

Identification of *trans*-Cinnamic Acid from *Scrophularia Oldhami* Oliver

By

Lin Keun Woo*, Choung Sang Suh** and Kuk Hyun Shin*

Identification of *trans*-cinnamic acid and *p*-hydroxycinnamic acid for estimation of the sequence of *p*-methoxycinnamic acid.

(Received Dec. 30, 1968)

禹麟根* 徐廷祥** 申國鉉* : 玄蔘成分 *trans*-Cinnamic Acid 의 同定

玄蔘의 成分 *p*-methoxycinnamic acid 의 生合成 機構을 檢討하는 豫備試驗에서 *trans*-cinnamic acid 의 出現을 同定하였다.

Previous studies^{1,2)} on roots of *Scrophularia Oldhami* Oliver in this laboratory, have resulted in the isolation of *p*-methoxycinnamic acid, and its serum level has been discussed. In this paper, an attempt was made to identify the occurring of *trans*-cinnamic acid in the plant, since the component is implied as an intermediate in the formation of *p*-methoxycinnamic acid.

It has been suggested that methyl *p*-coumarate might be regarded as an intermediate in the metabolism of methyl *p*-methoxycinnamate by *Lentinus lespideus* and possibly in the biosynthesis of it³⁾. It was shown that anethol was synthesized from phenylalanine through phenylpyruvic acid, cinnamic acid, and 4-hydroxycinnamic acid by *Foeniculum vulgare*⁴⁻⁶⁾. Also, an acetone precipitate from an extract of spinach⁷⁾ and crude homogenates of pea seedlings or microsomes from the homogenates⁸⁾ catalyzed the hydroxylation of cinnamic acid to *p*-coumaric acid.

In view of those results, it might be supposed the presence of *trans*-cinnamic acid together with *p*-methoxycinnamic acid as an intermediate. Therefore, the distribution of *trans*-cinnamic acid, *p*-hydroxycinnamic acid and *p*-methoxycinnamic acid resolved into a study of the identification of them for further study using tracer.

EXPERIMENTAL

Isolation of *p*-methoxycinnamic acid.—It was conducted in the same procedure as described in a previous paper¹⁾.

Isolation of *trans*-cinnamic acid—500 g. of fresh roots of *Scrophularia Oldhami* Oliver were

* Natural Products Research Institute, Seoul National University, Seoul, Korea

** Department of Chemistry, Chon Puk University, Chon Joo, Korea.

extracted with 2 L. of 95% ethanol on 60° water bath for 3 hr. and the filtrate was concentrated under reduced pressure at 60° until a syrupy residue was left. It was then extracted with 300 ml. of ether and evaporated to a volume of 50 ml. and shaken with 10% of aqueous NaHCO_3 solution. The aqueous layer was acidified with NHCl and resulting precipitate was crystallized from 95% ethanol to give 0.34 g. of *p*-methoxycinnamic acid.

The filtrate freed from *p*-methoxycinnamic acid was extract with ether and the ethereal layer concentrated. From the residue, *p*-methoxycinnamic acid was removed by recrystallizing with water. The aqueous layer was shaken with ether and the ethereal solution was evaporated to dryness. This final residue was crystallized from 70% ethanol to give 0.12 g. of colorless crystals, m.p. 128°. Repeated recrystallization from 70% ethanol gave m.p. 133° undepressed upon admixture with an authentic sample, $\nu^{\text{KBr}}_{\text{Max}}$ 2550, 1690, 1615, 968 and 702 cm^{-1} .

Paper chromatography.—The filtrate freed from *p*-methoxycinnamic acid was shaken with ether and the organic layer was evaporated and then dissolved in 95% ethanol to be applied for chromatography.

An ascending paper chromatographic procedure was carried out on Whatman No. I paper at room temperature. The solvent system was $n\text{-BuOH}$: 3% NH_4OH (1 : 1) and detecting reagents were 0.4% brom phenol blue(A) and diazotized sulfanilic acid-10% NaHCO_3 . (B)

The R_f values of *p*-methoxycinnamic acid, *p*-hydroxycinnamic acid, and *trans*-cinnamic acid were identical with those of an authentic sample. (See Table I).

TABLE I. R_f values of components

Component detected	R_f		Color by Reagent	
	Reference	Sample	(A)	(B)
<i>p</i> -Hydroxycinnamic acid	0.18	0.18	Blue	Red
<i>p</i> -Methoxycinnamic acid	0.39	0.39	Blue	—
<i>trans</i> -Cinnamic acid	0.45	0.45	Blue	—
Unidentified	—	0.09	Blue	Orange

REFERENCES

- (1) W.S. Woo, *J. Pharm. Soc. Korea*, 7, 55(1963).
- (2) W.S. Woo *J. Pharm. Sci.*, 57, 27 (1968).
- (3) H. Shimazono and F.F. Nord, *Arch. Biochem. Biophys.* 78, 263 (1958).
- (4) K. Kaneko, *Chem. Pharm. Bull.*, 8, 875(1960).
- (5) *Ibid.*, 9, 108 (1961).
- (6) *Ibid.*, 9, 108 (1961).
- (7) C.G. Mead, *Proc. Natl. Acad. Sci.*, 52, 1482(1964).
- (8) W. David and E.C. Eric, *Arch. Biochem. Biophys.*, 122, 252, 256 (1961).