

## Specific Association of Riboflavin and Penicillin Derivatives in Chloroform Solution

Byung Sul Yu

(Received September 10, 1974)

**Abstract**—From the measurements of infrared and fluorescence spectra riboflavin - 2', 3', 4', 5'-tetraacetate has been found to associate with penicillin-V more than strongly with themselves. They form the 1 : 1 cyclic hydrogen bonded dimer through the imino and the 2-C carbonyl groups of the isoalloxazine ring and the imino group of the penicillin-V residue. Penicillin-V is an effective quencher of the fluorescence of riboflavin tetraacetate. The quenching appears mainly due to the coplanar interaction through hydrogen bonds partly due to the collision interaction with the penicillin ring.

It has been known that flavin deficiency from the use of antibiotics, in particular penicillin, oxytetracycline and chlortetracycline<sup>1-6)</sup>.

Several investigator have previous reported that the mechanism of appearance of riboflavin deficiency caused by antibiotics<sup>2-5)</sup>.

They suggested the inhibition of flavin enzyme is due to interaction between chlortetracycline and riboflavin<sup>7)</sup>.

The use of infrared spectroscopy has become customary for detecting the specific interaction of biologically important molecules in chloroform solutions. The method has been extended to the survey of the interaction of other kinds of molecules like drugs<sup>8-10)</sup>.

This paper will report that some of antibiotics such as penicillins, can inhibit the enzyme due to interaction with riboflavin which might be present in flavoenzyme. The results might help to interpret the nature of previously reported theory<sup>6)</sup>.

### EXPERIMENTAL

**Materials** — Riboflavin - 2', 3', 4', 5'- tetraacetate was prepared by the method described

From the College of Pharmacy, Seoul National University, Seoul, Korea.

previously<sup>7)</sup>. A yellow flaky crystal was obtained (3 mg) (mp 246°C Found; C, 53.95; H, 5.22; N, 10.41%).

Crude 6-phenoxyacetamido penicillinic acid (penicillin-V) was obtained by the addition of hydrochloric acid to commercial sodium salt. Resultant precipitate was recrystallized from hot water.

**Procedures**—For the measurement of infrared spectra the samples were dissolved in chloroform-d (E. Merck Co.) which was purified by passing through alumina gel column.

Infrared spectra were measured with a Perkin Elmer Model 421 ( $3\mu$  region) and Beckmann Model 20 A infrared spectrophotometer. Fused quartz cell ranging 5 mm was used for measurement in the  $3700\text{--}3000\text{ cm}^{-1}$  region and potassium bromide cell ranging 0.2 mm was used for  $1800\text{--}1500\text{ cm}^{-1}$  region. The infrared spectra shown in the figures of this paper were given in the absorbance scale, which was calculated from the absorbed transmission which the aid of the solvent curves as base lines. None deuterated chloroform was used as the solvent for the measurements of visible-ultraviolet and fluorescence spectra. The solvent was purified by distillation and by successive passage through alumina gel column.

Visible-ultraviolet spectra were recorded with Hitachi 124 spectrophotometer. The fluorescence measurements were carried out with a Hitachi 204 fluorescence spectrophotometer equipped with a mercury lamp and a grating monochromator. The activating wavelength was set at  $360\text{ m}\mu$  and the fluorescence was read in the range  $380\text{--}640\text{ m}\mu$ . A fused quartz cell of 1 cm width and 1 cm length with a cap was used.

To avoid self quenching at higher concentration, the solution of riboflavin derivatives should have a concentration less than  $10^{-4}\text{ M}$ .

## RESULTS and DISCUSSION

**Infrared Spectra**—Infrared spectra of the  $0.004\text{ M}$  deuteriochloroform solution of riboflavin tetraacetate, penicillin-V and the same mole mixture of them are given in figure 1.

