

Studies on the Plant Pathogenic Corynebacteria; The Synthesis of B Group Vitamins by Plant Pathogenic Bacteria

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Corynebacterium 屬 植物病原細菌에 관한 研究

植物病原細菌의 Vitamin B 群의 合成

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Abstract

The results of studies on the synthesis of B group vitamins by plant pathogenic bacteria indicate that most bacteria utilize thiamine, nicotinic acid, biotin and P-Aminobenzoic acid as growth factors.

Riboflavin (vitamin B₂) was produced by most bacterial genera including the Corynebacteria but with the exception of *C. rathay* and *C. fascians*. The results suggest that the ability to produce riboflavin is not a generic characteristic of *Corynebacterium*, and that the accuracy of the ultra-violet light method (one of the diagnostic tests for potato bacterial ring rot disease caused by *Corynebacterium sepedonicum*) must be reconsidered.

Introduction

There are many reports on the vitamin requirements of phytopathogenic Corynebacteria. Mueller⁸⁾ concluded that the nonexacting strains of *C. diphtheriae* required no accessory factors whereas the exacting strains required biotin, nicotinic acid, and pantothenic acid, or B-alanine for normal growth. Starr¹⁴⁾ found that most of the plant pathogenic Corynebacteria required thiamin, biotin, and nicotinic acid. Mohanty⁷⁾ noted the stimulatory effect of thiamin on *C. fascians*. McLachlan and Thatcher found that *G. sepedonicum* is able to grow in a vitamin-free medium and stated that only pantothenic acid is essential for growth since the synthesis of pantothenic acid is a limiting factor of the growth rate of the organism. Ramamulthi¹⁰⁾ reported that thiamin is essential to nearly all plant pathogenic Corynebacteria, and pantothenic acid is essential to *C. flaccumfaciens*. He also

stated that biotin and niacin are required by *C. fascians* and *C. michiganense*.

The ultraviolet light method is one of the diagnostic tests for potato ring rot caused by *C. sepedonicum*. It was first developed by Iverson, and Kelly⁴⁾ and is based on the presence of a byproduct of bacterial metabolism which emits a green fluorescent glow under ultraviolet light. This method has become a standard test in potato ring rot control programmes. Skaptason¹³⁾ reported that the byproduct of *C. sepedonicum* which fluoresced under ultraviolet light was riboflavin. It has also been reported from *C. diphtheria* (the type species of *Corynebacterium*) by Evans, Hardley, and Happedol²⁾ but studies have never been carried out on the synthesis of riboflavin by other plant pathogenic Corynebacteria.

On the basis of the above information, the authors considered that studies on vitamin B group synthesis by plant pathogenic bacteria were needed in order to

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reconfirm the accuracy of the ultra-violet test as a method of detecting infected potato tubers and to investigate the specific vitamin requirements of phytopathogenic bacteria.

Materials and Methods

(a) Test organisms

Table 1 shows the species and strains of bacteria used in the study. Thirty-eight species of bacteria were assayed for production and utilization of B vitamins^{1,3,9,15} Ten species of Corynebacteria as the test organisms and 28 other species belong in each genera of Pseudomonas, Erwinia, Xanthomonas, Agrobacterium, Bacillus and Aerobacter as the control were sampled.

Table 2 shows the species of lactic acid bacteria used for the microbioassay of B vitamins.

