Manual Liquid Based Cytology for Pap Smear Preparation and HPV Detection by PCR in Pakistan

Shehla Akbar1, Shgufta Nasir Pervez2, Walayat Shah1*

Abstract

This study was conducted on female patients with different gynecological problems attending the gynecology out-patient departments of two tertiary care hospitals in Peshawar city of Khyber Pakhtunkhwa, Pakistan between August 2012 and October 2013. The 200 patients had an age range of 21-65 years. Smears were taken with cervical brushes and preserved in preservative medium and processed for manual liquid based cytology (MLBC) for Pap staining. Out of 200 collected samples, 30 samples were found inadequate on cytology. Of the remaining 170 samples, 164 (96.47%) were normal, 5 (2.94%) were of atypical squamous cells of unknown significance (ASCUS) and 1 (0.6%) was of high grade squamous intraepithelial lesion (HSIL). On PCR all the samples were positive for beta globin gene fragment including those reported inadequate on cytology. Out of the 5 ASCUS samples, 2 samples were positive for HPV, one each for HPV 16 and HPV 18, and the rest of the 3 samples were negative for HPV DNA. The 1 sample of HSIL was positive for HPV 16 on PCR. Out of 164 normal samples on cytology, only 1 sample was HPV 16 positive. So overall, 4 (2%) out of 200 samples were positive for HPV DNA, where 3 were HPV 16 (1.5%), and 1 was HPV 18 (0.5%) positive, and thus the ratio of infection with of HPV 16 to HPV 18 was 3:1 in the general population. In conclusion, PCR based HPV detection is a more sensitive method for screening of HPV infection than cytology as sample inadequacy does not affect the results. However, it can be combined with cytology methods in a HPV positive female to achieve the maximum results.

Keywords: Cervical cancer - HPV - Papanicolaou stain - manual liquid based cytology - Pakistan

Introduction

Cervical cancer is the third most commonly occurring cancer (Mathers et al., 2010) among women Worldwide and is one of the major public health encumbrances. Human papillomavirus (HPV) is the main culprit (zur Hausen, 1999) in the causation of cervical cancer. It is anticipated that at some point in their lives, around 79% of women, globally, get HPV infection (Koutsky, 1997). The rates of attaining an HPV infection peak soon after the age when most young women become sexually active (Dunne and Markowitz, 2006). Unprotected sexual contact is the main route of transmission (Garnett et al., 2002) with a high probability of transmission estimated to be 50-80%. The first step which is essential for cervical cancer to develop is infection of the cervix by one of the high risk types of HPV carrying oncogenic potential (Schiffmann, 2007).

The facts gathered so far about cervical cancer shows that this can be controlled through primary as well as secondary preventive measures. Primary prevention relies on the general awareness of women about risk factors and requisite changes in life style which is a main question in the developing countries where educational level of the women is yet to progress considerably (Nandini, 2012). Therefore, in these countries the center of attention is secondary prevention through early cytologic detection and conventional Pap smear cytology happens to be the foundation of it. However, its widespread use has not been possible in these countries (Nandini, 2012) due to shortage of resources, technical staff and other amenities. Also its sensitivity is said to reduce to less than 50% when there is presence of obscuring blood, inflammatory cells or broad areas of overlapping epithelial cells, even though the rate of invasive cervical cancer is decreased through this method of screening. Around 30% cases (ACOG Committee, 2009) reported negative on Pap cytology test recently done turned out to have cervical cancer. Liquid based cytology has recently become an alternative to conventional pap staining (CPS) (Nandini, 2012) in the detection of intraepithelial lesions as well as in invasive carcinoma of the uterine cervix. Manual Liquid Based Cytology (MLBC) is a technique that enables the cells to be suspended in a monolayer (Kavatkar et
Materials and Methods

A total of 200 female patients with gynecological problems were enrolled consecutively in the study. A specifically designed proforma was filled from patients prior to sample collection. Written consents were obtained from the patients and the concerned department and ethical committee. Sample collection was done between August 2012 and October 2013.

Married women of age 21 years to 65 years, who came to the gynecologic OPD for reasons like abnormal uterine bleeding, abnormal vaginal discharge, pelvic or lower abdominal pain, dyspareunia, post-coital bleeding, irregular menstrual cycles, were included in the study.

