Highly incidence of Hepatocellular carcinoma in HBV X transgenic mice

Dae-Yeul Yu¹, Hyung-Bae Moon², Yong-Mahn Han¹, Chul-Ho Lee¹, Byung-Hwa Hyun¹, Seishi Murakami³ and Kyung-Kwang Lee¹

Chronic infection of hepatitis B virus (HBV) has been regarded as one of the major causative agents of hepatocellular carcinoma (HCC). Several mechanisms of oncogenic role of HBV have been proposed including gene activation by integration of the viral cis-elements, induction of recombination by HBV subgenome, imbalanced accumulation of viral proteins, and transactivation by viral proteins. Recently the X-gene product was found to be a viral transactivator (1). Since then, a variety of genes have been shown to be activated by the X-protein (2-5). Activating a broad spectrum of genes, the X-gene was implicated in carcinogenesis as one of the causative factors (6).

The X-gene is one of the four genes in the HBV genome (7). It encodes a polypeptide of 154 aa, the amino acid sequences of which are well conserved among mammalian hepadna viruses such as woodchuck hepatitis virus (WHV) and ground squirrel hepatitis virus (GSHV) (8). It is noteworthy that duck hepatitis B virus (DHBV) lacks the X-gene and no HCC developes in DHBV infection in contrast to frequent occurrence of HCC in WHV and less frequent but significant occurrence in GSHV (8). Most of the primary tumors and tumor-derived cell lines have some or all of the X-region and upstream pre-S/S sequences integrated into the host genome (9). In addition, most of the primary tumors produced X-region transcripts, whereas relatively few made transcripts from other regions of the genome (10,11). Many of these integrated fragments make hepatitis-B-virus-encoded X antigen (HBxAg) capable of trans-activation both in vitro (12,13) and in vivo (14) although the natural targets of HBxAg transactivation in liver diseases, including HCC, remain to be clearly identified. Altogether, these properties provide a strong basis on which to suspect the HBxAg as the most likely cause of HBV-associated hepatic cell transformation. evidence that HBxAg contributes to hepatocarcinogenesis was reported using a transgenic mouse system by one research group (15,16,17). Altered foci, adenoma, and carcinoma appeared in their HBx transgenic mice with persistently high levels of HBx expression. However, these observations have not been reproducible in other transgenic mouse systems using the X-gene (18-22). Therefore, the oncogenic role of HBx has been strongly

¹Korea Research Institute of Bioscience and Biotechnology, Taejon, 305-333, Korea

²Wonkwang University, Iksan, 570-749, Korea and

³Cancer Research Institute of Kanazawa University, 920, Japan

suspected, and the positive role of HBx in the hepatocarcinogenesis remains controversial.

Result and Discussion

We have created transgenic mice bearing HCC by expressing the X-gene to present a useful model for defining the molecular events that follow the expression of the viral X-gene and are responsible for the development of liver cancer. 279 young mice were born from the 946 embryos transferred, however only 3 transgenic founder mice were determined. Integration rate of the transgene on mice chromosomes was very low level comparing to the results of other authors (24) and many mice were also dead in early stage of the growth before examining expression of the X-gene. It might be caused due to the physiological problem by overexpression of the X-protein in liver of transgenic mice. Despite difficulty in getting the HBx transgenic mice, we luckily got one transgenic founder mouse (HEX-3 line) stably transmitting the transgene to its progenies. The expression level of the X-mRNA in tissues was a little different from that of Kim et al (15). It was reported that HBV X mRNA was highly expressed in the liver and testis and weakly in the kidney (15). However, the expression level in HEX-3 line mice was high in the testis and kidney, but very low in the liver. RT-PCR result showed the presence of the transcript in the heart as well. However, expression of the X-protein by immunohistochemical analysis was positive only in liver tissues of all the transgenic mice examined. We have not examined the reason why post-transcriptional expression did not occurr in kidney, testis, and heart of HBx transgenic One of the possibilities is the lack of some factors necessary for post transcriptional-regulation of the X-protein in these tissues (25). There were mild to moderate pleomorphic nuclei or enlarged nuclei and multiple small vacuolar cytoplasmic changes in all HBx transgenic mice from the age of 6 months to 18 months. In 3 of the 4 HBx transgenic mice which did not have grossly diagnosed hepatocellular carcinomas microscopically diagnosed, small hepatocellular carcinomas were found. Grossly diagnosed hepatocellular carcinomas were found in 5 of the 7 HBx transgenic mice after age of 11 months. Overall incidence of the hepatocellular carcinoma was 8 out of the 10 HBx transgenic mice (80%) and their age ranged from 6 to 18 months. The incidence of hepatic tumor may be comparable to that reported in CD-1 strain (15,16). Susceptibility to hepatocarcinogenesis varies in different mouse strains. Some strains, including the C57BL6J, C57BL10Sn, DBA/2J and Balb/c, have very low spontaneous liver tumor incidences (\langle 4\% of animals develop liver tumors), whereas others, such as the C3H/HeJ, have high incidences (40-50%) (26). As our transgenic mice were derived from F1 hybrid (C57BL/6 × DBA), the spontaneous incidence of hepatic tumors in these mice is expected to be lower than 4%. Therefore, 80% hepatic tumor incidence in our transgenic mice, generated by the expression of HBV X gene, may be very significant. Koike et al. (16) reported that low levels of HBx expression by Northern blot hybridization were insufficient for initiating tumor formation in mouse liver. E1 heterozygous line with relatively low levels of HBx expression developed Hepatic tumors in only 1 out of 16, which is comparable to the incidence of hepatic tumors reported in normal CD-1 strain (27,28). However, although the mRNA level expressed in the liver of our HBx transgenic mice was very low according to Northern blot analysis (Fig.3), the incidence of hepatic tumors was comparable to that in the H9 transgenic mouse line with high level expression of HBx as reported by Koike et al. (16). Therefore, our result suggests that continued expression of the X-gene as shown in Fig. 4A and 4B, even though at very low level, but at the appropriate sites of liver, may be responsible for the development of hepatic tumors at high incidence.