Table 1. Genera and species of bacteria assayed for synthesis and utilization of B group vitamins

Genus	Species
<i>Corynebacterium</i>	<i>insidiosum</i>
	<i>sepedonicum</i>
	<i>michiganense</i> (2 strains)
	<i>rathay</i>
	<i>fascians</i> (2 strains)
	<i>poinsettiae</i>
	<i>tritici</i>
	<i>flaccumfaciens</i>
	<i>vesiculare</i>
	<i>oortii</i> (3 strains)
	<i>Pseudomonas</i>
<i>marginalis</i>	
<i>syringae</i>	
<i>lachrymans</i> (4 strains)	
<i>coronafaciens</i>	
<i>conjac</i>	
<i>tabaci</i>	
<i>putida</i>	
<i>Erwinia</i>	<i>rodigiousus</i>
	<i>oryzicola</i>
	<i>schuykilliensis</i>
	<i>mori</i>
	<i>caryophylli</i>
	<i>aroideae</i>

milletiae
carotovora
herbicola

<i>Xanthomonas</i>	<i>pruni</i> (2 strains) <i>oryzae</i> (2 strains) <i>citri</i> <i>hyacinthi</i> <i>vesicatoria</i>
<i>Agrobacterium</i>	<i>tumefaciens</i> <i>radiobacter</i>
<i>Bacillus</i>	<i>subtilis</i> <i>natto</i> <i>megaterium</i>
<i>Aerobacter</i>	<i>aerogenes</i>

Table 2. Test organisms used for microbioassay of B group vitamins

Test organisms	Vitamins
<i>Lactobacillus fermenti</i> 36 ATCC 9338 (IAM 1083)	Thiamine
<i>Lactobacillus casei</i> ATCC 7469	Riboflavin Pyridoxal Folic acid
<i>Lactobacillus arabinosus</i> ATCC 8014 (IAM 1084)	Nicotinic acid Pantothenic acid Biotin P-Aminobenzoic acid

(b) Strain maintenance^{5,9)}

The strains used for the assay of B-vitamins were maintained in Snieszko and Bond's medium; Bacto peptone 0.3%, bacto tryptose 0.3%, bacto yeast extract 0.3%, dextrose 0.3% (percentages are W/V). The pH was adjusted to 7.0 with normal NaOH. This medium supported good growth of all the strains. The lactic bacteria were top cultured at 37°C for 20 hours on a medium containing bacto yeast 2%, proteose pepton 0.5%, dextrose 1%, KH₂PO₄ 0.2% and were then maintained in a refrigerator at 5°C. They were recultured once every 3 days on the above medium.

(c) **Assay test**¹¹⁾

The test organisms were shake cultured at 160rpm at 25°C for 10 days in Snieszko and Bond's broth. The free type B-vitamins were then extracted by heat treatment at 80°C for 30 minutes.

(d) **Microbioassay of bacteria**^{59,12)}

A uniform medium for microbiological determination of the B-vitamins with various lactic acid bacteria (Table 3) was employed for the test. The double strength medium was added (5ml per test tube) to test tubes containing 5ml each of dilute standard vitamins

or test materials. The tubes were then autoclaved at 10 p.s.i. for 5min, inoculated with the lactic acid bacteria and washed 3 times with 0.85% NaCl solution with the rotor running at 3,000rpm for 10min. After the inoculated microbioassay media had been incubated at 35°C for 20 hours, they were sterilized at 100°C for 20 hours, and the turbidity was then measured at 640 mu. The quantities of Bgroup vitamins were calculated from standard curves prepared from the microbioassay of the standard vitamin solutions.

Table 3. Double concentration basal medium for microbioassay of B group vitamins

Vitamins		Amino acids	
Thiamine-HCl	1 mg	Casein hydrolysate	10 g
Riboflavin	1 mg	L-Tryptophane	400 mg
Nicotinic acid	2 mg	L-Cysteine-HCl	1.6 g
Biotin	8 r	L-Asparagine	200 mg
Ca-pantothenate	1 mg	L-Tyrosine	400 mg
P-Aminobenzoic acid	200 r	Minerals	
Pyridoxine-HCl	400 r	KH ₂ PO ₄	6 g
Pyridoxal-HCl	1 mg	MgSO ₄ ·7H ₂ O	400 mg
Pyridoxamine-2HCl	400 r	FeSO ₄ ·7H ₂ O	20 mg
Folic acid	20 r	MnSO ₄ ·4H ₂ O	20 mg
Bases		NaCl	20 mg
Adenine sulfate	20 mg	Others	
Guanine-HCl	20 mg	Na-acetate	40 g
Xanthine	20 mg	Glucose	40 g
Uracil	20 mg		