Married women below 21years and above 65 years, those having menstrual bleeding at the time of sample collection, those with total abdominal hysterectomy, visible polyps which could bleed readily, drug history of oral contraceptives, radiotherapy, use of vaginal creams, disinfectants and lubricants within the previous 24 hours, severe genital atrophy, pregnancy and women who had a pap test with in the previous three months were excluded from the study.

Cervical scrapings were obtained from the cervix near the squamo-columnar junction with a cytobrush and the tips were snapped-off into a 50ml sterile plastic bottle containing (Martinez-Contreras et al., 2008) 14mls PBSEDTA. Once detached from the cytobrush and suspended in the PBSEDTA medium (137 mM sodium chloride (NaCl), 2.7 mM potassium chloride (KCl), 10 mM disodium phosphate (Na2HPO4), 2 mM di-potassium hydrogen phosphate (KH2HPO2), 25 mM disodium EDTA, (pH 7.4), each sample was fixed by addition of 2 ml 96% ethanol. The collected samples placed in a bottle of preservative, were sealed, labeled and the specimen taken to the pathology laboratory at the Institute of Basic Medical Sciences (IBMS) in solution, stored and processed for cytological examination and DNA extraction.

In the laboratory, cell suspensions were mixed with the help of a vortexer and 11ml of it was transferred into a 15 ml round bottom falcon tube.1.5ml of alcoholic agar solution (Rooste et al., 2001) was added to it. Centrifugation of the falcon tubes was done for 15 minutes at 2000rpm. Cervical cells were trapped in a 1ml volume of spontaneously produced agar-gel at the bottom of the test tube. The overlying solution was discarded by pouring. Vortex-mixing the agar-gel portion at the bottom caused a rapid shift from gel to solution form, resulting in a viscous cell suspension. With the help of an adjustable jester, 200μl of the cell suspension was taken up and applied drop wise to glass slides, and in this way cytology slides were made. The slides were air-dried and stained using Papanicolaou stains, which were then interpreted by a histopathologist for cytology.

DNA extraction

From the remaining cell suspension, 250μl was taken for DNA extraction. DNA was extracted using Genomic DNA mini kit (invitrogen by life technologies, USA) following strictly the manufacturer’s instructions. The extracted DNA was run on 1% agarose gel to confirm that enough DNA has been extracted and to assess the quality of the genomic DNA extracted.

The integrity of extracted DNA was checked by PCR using primer set against a segment of β-globin gene used as an internal-control. The samples were then subjected to a generic screen for the presence of HPVVDNA using GP5+/6+ primers which target the conserved sequences.
in the HPV L1 region and can detect a number of low and high risk mucosal HPVs. Next type specific primers were used to detect the presence of HPV-16 and HPV-18 DNA (the most commonly found HR-HPVs in cervical cancer) in the samples. All the primers were selected from previous published work (Romero-Pastrana, 2012) and shown in Table 1.

PCR was carried out in a 25 µl reaction volume containing 2 µl genomic DNA, 1 µl (0.5 µl each) of the forward and reverse primers, 2.5 µl of PCR buffer, 1 µl of MgCl2, 1µl of dNTP mix, 1 µl of Taq DNA polymerase and 16.5 µl of sterile PCR water. All the above products are manufactured by Life technologies, USA, under the label of invitrogen. PCR was carried out in MultiGeneoptimax by Labnet International. The Cycling conditions were kept as follows: 5 min of denaturation at 94°C, followed by 35 cycles of 30 seconds of denaturation at 94°C, 30 seconds of annealing, and 30 seconds of extension at 72°C. Before the reactions were cooled to room temperature, an additional incubation for 10 minutes at 72°C was performed. Annealing temperatures for all four primer pairs were adjusted at optimum temperatures.

5 µl of the PCR product was run on 2% agarose gel along with a 100bp molecular weight marker (Invitrogen 100bp DNA Ladder, Life technologies, USA), to estimate the band size of the amplicon. Gel images were analyzed and saved using a major Science Gel Documentation System.