transgenic mice for the X-gene have been reported by several groups (3,15,19,20,21,22,31,29,30). Kim et al. (15) observed high incidence of HCC in transgenic mice by microinjecting a 1.15kb HBV subtype adr DNA fragment, which spans nucleotide positions 707 to 1856 in the viral genome, into single-cell embryos derived from outbred CD-1 mice. In contrast, other HBx transgenic lines generated in different mouse strains developed no obvious hepatic pathology, although they expressed the X-gene in liver cells and the HBx protein could be detected in some cases (3,19,20,21,22,23,30). However, in these cases the identity of the used for generating transgenic mice mouse strain might not have been an absolute criterion for the induction of hepatic tumors in the transgenic mice, because we used the same F1 hybrid mouse (C57BL/6×DBA) as that used by Purfumo et al. (21) and Terradillos et al. (23). One of the critical factors responsible for the difference in the outcomes of our experiments and perfumo et al. (21) could be the source of the X-gene used to generate the transgenic mice. We used the adr type of X-gene whereas Perfumo et al. used the ayw type; the nucleotide homology between these two gene types is only 39%. It could be that some elements responsible for inducing neoplasia in liver by the HBx might be deleted in the sequence of the X-gene of ayw type used in generation of HBx transgenic mice by Perfumo et al. (21).

Another possibility is that transgene integration site in the genome of HBx transgenic mice affected the incidence of hepatic tumor induction.

The HEX-3 transgenic mouse has been stably transmitting its transgenic trait to the progeny through generation until now. We are now trying to expand the HBx transgenic mice stock to present a useful model for defining the molecular events responsible for the development of liver cancer.

References

- 1. Twu JS, Schlomer RH. Transcriptional transactivating function of hepatitis B virus. J Virol 1987; 61: 3448-3453.
- 2. Twu JS, Chu K, Robinson WS. Hepatitis B virus X gene activates kB-like enhancer sequences in the long terminal repeat of immunodeficiency virus 1. Proc Natl Acad Sci