(Amounts per 1 liter of double concentration medium, pH 6.5-6.8)

Table 4. Preparation of the standard B-vitamins

Vitamins	Solvent	Concentration/ml
B ₁ -HCl	0.02N acetic acid	200 r
B ₂	0.3% acetic acid	50 r
NiA	0.02N acetic acid	200 r
Biotin	0.02N acetic acid	8 r
Ca-paA	25% ethanol	200 r
PABA	25% ethanol	40 r
PIN-HCl	25% ethanol	100 r
PAM-HCl	25% ethanol	100 r
PAL-HCl	25% ethanol	100 r
FA	25% ethanol	10 r

Results and Discussion

Results for the tests of 48 cultures of bacteria, representing 39 species classified in 8 genera, are presented in table 5 and 6. The quantities of B group vitamins are tabulated as μ g of free type vitamin per 1ml of Snieszko and Bond's medium. The values obtained by microbiological assay of the cultured media with various bacteria accurately indicate the B group vitamins synthesized by the bacteria and released into the medium after heat treatment.

Table 5. The synthesis of B group vitamins by bacteria

Organism	B-vitamin(mr per 1ml of medium)			
	B ₁	B ₂	Pal	NiA
<i>Corynebacterium</i>				
<i>insidiosum</i>	18.0	450.0	40.0	0
<i>sepedonicum</i>	51.0	1150.0	74.0	-1300
<i>michiganense</i> ¹⁾	7.0	100.0	42.0	- 400
<i>michiganense</i> ²⁾	- 34.0	500.0	47.0	-1100
<i>rathay</i>	4.0	-100.0	49.0	0
<i>fascians</i> ¹⁾	- 21.0	-400.0	40.0	1400
<i>fascians</i> ²⁾	- 35.0	0	47.0	-1400
<i>poinsettiae</i>	- 72.0	200.0	47.0	- 600
<i>tritici</i>	16.5	0	40.0	0
<i>flaccumfaciens</i>	626.0	450.0	257.0	1000
<i>vesiculare</i>	- 14.0	300.0	207.0	2100
<i>oortii</i> ¹⁾	1776.0	500.0	9.5	- 450
<i>oortii</i> ²⁾	976.0	550.0	9.5	- 500
<i>oortii</i> ³⁾	1256.0	100.0	9.5	- 400
<i>Pseudomonas</i>				
<i>eriotryae</i>	14.0	100.0	162.0	300
<i>marginalis</i>	4.0	800.0	81.0	1800
<i>syringae</i>	- 28.5	550.0	167.0	7000
<i>lachrymans</i> ¹⁾	2.5	450.0	25.0	800
<i>lachrymans</i> ²⁾	- 36.0	1200.0	122.0	3650
<i>lachrymans</i> ³⁾	- 37.5	1300.0	87.0	4100
<i>lachrymans</i> ⁴⁾	- 1.0	750.0	42.0	4250
<i>coronafaciens</i>	- 16.0	1700.0	25.0	3250
<i>conjac</i>	24.5	-500.0	65.5	500
<i>tabaci</i>	17.5	0	71.0	11100
<i>putida</i>	5.5	1950.0	172.0	0
<i>rodigiousus</i>	- 8.0	1350.0	73.0	-1600
<i>oryzicola</i>	976.0	350.0	112.0	3100
<i>schuyllkilliensis</i>	426.0	250.0	167.0	2750
<i>mori</i>	31.0	-150.0	74.0	2450
<i>caryophylli</i>	22.0	550.0	32.0	2600
<i>Erwinia aroideae</i>				
<i>milletiae</i>	- 28.5	50.5	900.0	0
<i>herbicola</i>	8.0	41.0	50.0	- 850
<i>carotovora</i>	- 15.5	25.0	500.0	0
<i>Xanthomonas pruni</i> ¹⁾				
<i>pruni</i> ²⁾	- 18.0	—	—	—
<i>oryzae</i> ¹⁾	4.0	44.0	500.0	650
<i>oryzae</i> ²⁾	- 4.0	48.5	500.0	1200
<i>citri</i>	- 64.0	—	—	—

