

P 99

Cryopreservation of Embryogenic Callus in Sweetpotato

Jong-Suk Park, Suk-Weon Kim¹, Dong-Su In², Jong-Seon Eun^{3*}

Mokpo Experiment Station, NHAES, RDA, Muan, Jeollanam-do 534-833

¹Genetic Resources Center and ²Eugentech Inc., KRIBB, Oun-dong 52, Yuseong-gu, Daejeon, 305-333

³Department of Horticulture, Chonbuk National University, Jeonju 561-756, Korea

Objectives

Sweetpotato is an important food crop in the subtropics and tropics mainly propagated vegetatively and conserved by filed gene banks. These method are not desirable for long-term storage. Embryogenic callus might provide alternative system for germ-plasm storage if cryopreservation of apical meristem are not readily available and it is also important because they are frequently used in genetic transformation protocol

Materials and Methods

1. Materials: Embryogenic callus derived from apical meristem of *Ipomoea batatas* L. cv. Yulmi
2. Methods
 - Pretreatment of 0, 1, 10 mg/L ABA in liquid MS medium containing 0.4 M sucrose and 1 mg/L 2,4-D
 - Cryoprotection solution of various concentration of glycerol at 0.54, 1.08, 1.62, 2.0 M, DMSO at 0.64, 1.28, 2.0 M only and

combination of 2.0 M DMSO + 0.54 M glycerol, 0.64 M DMSO + 2.0 M glycerol

Results and Discussion

Cryopreservation of embryogenic callus derived from apical meristem culture was attempted by slow prefreezing method (two-step method) with various cryoprotectants in sweetpotato cv. 'Yulmi'. Precultured embryogenic calli on medium containing 10 mg/L ABA prior to slow prefreezing in liquid nitrogen indicated higher survival rate than 1.0 mg/L ABA pretreatment. The cryoprotectant comprising 1.28 M DMSO in 0.4 M sucrose solution gave the best survival (over 46%) of sweetpotato cells exposed to liquid nitrogen as determined by TTC reduction and FDA staining method. Cryopreserved calli cultured on MS medium with 1.0 mg/L 2,4-D were grown for 4 weeks in the dark and induced embryos after another 4 weeks. They were subcultured on MS medium supplemented with 0.1 mg/L 2,4-D + 0.1 mg/L kinetin for 2 weeks and regenerated into normal plantlets in MS basal medium.

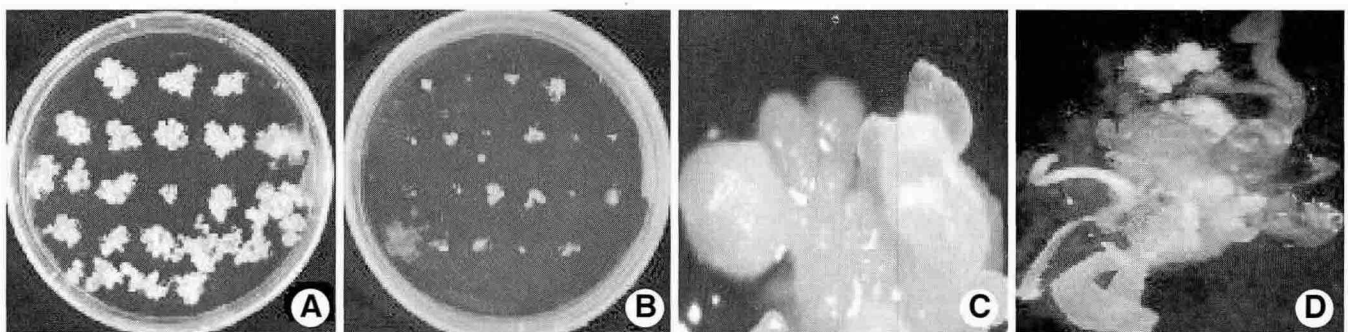


Figure 1. Cryopreservation of embryogenic callus. A, non-cryopreserved callus; B, cryopreserved callus by 1.28 M DMSO cultured on MS medium with 1.0 mg/L 2,4-D for 8 weeks; C, D, somatic embryos, and plantlets regenerated from cryopreserved embryogenic callus.