- USA 1989; 86: 5168-5172.
- 3. Balsano C, Avantaggiati ML, Natoli G, De Marzio E, Will H, Perricaudet M, Levrero M. Full-length and truncated versions of the hepatitis B virus X protein transactivate the c-myc protooncogene at the transcriptional level. Biochem Biophys Res Commun 1991;176: 985-992.
- 4. Twu JS, Lai MY, Chen DS, Robinson W. Activation of protooncogene c-jun by the X protein of hepatitis B virus. Virology 1993;192: 346-350.
- Colgrove R, Simon G, Ganem D. Transcriptional activation of homologous and heterologous genes by the hepatitis B virus X gene product in cells permissive for viral replication. J Virol 1989; 63: 4019-4026.
- 6. Wollersheim M, Debelka U, Hofschneider PH. A transactivating function encoded in the hepatitis B virus X gene is conserved in the integrated state. Oncogene 1988; 3: 545-552.
- 7. Koike K, Akatsuka T, Miyamura T. Characterization of hepatitis B virus X gene: In vitro translation of mRNA from COS-1 cells transfected with the X gene. Virology 1988;163: 233-235.
- 8. Koike K. Hepatitis B virus HBx gene and hepatocarcinogenesis. Intervirology 1995; 38: 134-142.
- 9. Matsubara K, Tokino T. Integration of hepatitis B virus DNA and its implications for hepatocarcinogenesis. Mol Biol Med 1990; 7: 243-260.
- 10. Diamantis ID, McGandy CE, Chen TJ, Liaw YF, Gudat F, Bianchi L. Hepatitis B X gene expression in hepatocellular carcinoma. J Hepatol 1992; 15: 400-403.
- 11. Paterlini P, Poussin K, Kew M, Franco D, Brechot C. Selective accumulation of the X transcript of hepatitis B virus in patients negative for hepatitis surface antigen with hepatocellular carcinoma. Hepatology 1995; 21: 313-321.
- 12. Zahm P, Hofschneider PH, Koshy R. The HBV X-ORF encodes a trans-activator: a potential factor in viral hepatocarcinogenesis. Oncogene 1988; 3: 169-177.
- 13. Wollersheim M, Debelka U, Hofschneider PH. A trans-activating function encoded in the hepatitis B virus X gene is conserved in the integrated state. Oncogene 1998; 3: 545-552.
- Balsano C, Billet O, Benoun M, Cavard C, Zider A, Grimber G, Natoli G, Briand P, Levrero M. Hepatitis B virus X gene product acts as a transactivator in vivo. J Hepatol 1994; 21: 103-109.
- 15. Kim CM, Koike K, Saito I, Miyamura T, Jay G. Hepatitis B virus HBx gene induces liver cancer in transgenic mice. Nature 1991;351: 317-320.
- 16. Koike K, Moriya K, Iino S, Yotsuyanagi H, Endo Y, Miyamura T, Kurokawa K. High-level expression of hepatitis B virus HBx gene and hepatocarcinogenesis in transgenic mice. Hepatology 1994; 19(4): 810-819.
- 17. Ueda H, Ullrich SJ, Gangemi JD, Kappel CA, Ngo L, Feitelson MA, Jay G. Functional inactivation but not structural mutation of p53 causes liver cancer. Nat Genet 1995; 9: 41-47.
- 18. Balsano C, Billet O, Benoun M, Carvard C, Zider A, Grimber G, Natoli G, Briand P, Levreno M. The hepatitis B virus X gene product transactivates the HIV-LTR in vivo.

- Arch Virol 1993; 8: 63-71.
- Billet O, Grimber G, Levrero M, Seye KA, Briand P, Joulin V. In vivo activity of the hepatitis B virus core promoter: tissue specificity and temporal regulation. J Virol 1995;69: 5912-5916.
- 20. Lee T-H, Finegold MJ, Shen RF, DeMayo JL, Woo SLC, Butel JS. Hepatitis B virus transactivator X protein is not tumorigenic in transgenic mice. J Virol 1990;64: 5939-5947.
- 21. Perfumo S, Amicone L, Colloca S, Giorgio M, Pozzi L, Tripodi M. Recognition efficiency of the hepatitis B virus polyadenylation signal is tissue specific in transgenic mice. J virol 1992;66: 6819-6823.
- 22. Yen TSB. Hepadnaviral X gene: Review of recent progress. J Biomed Sci 1996;3: 20-30.
- 23. Terradillos O, Billet O, Renard C-A, Levy R, Molina T, Briand P, Buendia MA. The hepatitis B virus X gene potentiates c-myc-induced liver oncogenesis in transgenic mice. Oncogene 1997; 14: 395-404.
- 24. Wall RJ, Bolt DJ, Frels WI, Hawk HW, King D, Pursel VG, Rexroad CE, Rohan Jr. and R.M. Transgenic farm animals: current state of the art. Agrobiotech News and Information 1990; 2(3): 391-395.
- 25. Huang, ZM, Yen TBS. Role of hepatitis B virus posttranscriptional regulatory element in intronless transcripts. Mol Cell Biol 1994; 15: 3864-3869.
- 26. Drinkwater NR. Genetic control of hepatocarcinogenesis in inbred mice. In Colburn,N,H. (ed.), Genes and Signal transduction in multistage carcinogenesis. Dekker Inc., New York, 1989: 3-17.
- 27. Homburger F, Russfield AB, Weisburger JH, Lim S, Chak SP, Weisburger EK. Aging changes in CD-1 HaM/ICR mice reared under standard laboratory conditions. J Natl Cancer Inst 1975; 55: 37-43.
- 28. Percy DH, Jonas AM. Incidence of spontaneous tumors in CD-1 HaM/ICR mice. J Natl Cancer Inst 1971; 46: 1045-1053.
- 29. Slagle BL, Lee TH, Medina D, Finegold MJ, Butel JS. Increased sensitivity to the hepatocarcinogen diethylnitrosamine in transgenic mice carrying the hepatitis B virus X gene. Mol Carcinog 1996; 15: 261-269.
- 30. Reifenberg K, Lohler J, Pudollek HP, Schmitteckert E, Spindler G, Kock J, Schlicht HJ. Long-term expression of the hepatitis B virus core-e-and X-proteins does not cause pathologic changes in transgenic mice. J Hepatol 1997; 26(1): 119-130.