<i>hyacinthi</i>	26.0	47.0	0	0
<i>vesicatoria</i>	18.0	54.0	—	1200
<i>Agrobacterium</i>				
<i>tumefaciens</i>	— 35.0	282.0	1650.0	350
<i>radiobactor</i>	— 52.5	47.0	1150.0	250
<i>Bacillus subtilis</i>	— 35.0	162.0	—750.0	— 300
<i>natto</i>	11.5	14.0	0	— 300
<i>megaterium</i>	14.0	25.0	100.0	— 600
<i>Aerobacter aerogenes</i>	— 6.5	42.0	350.0	0

Table 6. The synthesis of B group vitamins by bacteria

Organism	B-vitamin(mr per 1ml of medium)		
	Ca-PaA	Bio	PABA
<i>Corynebacterium</i>			
<i>insidiosum</i>	19250	6.80	— 260
<i>sepedonicum</i>	1400	4.20	— 120
<i>michiganense</i> ¹⁾	6770	8.60	— 110
<i>michiganense</i> ²⁾	2970	0.20	— 80
<i>rathay</i>	7470	2.80	— 90
<i>fascians</i> ¹⁾	3770	3.90	220
<i>fascians</i> ²⁾	3970	6.60	— 50
<i>poinsettiae</i>	820	8.60	— 220
<i>tritici</i>	12570	1.00	— 120
<i>flaccumfaciens</i>	— 530	4.70	420
<i>vesiculare</i>	12570	—4.18	— 175
<i>oortii</i> ¹⁾	— 930	3.72	— 170
<i>oortii</i> ²⁾	— 830	3.92	— 185
<i>oortii</i> ³⁾	— 840	4.02	— 178
<i>Pseudomonas eriobotryae</i>	11970	0.20	— 160
<i>marginalis</i>	16170	0	— 145
<i>syringae</i>	12170	0	— 110
<i>lachrymans</i> ¹⁾	1970	—1.10	— 220
<i>lachrymans</i> ²⁾	15970	—3.50	— 60
<i>lachrymans</i> ³⁾	15070	—1.80	— 10
<i>lachrymans</i> ⁴⁾	16070	—4.50	— 155
<i>coronafaciens</i>	6370	0.20	— 290
<i>conjac</i>	—1180	—4.80	240
<i>tabaci</i>	10270	—3.93	— 270
<i>putida</i>	5620	—4.40	120
<i>rodigiousus</i>	10570	—4.00	— 165
<i>schuylkilliensis</i>	2470	2.40	— 270
<i>mori</i>	11170	8.10	— 260
<i>caryophylli</i>	—	—	—
<i>Erwinia aroideae</i>	670	—4.90	— 90

<i>milletiae</i>	3870	-4.80	240
<i>carotovora</i>	670	-4.90	20
<i>herbicola</i>	3620	-4.80	- 40
<i>Xanthomonas pruni</i> ¹⁾	6570	1.00	- 245
<i>pruni</i> ²⁾	—	—	—
<i>oryzae</i> ¹⁾	3620	4.52	—
<i>oryzae</i> ²⁾	3320	2.70	- 70
<i>citri</i>	- 140	—	—
<i>hyacinthi</i>	—	19.10	- 40
<i>vesicatoria</i>	—	—	—
<i>Agrobacterium</i>			
<i>tumefaciens</i>	670	0.50	- 250
<i>radiobacter</i>	- 860	22.10	- 340
<i>Bacillus subtilis</i>	9070	18.10	- 300
<i>natto</i>	9770	0.30	- 300
<i>megaterium</i>	- 750	13.10	- 270
<i>Aerobacter aerogens</i>	5470	-3.50	- 290

Among the plant pathogenic *Corynebacteria*, *C. michiganense*, *C. fascians* and *C. poinsettiae* could not synthesize vitamin B₁ (thiamin). About half the species from the other genera also could not synthesize thiamin and particularly those from the genus *Pseudomonas*.