The spectrum of riboflavin tetraacetate a strong sharp band is observed at  $3380\text{ cm}^{-1}$  and a broad band at  $3200\text{ cm}^{-1}$ . Since both of the bands disappear on the deuterium and methyl substitution at the 3-N position, they are associated with the N-H stretching vibrations. The apparent extinction coefficient of the  $3380\text{ cm}^{-1}$  band increases and that of the  $3200\text{ cm}^{-1}$  band increases and that of the  $3200\text{ cm}^{-1}$  band decreases with dilution of the solution. Therefore the former band is assignable to the nonbonded N-H stretching

vibration and the latter is considered to be related to the bonded N-H stretching vibration as discussed previously<sup>7)</sup>,

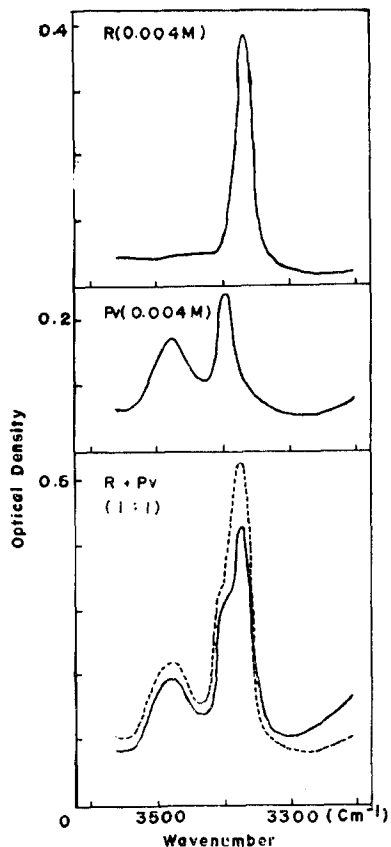


Fig. 1 — Infrared spectra of the deuteriochloroform solutions in a 5 mm quartz cell.

Solid line; observed spectra.

Dotted line; calculated sum of the upper two spectra.

The spectrum of penicillin-V shows two strong bands at  $3475$  and  $3400\text{ cm}^{-1}$  which are respectively due to O-H stretching vibration of the nonbonded carboxylic acid group and nonbonded N-H stretching vibration of imino group.

When both the solutions are mixed together the nonbonded bands decrease in intensity and new association bands appear at  $3300\text{--}3200\text{ cm}^{-1}$  region strong and broad. It is difficult to assign each association band of mixture to one or both components.

In order to get further information on the structure of R-PV complex in solution the spectra in the carbonyl stretching region were studied. In the spectrum of the dilute solu-

tion of riboflavin tetraacetate a strong band is observed at  $1745\text{ cm}^{-1}$  and two bands with medium intensity at  $1710\text{ cm}^{-1}$  and  $1690\text{ cm}^{-1}$  (Figure 2). On the deuteration of the 3-N position the strongest band remains unchanged whereas the  $1710\text{ cm}^{-1}$  band moves to  $1713\text{ cm}^{-1}$  and the  $1690\text{ cm}^{-1}$  band shifts to  $1675\text{ cm}^{-1}$ . The  $1745\text{ cm}^{-1}$  band which appears at such high frequency region must be the carbonyl stretching band of the acetyl groups as discussed previously<sup>7)</sup>. The  $1710\text{ cm}^{-1}$  band is considered to arise mainly from the 4-C carbonyl group and the  $1685\text{ cm}^{-1}$  band from the 2-C carbonyl stretching vibrations coupled with the N-H bending mode, since the 2-C carbonyl bond, being conjugated with the C=N bonds, should have lower frequency vibration than the 4-C carbonyl bond.

