

NMR Studies on the Structure of Human Annexin I

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Annexin I is a member of the annexin family of calcium dependent phospholipid binding proteins and has anti-inflammatory activity by inhibiting phospholipase A₂ (PLA₂). Recent X-ray crystallographic study of annexin I identified six Ca²⁺ binding sites, which was different types (type II, III) from the well-known EF-hand motif (type I). In this work, the structure of annexin I was studied at atomic level by using ¹H, ¹⁵N and ¹³C NMR (nuclear magnetic resonance) spectroscopy, and the effect of Ca²⁺ binding on the structure of annexin I was studied, and compared with that of Mg²⁺ binding. When Ca²⁺ was added to annexin I, NMR peak change was occurred in high- and low-field regions of ¹H-NMR spectra. NMR peak change by Ca²⁺ binding was different from that by Mg²⁺ binding. Because annexin I is a larger protein with 35 kDa molecular weight, site-specific (amide-¹⁵N, carbonyl-¹³C) labeling technique was also used. We were able to detect methionine, tyrosine and phenylalanine peaks respectively in ¹³C-NMR spectra, and each residue was able to be assigned by the method of doubly labeling annexin I with [¹³C]carbonyl-amino acid and [¹⁵N]amide-amino acid. In ¹³C-NMR spectra of [¹³C]carbonyl-Met labeled annexin I, we observed that methionine residues spatially located near Ca²⁺ binding sites were significantly effected by Ca²⁺ binding. From UV spectroscopic data on the effect of Ca²⁺ binding, we knew that Ca²⁺ binding sites of annexin I have cooperativity in Ca²⁺ binding. The interaction of annexin I with PLA₂ also could be detected by using heteronuclear NMR spectroscopy. Consequently, we expect that the anti-inflammatory action mechanism of annexin I may be a specific protein-protein interaction. The residues involved in the interaction with PLA₂ can be identified as active site by assigning NMR peaks effected by PLA₂ binding.