### Results

#### Cytology findings

Pap-stained MLBC specimens were screened by histopathologist, according to the Bethesda system 2001 (Moriarty et al., 2002) into normal cells (normal), atypical squamous cells of unknown significance (ASCUS), and cells with low- and high grade intraepithelial lesions (LSILs and HSILs, respectively) and squamous cell carcinoma. The cytological diagnosis was made without knowledge of the PCR results for HPV-DNA. Among 200 clinical samples, 30 samples were reported inadequate either due to absence of endocervical cells, obscured by blood or mucus, less cellularity or some staining problems, 164 were normal out of the rest of the 170 samples which makes around 96.47%, 5 samples were reported to have ASCUS (2.94%), no samples were reported to have LSIL (0%) and 1 sample was reported have HSIL (0.6%).

#### PCR findings

All the samples were found positive for β-globin primers, including those reported inadequate on cytology, showing the integrity of the DNA extracted from the cervical cell suspension left over after slide preparations for Pap smear (Figure 1).

Only 4 out of 200 samples (2%) were found positive for HPV DNA with HPV general primers (Figure 2). On

### Table 1. Sequences of Primers Used in the Study

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Primer</th>
<th>Primer sequence 5'-3'</th>
<th>Primer Length (No. of bases)</th>
<th>Tm °C</th>
<th>Amplicon size (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HPV 16</td>
<td>F</td>
<td>Ttaggcacactgaccaacca</td>
<td>22</td>
<td>68</td>
<td>207</td>
</tr>
<tr>
<td>HPV 16</td>
<td>R</td>
<td>Taatccgtcctttgtgagct</td>
<td>22</td>
<td>64</td>
<td></td>
</tr>
<tr>
<td>HPV 18</td>
<td>F</td>
<td>Tccgtgtggtcatccacgagc</td>
<td>22</td>
<td>72</td>
<td>274</td>
</tr>
<tr>
<td>HPV 18</td>
<td>R</td>
<td>Caactgtgcatcattggac</td>
<td>22</td>
<td>66</td>
<td></td>
</tr>
<tr>
<td>β-gl</td>
<td>F</td>
<td>Gaagagccaagcaggttgac</td>
<td>20</td>
<td>62</td>
<td>268</td>
</tr>
<tr>
<td>β-gl</td>
<td>R</td>
<td>Caactgtgcatcattggac</td>
<td>20</td>
<td>60</td>
<td></td>
</tr>
<tr>
<td>GP</td>
<td>5+</td>
<td>Ttgtagctgtaggtgagattc</td>
<td>23</td>
<td>62</td>
<td>~145</td>
</tr>
<tr>
<td>GP</td>
<td>6+</td>
<td>Gaaaataaactgtaaatcatc</td>
<td>25</td>
<td>60</td>
<td></td>
</tr>
</tbody>
</table>

*F, forward primer; R, reverse primer; Tm, melting temperature; GP, general primer; β-gl, beta globin*.

![Figure 1. Representative Pictures of PCR Product with Beta Globin Primers.](image)

From left to right- position 1 shows 100bp DNA-ladder for comparison of bands, position 2 is kept empty and position 3 and onwards are bands of beta-globin gene segment which is around 268bp size

![Figure 2. PCR Results for HPV General Primers.](image)

Position 1 on the left in each of the upper and lower panel is from known HPV-positive sample used as control for PCR results validation and band size confirmation. The last position on the right in the lower panel is negative control. Two bands in the center of the upper and 2 bands on the lower panel are of the samples found positive on PCR with general primers
type specific PCR 3 (1.5%) samples were found positive for HPV 16 (Figure 3), and 1 (0.5%) sample for HPV 18, making HPV 16 to 18 infection ratio as 3:1 in the general population sampled for this study.

We further compared the cytology findings with PCR results and it was found that out of 5 samples reported to have ASCUS, 1 sample was positive for HPV 18 DNA and 1 for HPV 16 DNA and the rest of the 3 were negative for both HPV general primers as well as type specific primers. The 1 sample reported as HSIL on cytology was HPV 16 positive on PCR. Out of the rest of the normal samples on cytology, only 1 sample was found HPV 16 positive on PCR. The ages of all these HPV positive patients were between 35-45 years except the one patient with HSIL who was a 60 year old lady.