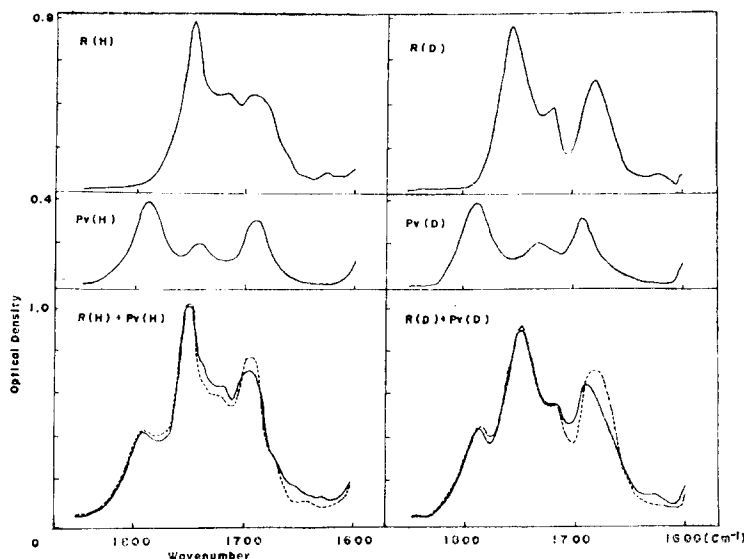


Fig. 2 — Infrared spectra of the carbonyl stretching region of riboflavin tetraacetate (R) and penicillin-V (Pv). The concentration of each solute is 0.004 M and path length is 0.2 mm.

H; nondeuterated compound, D; N-deuterated compound.  
Solid line; observed spectra. Dotted line: calculated sum of the upper two spectra.

The spectrum of penicillin-V shows three medium bands at  $1800$ ,  $1740$  and  $1710\text{ cm}^{-1}$  which were assigned to the carbonyl stretching vibration band and N-H bending. The  $1800\text{ cm}^{-1}$  band which appears at such high frequency region must be the carbonyl stretching band of 7-C in penicillin-V. The  $1745\text{ cm}^{-1}$  band is considered to arise from non-bonded N-H vibration because on the deuteration of the imino group the band moves to  $1735\text{ cm}^{-1}$  and the  $1690\text{ cm}^{-1}$  band is assignable to the carbonyl stretching band of free carboxylic acid of 2'-C.

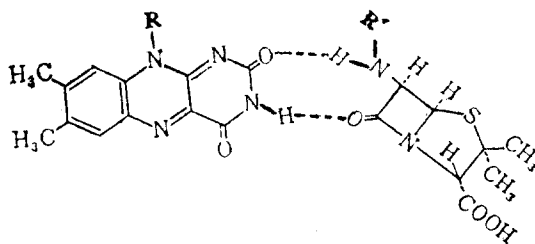
When both the solutions are mixed together, new association bands appear at 1730 and 1650  $\text{cm}^{-1}$  and the band at 1690  $\text{cm}^{-1}$  become weak. The 1745  $\text{cm}^{-1}$  band remains unchanged. The 1710 and 1690  $\text{cm}^{-1}$  bands of the coupled vibration of the carbonyl stretching and the N-H bending motions of riboflavin tetraacetate shift to 1700 and 1665  $\text{cm}^{-1}$  respectively by the formation of hydrogen bonds.

The spectrum of penicillin-V in the 1800-1600  $\text{cm}^{-1}$  region shows the carbonyl stretching band at 1800  $\text{cm}^{-1}$ , which becomes weak and shifts to the lower frequency when riboflavin is added to the solution (Figure 2).

It is clear that a hydrogen bond is formed at the carbonyl group of 7-C. On the other hand in the N-deuterated compounds the bands at 1745 and 1713  $\text{cm}^{-1}$  of the mixture are unaffected. Only the 1675 and 1800  $\text{cm}^{-1}$  bands of carbonyl groups decrease in intensity and new band appears 1710  $\text{cm}^{-1}$ . Since only the 1675  $\text{cm}^{-1}$  band of N-deuterated compound shifts to lower frequency by the addition of penicillin-V, the 2-C carbonyl group of riboflavin tetraacetate seems to be used for the formation of hydrogen bond which penicillin-V as previous condition<sup>8)</sup>.