Nicotinic acid, pantothenic acid, P-aminobenzoic acid and biotin were generally utilized by the bacteria tested in this study. The synthesis of nicotinic acid was detected from 3 species of *Corynebacteria*; *C. fascians* (rough type), *C. flaccumfaciens* and *C. vesiculare*, and pantothenic acid from all except *C. poinsettiae*, *C. flaccumfaciens* and *C. cortii*. *C. vesiculare* was the only *Corynebacterium* which reduced the concentration of biotin in the culture medium below that of the standard medium. Most bacteria of other genera utilized biotin (except genus *Xanthomonas*, *Agrobacterium* and *Bacillus*. P-amino benzoic acid was utilized not only by *Corynebacteria* (except of rough type *C. fascians* and *C. flaccumfaciens*) but also by most bacteria of the other genera. This facts indicate that most bacteria require at least one of the following amino acid for their normal growth; B₁, nicotinic acid, pantothenic acid, P-amino-benzoic acid or biotin.

Similar results have been found from previous studies of plant pathogenic *Corynebacteria* by other workers. Stare¹⁴⁾ studied the vitamin requirements of phytopathogenic *Corynebacteria* and showed that most

of them required thiamin, biotin and nicotinic acid. Mohanty⁷⁾ noted the stimulatory effect of thiamin on *C. fascians*. McLachlan and Thatcher⁶⁾ found that *C. sepedonicum* could synthesize all the vitamins required for its nutrition but that the rate of synthesis of pantothenic acid is a factor limiting the rate of growth of the organism. Ramamurthi¹⁰⁾ reported that thiamin is essential for the growth of most plant pathogenic *Corynebacteria*, pantothenic acid in addition to thiamin is essential to *C. flaccumfaciens*. He also found that biotin and niacin are required by *C. fascians* and *C. michiganense*.

Pyridoxal was synthesized by all the bacteria studied except *Bacillus subtilis*.

The ultraviolet light test method is based on the fluorescence under ultraviolet light of a green pigment which is a byproduct of bacterial metabolism. The test is one of the diagnostic tests for potato bacterial ring rot disease caused by *C. sepedonicum* and is used in control programmes for the detection of this disease. Skaptason¹³⁾ reported that this byproduct fluorescent compound was riboflavin. The synthesis of riboflavin by other plant pathogenic *Corynebacteria* has never been studied although its synthesis has been reported from *G. diphtheria*, the type species of *Corynebacterium* (Evans, Hardley and, Happold)²⁾

The present study was carried out to determine whether the synthesis of riboflavin is a generic char-

acteristic of *Corynebacterium* or a specific characteristic of *C. sepedonicum* and *C. diphtheria*. It was found that riboflavin was produced by all bacteria studied with the exception of *C. rathay*, *P. conjae* and *P. mori*. The ability to synthesize riboflavin, therefore, is not a generic characteristic of *Corynebacterium*. The value of the ultra-violet light method as a diagnostic test of potato bacterial ring rot disease must be reassessed since the absence of fluorescence would

indicate that the tubers are free of the disease but presence of fluorescence is not confirmation of the disease since other species of *Corynebacterium*, and indeed of the other genera, also produce riboflavin. Similar results were also obtained by Skaptason in 1942¹³⁾.

In general, it was found that the ability to synthesize B group vitamins by bacteria was a characteristic of the species or of an individual strain of the species and was not related to its classification.