Discussion

Cervical cancer holds second position in the list of most frequently occurring cancers amongst women throughout the world and as compared to the developed countries, most of it occurs in developing countries (Ferlay et al., 2005; GLOBOCAN, 2012) for which the main reason is shown to be the failure either to begin or then continue successful cervical-cancer screening programs (Wright and Kuhn, 2012). As for screening tests, cytology in association with strong healthcare facilities, has considerably aided in the prevention of cervical cancer in countries that have enough resources to set up and maintain well-conducted programs (Wright and Kuhn, 2012).

The mortality rate of cervical cancer in Pakistan is higher than the incidence rate as compared to various western countries. This elevated mortality rate is assigned to the delayed presentation of cases to the medical facility in Pakistan (Badar et al., 2007) where above 70% of cancer cases come to seek medical help when the disease has moved to a higher stage (Bhurgri, 2004).

From the facts gathered so far from the very few research projects on human papillomavirus and cervical cancer in Peshawar (Khattak et al., 2006), it seems that still the exact statistical data on the incidence or prevalence of the disease in this population is not known. Health care facilities happen to be insufficiently supplied and the existing ones are inclined to focus more on the curative aspect, rather than on the preventive aspect of diseases and health problems. Women are seldom informed about prevention and advancements such as screening for early detection of cervical cancer; consequently, there is no demand from the female population and no genuine political motivation to set up screening programs. Another major reason seems to be extensive poverty.

Regarding the prevalence of HPV, there are barely any collected figures on the viral presence in the non-cancerous specimens from the general female population of Peshawar. Substantiate research that would support in beginning an immunization program against HPV in this kind of population is undoubtedly deficient. From our results it can be seen that HPV do occur in this part of the country, and among the general population the most prevalent type of HPV is 16 and HPV 18. HPV was most frequent in age group 35-45 years old. The most disappointing fact is that the general population including the very highly educated women does not have the knowledge about cervical cancer and its etiological factor plus the health professionals are least interested in educating them about such issues. The gynecologist here relies only on cytology for all kind of patients and if it is negative, the patient is considered negative. From our results we have seen that cytology is prone to false-negative results and for such women who come to see a doctor only when they become symptomatic, this is a dangerous situation for if the women is actually positive for HPV, it cannot be said that the next time when she visits a doctor what stage of the disease she might be in.

Sherman et al. (2002) suggested that about 15% of women in annual screening programs who concurrently have a negative Pap test and a positive oncogenic HPV test will have a subsequent abnormal Pap test within 5 years. All these problems could be solved by initiating a proper screening program throughout the country. From our results we have conclude that PCR-based tests are the best for all these cases if conducted at a very basic health facility by a trained medical staff and only the positive cases should be referred to a tertiary care set up to be aided by cytology and further keeping in mind that most of the HPV infections in women below 30 years of age are cleared spontaneously so in this way giving the patient some time to clear the infection as well as closely following her for it through repeat cytology/colposcopy. In addition most of the samples giving ASCUS on cytology results belong to women in the age group above 30 years.
further explaining the need to start screening for this age group. Although many studies supported the use of HPV DNA testing for primary screening largely due to a higher sensitivity, the data nevertheless, suggested that the specificity and positive predictive values might be lower when compared with cytology-based assays (Junyangdikul et al., 2013; Lui, 2013; Priebe, 2013; Siriannkugl et al., 2014).

In different studies carried out at different institutions of Pakistan, the most frequent explanation for a low Pap test coverage is either lack of knowledge regarding it, or regarding its usefulness, instead of economic limitations or accessibility (Bokutz, 2008). We also noted that patient’s information level regarding cervical cancer was very low and literally not a single patient knew about cervical cancer etiology and immunization for its prevention or the mode of transmission of the causative agent. Improving awareness of the general population on the subject of cervical cancer and its screening is a significant step in improving the coverage of Pap test amongst the women of Peshawar because any harmonized cancer screening attempts are doubtful to be successful without any well-grounded knowledge amongst the targeted population concerning its aims and effectiveness.

References


