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Induction of Heme oxygenase-1 and Overproduction of Carbon Monoxide Protect Cells from Apoptosis Caused by Peroxynitrite

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Stimulated macrophages undergo oxidative burst and oxidative stress by overproducing O2 through activation of NADPH-oxidase, a heme-containing membrane enzyme. Cells with minimal oxidative stress can survive from the toxicity of O₂ both by utilizing the SOD and GSH-peroxidase pathway. While severe oxidative stress can kill the cell, moderately stressed cells can survive by eliminating the O2 via induction of iNOS and overproduction of NO to convert the overporoduced O₂ into peroxynitrite (ONOO), a strong oxidizing product that is detoxified by intracellular GSH. However, when the production of ONOO is excessive, cellular GSH is depleted and cells undergo apoptotic cell death. In response to the GSH depletion and to survive from the ONOO induced cell death, cells upregulate the expression of heme oxygenase-1 (HO-1). HO is the rate-limiting enzyme degrading heme into bilivedin (bilirubin) and carbon monoxide (CO). Induction of HO-1 and elevation of HO activity will accelerate the heme degradation and prevent Fenton reaction, limiting the generation of hydroxyl radical (HO*) and additional expression of heme-requiring enzymes like the NADPH-oxidase and iNOS that produce O₂ and NO, respectively. Additionally, bile pigment antioxidants (biliverdin and bilirubin) produced by the enhanced HO activity will detoxify ONOO and the CO also produced by the HO activity will inhibit additional productions of O₂ and NO by binding to the heme contained in NADPH-oxidase and iNOS, respectively. Such multi-level cross talks between the enzymes that produce O₂, NO and CO will protect the oxidatively stressed aerobic cells from the ONOO-derived cell death. Thus, the cells with upregulated HO-1 overproducing CO were insensitive to the stimulation by LPS or peroxynitrite. Alternatively, when the cells were pre-exposed to increasing doses of CO generated from CO-releasing molecule, the LPS-derived overproductions of O2 (oxidative burst) and NO (iNOS induction) were inhibited in a dosedependent manner. This caused cells to undergo a smaller degree of oxidative stress, allowing better survival from a given dose of LPS with smaller inductions of iNOS and HO-1. Furthermore, the cells overexpressing HO-1 or exposed to exogenous CO-donor were resistant to the ONOOinducible apoptotic cell death. This CO-dependent inhibiton of ONOO derived apoptotic cell death was causatively associated with prevention or reversal of ONOO-dependent depolarization of mitochondrial transmembrane potential ($\Delta \psi_{\rm m}$).