Corynebacterium 屬 植物病原菌에 關한 研究

— 植物病原菌의 Vitamin B 群의 合成 —

적 요

본실험은 *Corynebacterium* 속 식물병원세균에 관한 연구의 일환으로서 *Corynebacterium* 속 식물병원세균의 종 및 속으로서의 특성을 구명함과 동시에 *C. sepedonicum* 이 생성하는 Rumiflabin 에 기초를 둔 ultra-violet method 의 재검토를 위하여 행하여 진 것으로서 vitamin B 群의 정량은 미생물 정량에 의하였다.

대다수의 식물병원세균은 Thiamine (vitamin B₁), Nicotinic acid, Biotin 및 안식향산을 생장물질로서 사용하였다. Riboflavin 은 *corynebacterium* 속 식물 병원세균 (*C. rathay* 및 *C. fascians* 는 제외) 은 물론 타속의 식물병원세균도 생성하는 점으로 *Corynebacterium* 속 식물병원 세균의 특성이 아니라는 점이 구명되었으며 *C. sepedonicum* 에 의하여 발생되는 감자의 윤부병과 청고병과의 감별에 사용되어온 Ultra-violet light 법은 재검토 되어야 한다.

References

1. Biseibutsu handbook henshuiuinkaihen (1964) handbook of microbiology. 1457p. Kibodo.
2. Evans, W.C., F.C. Happold, and W.R.C. Handly, (1939). The nutrition of *C. diphtheriae* (Types *mitis*, *gavis* and *Intermedium*). Britsh. J. Exp. Path. 20:41-48.
3. Fujida, A. (1955). Assay method of vitamin. 754 p. Namkodo.
4. Iverson, V.E. and H.C. Kelly. (1940). Control of bacterial ring rot of potatoes with special reference to the ultraviolet-light method for selecting disease-free seed stock. Montana state Coll. Agric. Exp. Sta., Bulletin 386. 1-15.
5. Iwai, K., O. Okinaka, and H. Yokomizo (1967). An uniform medium for microbiological determina-

tion of the B-vitamins with various lactic acid bacteria. Vitamins (Japan) 35:(5)387-394.

6. MacLachlan, D.S. and F.S. Thatcher. (1951). Studies in the nutrition of *Corynebacterium sepedonicum* (Spiek. and Kott.) Skapt. and Burkh. Canad. J. Bot. 29 246-259.
7. Mohanty, U. (1951). *Corynebacterium fascians* (Tilford) Dowson, its morphology, physiology, nutrition and taxonomic position. Brit. Mycol. Soc. Transac. 34:95-116.
8. Mueller, J.H. (1940). Nutrition of the diphtheria bacillus. Bact. Revs. 4:97-134.
9. Nihon kagakukai (1970). Experimental course of chemistry. 25 (biochemistry III). 406p. Maruzen Ltd.
10. Ramanurthi, C.S. (1959). Comparative studies on some gram positive phytopathogenic bacteria and their relationship to the Corynebacteria. Cornell Univ. Agric. Exp. Sta., Memorir 366. 1-52.
11. Sato, T., A. Maekawa, T. Suzuki, and Y. Sahaishi. (1966). Utilization of hydrocarbons for the formation of riboflavin and its coenzymes in *Eremothecium ashbyii*. Vitamins (Japan). 34(6):542-545.
12. Seibutsu handbook henshuiuinkaihen (1967). Handbook of biochemistry. 1126p. Kobodo.
13. Skaptason, J.B. (1943). Studies on the bacterial ring-rot disease of potatoes. Cornell Univ. Agric. Exp. Sta., Memorir 250. 1-30.
14. Starr, M.P. (1949). The nutrition of phytopathogenic bacteria. III. The gram-positive phytopathogenic *Corynebacterium* species. J. Bact. 57:253-258.
15. Vitamingaku henshuiuinkai (1956). Vitaminology. 885p. Kanehara Ltd.