Besides this, it is doubtless that the imino group of the riboflavin tetraacetate and the imino group of penicillin-V are employed for the complex formation. This fact indicates the importance of the cyclic dimer formation for the association of riboflavin tetraacetate and penicillin-V.

From the present results, however, the following structure can be proposed for the association.



It is generally believed that there is some kind of interaction between isoalloxazine ring and adenine moieties of the FAD molecule<sup>11-15)</sup> as a co-enzyme of flavoenzyme.

Antibiotics, especially penicillins or chlortetracycline, are known to be inhibitors in the flavoenzyme where FAD serves as a co-enzyme<sup>16, 17)</sup>

It is suggested, therefore, that penicillin-V inhibit the flavoenzyme by disrupting the structure of FAD; it takes the flavin moiety from the intramolecular adenine and isoalloxazine complex.

**Fluorescence Spectra** — Fluorescence spectra of  $1 \times 10^{-4}M$  chloroform solution of riboflavin tetraacetate are given in figure 3.

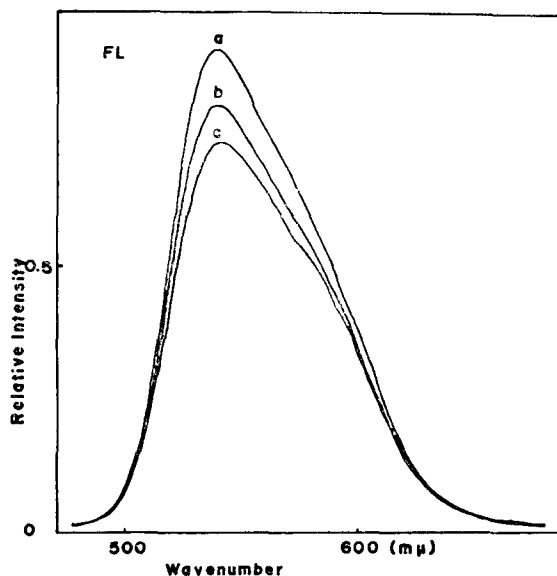


Fig. 3 — Fluorescence spectra of riboflavin tetraacetate in chloroform solution.

a ; R ( $1.2 \times 10^{-4}M$ )    b ; R ( $1.2 \times 10^{-4}M$ ) and Pv ( $1.2 \times 10^{-3}M$ )  
 c ; R ( $1.2 \times 10^{-4}M$ ) and Pv ( $1.2 \times 10^{-2}M$ )

When the same amount of penicillin-V was added to the solution, there was no change in the fluorescence spectrum of riboflavin tetraacetate. As the concentration of penicillin-V was increased, however, decisive quenching was detected. 15 percent quenching was observed for the solution containing  $1 \times 10^{-4}M$  riboflavin tetraacetate and  $1 \times 10^{-2}M$  penicillin-V.

The quenching appears to be related to the formation of the complex with penicillin-V through hydrogen bonds as discussed previously<sup>8)</sup>.

In this paper the number of the complex molecules at a given concentration did not considered.

**Ultraviolet and Visible Spectrum** — The spectrum of riboflavin tetraacetate and mixture of penicillin-V in chloroform are given in figure 4.

Spectrum in the same region were recorded for all of the mixture solutions that were

examined by the fluorescence measurements.

