone

(Compound 6)—crystallized from MeOH as pale yellow needles, mp. 180~182°; Rf, 0.18 (hexane-EtOAc=7:3, si gel); IR $\nu_{\rm max}^{\rm KBr}({\rm cm}^{-1})$: 3430 (OH), 1665 (conjugated C=O), 258, 1460, 1520 (aromatic); MS m/z (rel. int.): 328 (M+, 100), 196 (RDA with A ring, 2.0), 132 (RDA with B ring); UV, NMR; see Tables I, II and III.

5, 3', 4', 5'-Tetramethoxy-6, 7-methylenedioxyisoflavone (Compound 7)—crystallized from MeOH as needles, mp. $166\sim167^\circ$; Rf, 0.1 (hexane-EtOAc=6:4 si gel); UV $\lambda_{\rm max}^{\rm MeOH}$ nm (logs): 266.5(4.44), 323.5(3.80), unchanged by shift reagents; IR $\nu_{\rm max}^{\rm KBr}$ (cm⁻¹): 1668 (conjugated C=O), 1585 (conjugated C=C), 929 (methylenedioxy); MS m/z (rel. int.):386 (M+, 100), 194 (RDA with A ring, 3.0), 195 (RDA with B ring, 2.6); NMR: see Tables II and III.

6, 4'-Dihydroxy-5, 7-dimethoxyisoflavone

(Compound 8)—crystallized from MeOH as white needles, mp. $284\sim286^{\circ}$; Rf, 0. 23 (CHCl₃-MeOH=94:6, si gel); IR $\nu_{\rm max}^{\rm KBr}$ (cm⁻¹): 3400 (OH), 1630 (conjugated C=O), 1450, 1525 (aromatic); MS m/z (rel. int.): 314 (M+, 56.2), 196 (RDA with A ring, 5.3), 118 (RDA with B ring, 70.0); UV, NMR: see Tables I and II.

5, 7, 4'-Trihydroxy-6, 3'-dimethoxyisoflavone (Compound 9)—crystallized from MeOH as yellow needles, mp. $186 \sim 188^{\circ}$; Rf, 0. 33 (CHCl₃-MeOH=94:6, si gel); IR $\nu_{\text{max}}^{\text{KBr}}(\text{cm}^{-1})$: 3430(OH), 1630 (conjugated C=O), 1590, 1530, 1470 (aromatic); MS m/z (rel. int.): 330 (M+, 100), 182(RDA with A ring, 0.9), 148 (RDA with B ring, 9.0); UV, NMR: see Tables I, II and III.

Methylation of Compound 6 and Compound 8

To a solution of each compound (30 mg) in acetone (10 ml) was added (CH₃)₂SO₄ (1.0 ml) and K₂CO₃ (2.4 g), and stirred for 5 hr at 50°. After diluted with water the reaction mixture was extracted with ether. The ether layer was washed with water, dried with Na₂SO₄, evaporated and applied to column chromatography using hexane–EtOAc (7:3) to obtain pure permethylated tectorigenin (PM 6); mp. 176~178°; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ε): 261.0(4.54), 304.5 (3.84), unchanged by shift reagents; MS m/z

(rel. int.): 342 (M⁺, 406), 327 (M-CH₃, 100), 210 (RDA with A ring, 4.9), 132 (RDA with B ring, 59.5); NMR: see Tables II and III.

Preparation of Compound 7

Methylation of the authentic sample of 5, 3'-dihydroxy-4', 5'-dimethoxy-6, 7-methylene-dioxyisoflavone as described above gave a single product, which was in every respect (mp, MS, TLC and ¹HNMR) the same as Compound 7.

Oxidation of Compound 9

To a solution of Compound 9 (2 mg) in 5% KOH (3 ml) was added 3% H₂O₂ (1 ml) and the mixture was allowed to stand over night.

After decomposition of excess H_2O_2 with MnO_2 , the reaction mixture was acidified with dilute HCl, extracted with ether and submitted to TLC to detect vanillic acid and isovanillic acid. Rf, 0.25 and 0.18 (C_6H_6 -EtOAc-HOAc= 18:1:1); 0.42 and 0.32 (C_6H_6 -HOAc-H₂O= 6:7:3); 0.27 and 0.20 (toluene-formic acidethylformate=5:4:1); Color reaction with Gibbs reagent followed by NH₃ fuming: pink and blue, respectively.

Acknowledgements—This work was supported in part by the research grant from KOSEF.

(Received May. 8, 1990;

Accepted Sept. 31, 1990

Literature Cited

- Lee, S.O., Woo, W.S., Woo, E.H. and Kim, K.S.: Kor. J. Pharmacogn. 20, 219 (1989).
- 2. Woo, W.S.: Techniques in natural products research, Min Eum Sa, Seoul (1984).
- Banerji, A., Murti, V.V.S. and Seshadri, T.R.: Curr. Sci. 34, 431 (1965).
- Morita, N., Arisawa, M., Kondo, Y. and Takemoto, T.: Chem. Pharm. Bull. 21, 600 (1973).
- King, F.E., King, T.J. and Warwick, A.J.: J. Chem. Soc. 96 (1952).
- 6. Morita, N., Shimokoriyama, M., Shimizu, M. and Arisawa, M.: Yakugaku Zasshi 92, 1052

Vol. 22, No. 1, 1991

(1972).

- Morita, N., Shimokoriyama, M., Shimizu, M. and Arisawa, M.: Chem. Pharm. Bull. 20, 730 (1972).
- Markham, K.R. and Chari, V.M.: Carbon-13 NMR spectroscopy of flavonoids. In "The Flavonoids. Advances in Research" (Eds. Har-
- borne, J.B. and J.B. and Mabry, T.J.), Chapman and Hall, London, p. 19 (1982).
- Agrawal, P.K. and Bansel, M.C.: Isoflavonoids. In "Carbon-13-NMR of flavonoids" (Ed. Agrawal, P.K.) Elsevier, Amsterdam, p. 183 (1989).
- Shawl, A.S., Dar, B.A. and Vishwapaul: J. Nat. Prod. 48, 849 (1985).