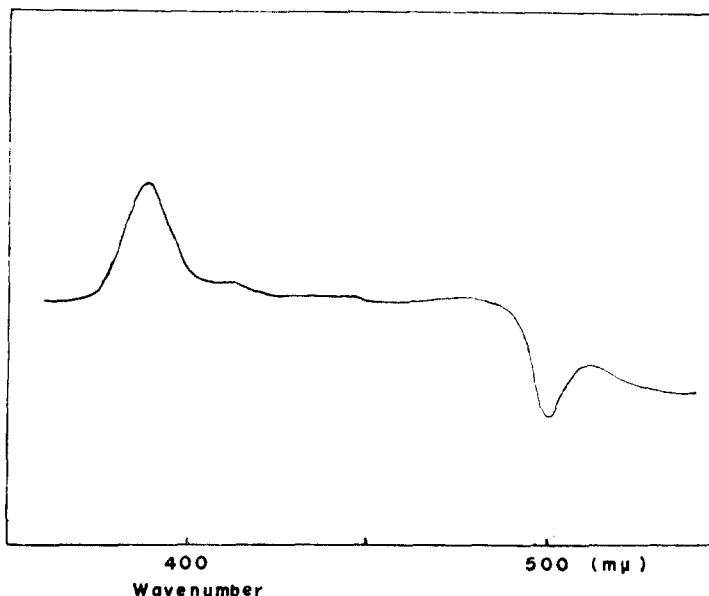


Fig. 4 — Differential UV spectrum of riboflavin tetraacetate ( $1.2 \times 10^{-4}M$ ) and mixture with penicillin-V ( $1.2 \times 10^{-8}M$ ) in chloroform solution.

The absorption at  $320 m\mu$  of the penicillin-V solution at  $1 \times 10^{-2}M$  is so strong that the spectrum of the mixture solution could not be recorded below  $320 m\mu$ . The shift and hypochromicity were appeared to be related to the formation of the complex with penicillin-V through hydrogen bonds, because the spectrum of riboflavin tetraacetate show red shift in  $490 m\mu$  region and blue shift in  $380 m\mu$  region when it is mixed together with penicillin-V.

The same phenomena were recognized from differential UV spectrum of mixture of  $5 \times 10^{-2}M$  penicillin-V.

**Acknowledgment** — The author expresses his sincere thank to Miss. E.J. Park for her experimental assistance. Thank is also due to Mr. S.U. Kim for the permission to use fluorescence spectrometer.

This work was partly supported by a grant from the Ministry of Education, Republic of Korea (1974).

## REFERENCES

1. H. M. Rauen and W. Rulf, *Arzneim-Forsch.*, **15**, 297 (1967)
2. Y. Kabaya, *Bitamin*(Kyoto), **20**, 130 (1959)
3. K. Hirano, *Bitamin* (Kyoto), **21**, 117 (1960); *ibid.*, 125; *ibid.*, 129; *ibid.*, 134.
4. N. B. Guarrant and J. M. Steel, *Proc. Soc. Exptl. Biol. Med.*, **98**, 542 (1958)
5. T. Tsukahara, *Japan. J. Microbiol.*, **5**, 41 (1961)
6. K. Yagi, *Biochemistry of Flavin*. Nankoto, Tokyo, 1956, p-265.
7. K. Yagi, Y. Yamamoto and M. Kobayashi, *J. Vitaminol.*(Kyoto), **14**, 271 (1968)
8. Y. Kyogoku and B. S. Yu, *Bull. Chem. Soc. of Japan*, **42**, 1387 (1969)
9. B. S. Yu and Y. Kyogoku, *ibid.*, **43**, 239 (1969)
10. Y. Kyogoku and B. S. Yu, *Chem-Biol. Interaction*, **2**, 117 (1970)
11. H. Weil-Marherbs, *Biochem. J.*, **40**, 363 (1946)
12. G. Weber, *ibid.*, **47**, 114 (1950)
13. L. G. Whitby, *ibid.*, **54**, 437 (1953)
14. J. C. M. Tsibris, D. B. McCormick and L. D. Wright, *Biochemistry*, **4**, 504 (1965)
15. G. Weber, "Flavins and Flavoproteins" ed. by E. Slater, Elsevier, Amsteldam, 1966, p-15
16. E. Takano, K. Yagi and Y. Okuda, 75th annual meeting of Japanese Pharmaceutical Society (1955)
17. R. Kuhn and P. Boulanger, *Z. Physiol. Chem.*, **241**, 233 (1936)